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(54) Title: EXPRESSION PROFILE OF PROSTATE CANCER

(57) Abstract: The present invention relates to compositions and methods for cancer diagnostics, including but not limited to, cancer markers. In particular, the present invention provides gene expression profiles associated with prostate cancers. Genes identified as cancer markers using the methods of the present invention find use in the diagnosis and characterization of prostate cancer. In addition, the genes provide targets for cancer drug screens and therapeutic applications.

EXPRESSION PROFILE OF PROSTATE CANCER

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FIELD OF THE INVENTION

The present invention relates to compositions and methods for cancer diagnostics, including but not limited to, cancer markers. In particular, the present invention provides gene expression profiles associated with prostate cancers. The present invention further provides novel markers useful for the diagnosis, characterization, and treatment of prostate cancers.

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BACKGROUND OF THE INVENTION

Afflicting one out of nine men over age 65, prostate cancer (PCA) is a leading cause of male cancer-related death, second only to lung cancer (Abate-Shen and Shen, Genes Dev 14:2410 [2000]; Ruijter et al., Endocr Rev, 20:22 [1999]). The American Cancer Society estimates that about 184,500 American men will be diagnosed with prostate cancer and 39,200 will die in 2001.

Prostate cancer is typically diagnosed with a digital rectal exam and/or prostate specific antigen (PSA) screening. An elevated serum PSA level can indicate the presence of PCA. PSA is used as a marker for prostate cancer because it is secreted only by prostate cells. A healthy prostate will produce a stable amount -- typically below 4 nanograms per milliliter, or a PSA reading of "4" or less -- whereas cancer cells produce escalating amounts that correspond with the severity of the cancer. A level between 4 and 10 may raise a doctor's suspicion that a patient has prostate cancer, while amounts above 50 may show that the tumor has spread elsewhere in the body.

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When PSA or digital tests indicate a strong likelihood that cancer is present, a transrectal ultrasound (TRUS) is used to map the prostate and show any suspicious areas.

Biopsies of various sectors of the prostate are used to determine if prostate cancer is present. Treatment options depend on the stage of the cancer. Men with a 10-year life expectancy or less who have a low Gleason number and whose tumor has not spread beyond the prostate are often treated with watchful waiting (no treatment). Treatment options for more aggressive cancers include surgical treatments such as radical prostatectomy (RP), in which the prostate is completely removed (with or without nerve sparing techniques) and radiation, applied through an external beam that directs the dose to the prostate from outside the body or via low-dose radioactive seeds that are implanted within the prostate to kill cancer cells locally. Anti-androgen hormone therapy is also used, alone or in conjunction with surgery or radiation. Hormone therapy uses luteinizing hormone-releasing hormones (LH-RH) analogs, which block the pituitary from producing hormones that stimulate testosterone production. Patients must have injections of LH-RH analogs for the rest of their lives.

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While surgical and hormonal treatments are often effective for localized PCA, advanced disease remains essentially incurable. Androgen ablation is the most common therapy for advanced PCA, leading to massive apoptosis of androgen-dependent malignant cells and temporary tumor regression. In most cases, however, the tumor reemerges with a vengeance and can proliferate independent of androgen signals.

The advent of prostate specific antigen (PSA) screening has led to earlier detection of PCA and significantly reduced PCA-associated fatalities. However, the impact of PSA screening on cancer-specific mortality is still unknown pending the results of prospective randomized screening studies (Etzioni et al., J. Natl. Cancer Inst., 91:1033 [1999]; Maattanen et al., Br. J. Cancer 79:1210 [1999]; Schroder et al., J. Natl. Cancer Inst., 90:1817 [1998]). A major limitation of the serum PSA test is a lack of prostate cancer sensitivity and specificity especially in the intermediate range of PSA detection (4-10 ng/ml). Elevated serum PSA levels are often detected in patients with non-malignant conditions such as benign prostatic hyperplasia (BPH) and prostatitis, and provide little information about the aggressiveness of the cancer detected. Coincident with increased serum PSA testing, there has been a dramatic increase in the number of prostate needle biopsies performed (Jacobsen et al., JAMA 274:1445 [1995]). This has resulted in a surge of equivocal prostate needle biopsies (Epstein and Potter J. Urol.,

166:402 [2001]). Thus, development of additional serum and tissue biomarkers to supplement PSA screening is needed.

SUMMARY OF THE INVENTION

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The present invention relates to compositions and methods for cancer diagnostics, including but not limited to, cancer markers. In particular, the present invention provides gene expression profiles associated with prostate cancers. The present invention further provides novel markers useful for the diagnosis, characterization, and treatment of prostate cancers.

In some embodiments, the present invention provides a method for characterizing prostate tissue in a subject, comprising: providing a prostate tissue sample from a subject; and detecting the presence or absence of expression of hepsin in the sample, thereby characterizing the prostate tissue sample. In some embodiments, detecting the presence of expression of hepsin comprises detecting the presence of hepsin mRNA. In other embodiments, detecting the presence of expression of hepsin mRNA comprises exposing the hepsin mRNA to a nucleic acid probe complementary to the hepsin mRNA. In yet other embodiments, detecting the presence of expression of hepsin comprises detecting the presence of a hepsin polypeptide. In some embodiments, detecting the presence of a hepsin polypeptide comprises exposing the hepsin polypeptide to an antibody specific to the hepsin polypeptide and detecting the binding of the antibody to the hepsin polypeptide. In some embodiments, the subject comprises a human subject. In some embodiments, the sample comprises tumor tissue. In some embodiments, the tumor tissue sample is a post-surgical tumor tissue sample and the method further comprises the step of c) identifying a risk of prostate specific antigen failure based on detecting the presence or absence of expression of hepsin. In some embodiments, characterizing prostate tissue comprises identifying a stage of prostate cancer in the tissue. In some embodiments, the stage includes but is not limited to, high-grade prostatic intraepithelial neoplasia, benign prostatic hyperplasia, prostate carcinoma, and metastatic prostate carcinoma. In some embodiments, the method further comprising the step of c) providing a prognosis to the subject. In some embodiments, the prognosis comprises a

risk of developing prostate specific antigen failure. In other embodiments, the prognosis comprises a risk of developing prostate cancer.

The present invention also provides a method for characterizing prostate tissue in a subject, comprising: providing a prostate tissue sample from a subject; and detecting the presence or absence of expression of pim-1 in the sample, thereby characterizing the prostate tissue sample. In some embodiments, detecting the presence of expression of pim-1 comprises detecting the presence of pim-1 mRNA. In other embodiments, detecting the presence of expression of pim-1 mRNA comprises exposing the pim-1 mRNA to a nucleic acid probe complementary to the pim-1 mRNA. In yet other embodiments, detecting the presence of expression of pim-1 comprises detecting the presence of a pim-1 polypeptide. In some embodiments, detecting the presence of a pim-1 polypeptide comprises exposing the pim-1 polypeptide to an antibody specific to the pim-1 polypeptide and detecting the binding of the antibody to the pim-1 polypeptide. In some embodiments, the subject comprises a human subject. In some embodiments, the sample comprises tumor tissue. In some embodiments, the tumor tissue sample is a postsurgical tumor tissue sample and the method further comprises the step of c) identifying a risk of prostate specific antigen failure based on detecting the presence or absence of expression of pim-1. In some embodiments, characterizing prostate tissue comprises identifying a stage of prostate cancer in the tissue. In some embodiments, the stage includes but is not limited to, high-grade prostatic intraepithelial neoplasia, benign prostatic hyperplasia, prostate carcinoma, and metastatic prostate carcinoma. In some embodiments, the method further comprising the step of c) providing a prognosis to the subject. In some embodiments, the prognosis comprises a risk of developing prostate specific antigen failure. In other embodiments, the prognosis comprises a risk of developing prostate cancer.

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The present invention further provides a method for characterizing prostate tissue in a subject, comprising: providing a prostate tissue sample; and detecting a decreased or increased expression relative to a non-cancerous prostate tissue control of two or more markers selected from the group consisting of HEPSIN, FKBP5, FASN, FOLH1, TNFSF10, PCM1, S100A11, IGFBP3, SLUG, GSTM3, IL1R2, ITGB4, CCND2, EDNRB, APP, THROMBOSPONDIN 1, ANNEXIN A1, EPHA1, NCK1, MAPK6,

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SGK, HEVIN, MEIS2, MYLK, FZD7, CAVEOLIN 2, TACC1, ARHB, PSG9, GSTM1, KERATIN 5, TIMP2, GELSOLIN, ITM2C, GSTM5, VINCULIN, FHL1, GSTP1, MEIS1, ETS2, PPP2CB, CATHEPSIN B, COL1A2, RIG, VIMENTIN, MOESIN, MCAM, FIBRONECTIN 1, NBL1, ANNEXIN A4, ANEXIN A11, IL1R1, IGFBP5, CYSTATIN C, COL15A1, ADAMTS1, SKI, EGR1, FOSB, CFLAR, JUN, YWHAB, NRAS, C7, SCYA2, ITGA1, LUMICAN, C1S, C4BPA, COL3A1, FAT, MMECD10, CLUSTERIN, PLA2G2A, thereby characterizing the prostate tissue sample. In some embodiments, the detecting comprises detecting three or more markers. In other embodiments, the detecting comprises detecting five or more markers. In still further embodiments, the detecting comprises detecting ten or more markers.

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The present invention additionally provides a method for characterizing prostate cancer in a subject, comprising: providing a tumor sample from a subject diagnosed with prostate cancer; and detecting decreased expression relative to a non-cancerous prostate tissue control of two or more cancer markers selected from the group consisting of IGFBP5, MADH4, NBL1, SEPP1, RAB2, FAT, PP1CB, MPDZ, PRKCL2, ATF2, RAB5A, and Cathepsin H, wherein decreased expression is diagnostic of metastatic prostate cancer. In some embodiments, the detecting comprises detecting three or more markers. In other embodiments, the detecting comprises detecting five or more markers. In still further embodiments, the detecting comprises detecting ten or more markers.

The present invention further provides a method for characterizing prostate cancer in a subject, comprising providing a tumor sample from a subject diagnosed with prostate cancer; and detecting increased expression relative to a non-cancerous prostate tissue of two or more cancer markers selected from the group consisting of CTBP1, MAP3K10, TBXA2R, MTA1, RAP2, TRAP1, TFCP2, E2-EPF, UBCH10, TASTIN, EZH2, FLS353, MYBL2, LIMK1, TRAF4, wherein increased expression is diagnostic of metastatic prostate cancer. In some embodiments, the detecting comprises detecting three or more markers. In other embodiments, the detecting comprises detecting five or more markers. In still further embodiments, the detecting comprises detecting ten or more markers.

In some embodiments, the present invention provides a kit for characterizing prostate cancer in a subject, comprising: a reagent capable of specifically detecting the presence of absence of expression of hepsin; and instructions for using the kit for

characterizing cancer in the subject. In some embodiments, the reagent comprises a nucleic acid probe complementary to a hepsin mRNA. In other embodiments, the reagent comprises an antibody that specifically binds to a hepsin polypeptide. In some embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

In other embodiments, the present invention provides a kit for characterizing prostate cancer in a subject, comprising: a reagent capable of specifically detecting the presence of absence of expression of pim-1; and instructions for using the kit for characterizing cancer in the subject. In some embodiments, the reagent comprises a nucleic acid probe complementary to a pim-1 mRNA. In other embodiments, the reagent comprises an antibody that specifically binds to a pim-1 polypeptide. In some embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

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In still further embodiments, the present invention provides a kit for characterizing prostate cancer in a subject, comprising: two or more reagents capable of 15 specifically detecting expression levels of two or more markers selected from the group consisting of FKBP5, FASN, FOLH1, TNFSF10, PCM1, S100A11, IGFBP3, SLUG, GSTM3, ATF2, RAB5A, IL1R2, ITGB4, CCND2, EDNRB, APP, THROMBOSPONDIN 1, ANNEXIN A1, EPHA1, NCK1, MAPK6, SGK, HEVIN, MEIS2, MYLK, FZD7, CAVEOLIN 2, TACC1, ARHB, PSG9, GSTM1, KERATIN 5, 20 TIMP2, GELSOLIN, ITM2C, GSTM5, VINCULIN, FHL1, GSTP1, MEIS1, ETS2, PPP2CB, CATHEPSIN B, CATHEPSIN H, COL1A2, RIG, VIMENTIN, MOESIN, MCAM, FIBRONECTIN 1, NBL1, ANNEXIN A4, ANEXIN A11, IL1R1, IGFBP5, CYSTATIN C, COL15A1, ADAMTS1, SKI, EGR1, FOSB, CFLAR, JUN, YWHAB, NRAS, C7, SCYA2, ITGA1, LUMICAN, C1S, C4BPA, COL3A1, FAT, MMECD10, 25 . CLUSTERIN, PLA2G2A, MADh4, SEPP1, RAB2, PP1CB, MPDZ, PRKCL2, CTBP1, CTBP2, MAP3K10, TBXA2F, MTA1, RAP2, TRAP1, TFCP2, E2EPF, UBCH10, TASTIN, EZH2, FLS353, MYBL2, LIMK1, GP73, VAV2, TOP2A, ASNS, CTBP, AMACR, ABCC5 (MDR5), and TRAF4; and instructions for using the kit for characterizing cancer in the subject. In some embodiments, the kit comprises reagents 30 capable of specifically detecting expression levels of three or more of the markers. In

other embodiments, the kit comprises reagents capable of specifically detecting expression levels of five or more of the markers. In still further embodiments, the kit comprises reagents capable of specifically detecting expression levels of ten or more of the markers. In some embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

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In yet other embodiments, the present invention provides a kit for characterizing prostate cancer in a subject, comprising: two or more reagents capable of specifically detecting decreased expression levels of two or more markers selected from the group consisting of IGFBP5, MADH4, NBL1, SEPP1, RAB2, FAT, PP1CB, MPDZ, PRKCL2, ATF2, RAB5A, and Cathepsin H; and instructions for using the kit for characterizing cancer in the subject. In some embodiments, the kit comprises reagents capable of specifically detecting decreased expression levels of three or more of the markers. In other embodiments, the kit comprises reagents capable of specifically detecting decreased expression levels of five or more of the markers. In still further embodiments, the kit comprises reagents capable of specifically detecting decreased expression levels of ten or more of the markers. In some embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

In an additional embodiment, the present invention provides a kit for characterizing prostate cancer in a subject, comprising: two or more reagents capable of specifically detecting increased expression levels of two or more markers selected from the group consisting of CTBP1, MAP3K10, TBXA2R, MTA1, RAP2, TRAP1, TFCP2, E2-EPF, UBCH10, TASTIN, EZH2, FLS353, MYBL2, LIMK1, TRAF4; and instructions for using the kit for characterizing cancer in the subject. In some embodiments, the kit comprises reagents capable of specifically detecting decreased expression levels of three or more of the markers. In other embodiments, the kit comprises reagents capable of specifically detecting decreased expression levels of five or more of the markers. In still further embodiments, the kit comprises reagents capable of specifically detecting decreased expression levels of ten or more of the markers. In some embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

The present invention also provides a method of screening compounds, comprising: providing a prostate cell sample; and one or more test compounds; and contacting the prostate cell sample with the test compound; and detecting a change in hepsin expression in the prostate cell sample in the presence of the test compound relative to the absence of the test compound. In some embodiments, the detecting comprises detecting hepsin mRNA. In other embodiments, the detecting comprises detecting hepsin polypeptide. In some embodiments, the cell is *in vitro*. In other embodiments, the cell is *in vitro*. In some embodiments, the test compound comprises an antisense compound. In other embodiments, the test compound comprises a drug.

The present invention further provides a method of screening compounds, comprising: providing a prostate cell sample; and one or more test compounds; and contacting the prostate cell sample with the test compound; and detecting a change in pim-1 expression in the prostate cell sample in the presence of the test compound relative to the absence of the test compound. In some embodiments, the detecting comprises detecting pim-1 mRNA. In other embodiments, the detecting comprises detecting pim-1 polypeptide. In some embodiments, the cell is *in vitro*. In other embodiments, the cell is *in vitro*. In some embodiments, the test compound comprises an antisense compound. In other embodiments, the test compound comprises a drug.

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The present invention provides a prostate cancer expression profile map comprising gene expression level information for two or more markers selected from the group consisting of: FKBP5, FASN, FOLH1, TNFSF10, PCM1, S100A11, IGFBP3, SLUG, GSTM3, ATF2, RAB5A, IL1R2, ITGB4, CCND2, EDNRB, APP, THROMBOSPONDIN 1, ANNEXIN A1, EPHA1, NCK1, MAPK6, SGK, HEVIN, MEIS2, MYLK, FZD7, CAVEOLIN 2, TACC1, ARHB, PSG9, GSTM1, KERATIN 5, TIMP2, GELSOLIN, ITM2C, GSTM5, VINCULIN, FHL1, GSTP1, MEIS1, ETS2, PPP2CB, CATHEPSIN B, CATHEPSIN H, COL1A2, RIG, VIMENTIN, MOESIN, MCAM, FIBRONECTIN 1, NBL1, ANNEXIN A4, ANEXIN A11, IL1R1, IGFBP5, CYSTATIN C, COL15A1, ADAMTS1, SKI, EGR1, FOSB, CFLAR, JUN, YWHAB, NRAS, C7, SCYA2, ITGA1, LUMICAN, C1S, C4BPA, COL3A1, FAT, MMECD10, CLUSTERIN, PLA2G2A, MADh4, SEPP1, RAB2, PP1CB, MPDZ, PRKCL2, CTBP1, CTBP2, MAP3K10, TBXA2F, MTA1, RAP2, TRAP1, TFCP2, E2EPF, UBCH10,

TASTIN, EZH2, FLS353, MYBL2, LIMK1, GP73, VAV2, TOP2A, ASNS, CTBP, AMACR, ABCC5 (MDR5), and TRAF4. In some embodiments, the map is digital information stored in computer memory. In some embodiments, the map comprises information for three or more markers. In other embodiments, the map comprises information for five or more markers. In still further embodiments, the map comprises information for ten or more markers.

The present invention also provides a prostate cancer expression profile map comprising gene expression level information for two or more markers selected from the group consisting of: IGFBP5, MADH4, NBL1, SEPP1, RAB2, FAT, PP1CB, MPDZ, PRKCL2, ATF2, RAB5A, and Cathepsin H. In some embodiments, the map is digital information stored in computer memory. In some embodiments, the map comprises information for three or more markers. In other embodiments, the map comprises information for five or more markers. In still further embodiments, the map comprises information for ten or more markers. In some embodiments, the prostate cancer is metastatic.

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The present invention further provides a prostate cancer expression profile map comprising gene expression level information for two or more markers selected from the group consisting of: CTBP1, MAP3K10, TBXA2R, MTA1, RAP2, TRAP1, TFCP2, E2-EPF, UBCH10, TASTIN, EZH2, FLS353, MYBL2, LIMK1, TRAF4. In some embodiments, the map is digital information stored in computer memory. In some embodiments, the map comprises information for three or more markers. In other embodiments, the map comprises information for five or more markers. In still further embodiments, the map comprises information for ten or more markers. In some embodiments, the prostate cancer is metastatic.

In some embodiments, the present invention provides a method for characterizing prostate tissue in a subject, comprising providing a prostate tissue sample from a subject; and detecting the presence or absence of expression of EZH2 in the sample, thereby characterizing the prostate tissue sample. In some embodiments, detecting the presence of expression of EZH2 comprises detecting the presence of EZH2 mRNA (e.g., including, but not limited to, by exposing the hepsin mRNA to a nucleic acid probe complementary to the hepsin mRNA). In other embodiments, detecting the presence of

expression of EZH2 comprises detecting the presence of a EZH2 polypeptide (e.g., including, but not limited to, by exposing the EZH2 polypeptide to an antibody specific to the EZH2 polypeptide and detecting the binding of the antibody to the EZH2 polypeptide). In some embodiments, the subject comprises a human subject. In some embodiments, the sample comprises tumor tissue. In some embodiments, characterizing the prostate tissue comprises identifying a stage of prostate cancer in the prostate tissue. In certain embodiments, the stage is selected from the group including, but not limited to, high-grade prostatic intraepithelial neoplasia, benign prostatic hyperplasia, prostate carcinoma, and metastatic prostate carcinoma. In some embodiments, the method further comprises the step of providing a prognosis to the subject (e.g., a risk of developing metastatic prostate cancer).

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In further embodiments, the present invention provides a kit for characterizing prostate cancer in a subject, comprising a reagent capable of specifically detecting the presence of absence of expression of EZH2; and instructions for using the kit for characterizing cancer in the subject. In some embodiments, the reagent comprises a nucleic acid probe complementary to a EZH2 mRNA. In other embodiments, the reagent comprises an antibody that specifically binds to a EZH2 polypeptide. In certain embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

In still other embodiments, the present invention provides a method of screening compounds, comprising providing a prostate cell sample; and one or more test compounds; and contacting the prostate cell sample with the test compound; and detecting a change in EZH2 expression in the prostate cell sample in the presence of the test compound relative to the absence of the test compound. In some embodiments, wherein the detecting comprises detecting EZH2 mRNA. In other embodiments, the detecting comprises detecting EZH2 polypeptide. In some embodiments, the cell is *in vitro*; while in other embodiments, the cell is *in vivo*. In some embodiments, the test compound comprises an antisense compound. In certain embodiments, the test compound comprises a drug.

In yet other embodiments, the present invention provides a method for characterizing inconclusive prostate biopsy tissue in a subject, comprising providing an

inconclusive prostate biopsy tissue sample from a subject; and detecting the presence of expression of AMACR in the sample, thereby characterizing the inconclusive prostate biopsy tissue sample. In some embodiments, detecting the presence of expression of AMACR comprises detecting the presence of AMACR mRNA (e.g., by exposing the AMACR mRNA to a nucleic acid probe complementary to at least a portion of the AMACR mRNA). In other embodiments, detecting the presence of expression of AMACR comprises detecting the presence of a AMACR polypeptide (e.g., by exposing the AMACR polypeptide to an antibody specific to the AMACR polypeptide and detecting the binding of the antibody to the AMACR polypeptide). In some embodiments, the subject comprises a human subject. In some embodiments, the presence of AMACR expression in the inconclusive biopsy tissue is indicative of prostate cancer in the subject. In certain embodiments, the method further comprises the step of detecting expression of a basal cell marker selected from the group consisting of 34βΕ12 and p63 and the absence of a basal cell marker expression and the presence of AMACR expression is indicative of prostate cancer in the subject.

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The present invention further provides a method of detecting AMACR expression in a bodily fluid, comprising providing a bodily fluid from a subject; and a reagent for detecting AMACR expression in the biological fluid; and contacting the bodily fluid with the reagent under conditions such that the reagent detects AMACR expression in the bodily fluid. In some embodiments, the bodily fluid is selected from the group consisting of serum, urine, whole blood, lymph fluid, and mucus. In certain embodiments, the presence of AMACR in the bodily fluid is indicative of cancer (e.g., prostate cancer).

The present invention additionally provides a kit for characterizing inconclusive prostate biopsy tissue in a subject, comprising a reagent capable of specifically detecting the presence or absence of expression of AMACR; and instructions for using the kit for characterizing inconclusive biopsy tissue in the subject. In some embodiments, the reagent comprises a nucleic acid probe complementary to at least a portion of an AMACR mRNA. In other embodiments, the reagent comprises an antibody that specifically binds to a AMACR polypeptide. In still other embodiments, the kit further comprises a second reagent, the second reagent capable of specifically detecting the expression of a basal cell marker selected from the group consisting of 34βΕ12 and p63.

In some embodiments, the instructions further comprise instructions for using the second reagent and the reagent for characterizing inconclusive biopsy tissue in the subject. In some embodiments, the instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

The present invention further provides a method of characterizing tissue in a subject, comprising providing a tissue sample from a subject, the tissue sample selected from the group consisting of breast tissue, ovarian tissue, lymph tissue, and melanoma tissue; and detecting the presence or absence of expression of AMACR in the sample, thereby characterizing the breast tissue sample.

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The present invention also provides a method of diagnosing cancer in a subject, comprising providing a tissue sample from a subject, the tissue sample selected from the group consisting of breast tissue, ovarian tissue, lymph tissue, and melanoma tissue; and wherein the subject is suspected of having cancer; and detecting the presence of expression of AMACR in the sample, thereby diagnosing cancer in the tissue sample of the subject.

The present invention provides a method of diagnosing cancer in a subject, comprising providing a blood sample from a subject suspected of having cancer; and detecting an immune response to AMACR in the blood sample, thereby diagnosing cancer in the subject. In some embodiments, the cancer is prostate cancer. In certain embodiments, detecting an immune response comprises detecting an antibody against the AMACR in the blood sample.

The present invention additionally provides a method of inhibiting the growth of cells, comprising providing a cell that expresses EZH2; and a reagent for inhibiting EZH2 expression in the cell; and contacting the cell with the reagent under conditions such that the expression of EZH2 in the cell is inhibited. In some embodiments, the reagent is an antisense oligonucleotide. In other embodiments, the reagent is a RNA duplex. In some embodiments, the reagent is a drug. In some embodiments, the cell is a prostate cancer cell. In some embodiments, the cell is in vitro; while in other embodiments, the cell is in vitro. In some embodiments, the contacting further results in a decrease in proliferation of the cell.

In certain embodiments, the present invention provides a method for characterizing prostate cancer in a subject, comprising providing a prostate tissue sample from a subject suspected of having prostate cancer; and detecting a decrease in expression of an annexin in the sample, thereby characterizing the prostate tissue sample. In some embodiments, the decrease in expression of an annexin is indicative of the presence of metastatic prostate cancer in the subject. In certain embodiments, the annexin is selected from the group including, but not limited to, Annexin 1, Annexin 2, Annexin 4, Annexin 6, Annexin 7, and Annexin 11.

In other embodiments, the present invention provides a method for characterizing prostate cancer in a subject, comprising providing a prostate tissue sample from a subject suspected of having prostate cancer; and detecting an increase in expression of a cterminal binding protein in the sample, thereby characterizing the prostate tissue sample. In some embodiments, the c-terminal binding protein is selected from the group consisting of c-terminal binding protein 1 and c-terminal binding protein 2. In certain embodiments, the increase in expression of a c-terminal binding protein is indicative of the presence of metastatic prostate cancer in the subject. In some embodiments, the expression of a c-terminal binding protein is indicative of an increased risk of PSA failure.

In other embodiments, the present invention provides a method for characterizing prostate cancer in a subject, comprising providing a prostate tissue sample from a subject suspected of having prostate cancer; and detecting an increase or decrease in expression of GP73, thereby characterizing the prostate tissue sample. In some embodiments, an increase in expression of gp73 is indicative of localized prostate cancer. In other embodiments, the prostate tissue sample is prostate cancer and a decrease in the expression of gp73 is indicative of metastatic prostate cancer.

DESCRIPTION OF THE FIGURES

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Figure 1 shows a gene expression profile of prostate cancer samples.

Figure 1a shows a dendrogram describing the relatedness of the samples. Figures

1b shows a cluster diagram of the samples groups compared against normal

adjacent prostate pool as a reference. Figures 1c shows a cluster diagram of the samples groups compared against commercial prostate pool reference.

Figure 2 shows functional clusters of genes differentially expressed in prostate cancer.

Figure 3 shows the expression of hepsin in prostate cancer samples as determined by Northern blot analysis and immunohistochemistry. Figure 3a shows Northern blot analysis of human hepsin (top) and normalization with GAPDH (bottom). NAT indicates normal adjacent prostate tissue and PCA indicates prostate cancer. Figure 3b shows tissue microarrays used for hepsin analysis. Figure 3c shows a histogram of hepsin protein expression by tissue type. Benign prostate hyperplasia (BPH). High-grade intraepithelial neoplasia (HG-PIN). Localized prostate cancer (PCA). Hormone-refractory prostate cancer (MET). Figure 3d shows Kaplan Meier Analysis.

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Figure 4 shows the expression of pim-1 in prostate cancer samples as determined by Northern blot analysis and immunohistochemistry. Figure 4a shows a histogram of pim-1 protein expression by tissue type as assessed from 810 tissue microarray elements. High-grade intraepithelial neoplasia (HG-PIN). Localized prostate cancer (PCA). Figure 4b shows a Kaplan-Meier analysis. The tope line represents patients with strong Pim-1 staining. The bottom line represents patients with absent/weak Pim-1 expression.

Figure 5 shows a comparison of gene expression profiles for normal adjacent prostate tissue and normal prostate tissue reference.

Figure 6 shows a focused cluster of prostate cancer related genes.

Figure 7 shows data for gene selection based on computed t-statistics for the NAP and CP pools.

Figure 8 shows an overview of genes differentially expressed in prostate cancer.

Figure 9 describes exemplary accession numbers and sequence ID Numbers for exemplary genes of the present invention.

Figure 10 provides exemplary sequences of some genes of the present invention.

Figure 11 an overview of the discovery and characterization of AMACR in prostate cancer utilized in some embodiments of the present invention.

Figure 12 describes a DNA microanalysis of AMACR expression in prostate cancer.

Figure 13 describes an analysis of AMACR transcript and protein levels in prostate cancer.

Figure 14 describes an analysis of AMACR protein expression using prostate cancer tissue microarrays.

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Figure 15 shows relative gene expression of AMACR in several samples.

Figure 16 shows AMACR protein expression PCA. Figure 19A shows AMACR protein expression in localized hormone naive PCA. Figure 19B shows strong AMACR expression in a naive lymph node metastasis. Error bars represent the 95% CI of the mean expression of the primary naive prostate cancer and corresponding lymph node metastases.

Figure 17 shows the hormonal effect on AMACR expression. Figure 17A shows PCA demonstrating strong hormonal effect due to anti-androgen treatment. Figure 17B shows Western Blot analysis representing the baseline AMACR expression in different prostate cell lines (Left) and Western Blot analysis of LNCaP cells for AMACR and PSA expression after treatment with an androgen or an anti-androgen for 24h and 48 hours (right).

Figure 18 shows AMACR over-expression in multiple tumors. AMACR protein expression was evaluated by immunohistochemistry on a multi-tumor and a breast cancer tissue microarray. Percentage of cases with positive staining (moderate and strong staining intensity) is summarized on the Y-axis. The left bar represents negative or weak staining and the right bar represents moderate or strong staining.

Figure 19 shows the results of laser capture microdissection (LCM) and RT-PCR amplification of AMACR in prostate cancer. LCM was used to isolate pure prostate cancer and benign glands and AMACR gene expression was characterized by RT-PCR in 2 radical prostatectomies. A constitutively expressed gene, GAPDH, was used as quantitative control of input mRNA. AMACR expression is barely detectable in benign glands, and is elevated in prostate cancer.

Figure 20 describes the identification and validation of EZH2 over-expression in metastatic prostate cancer. Figure 20a shows a cluster diagram depicting genes that molecularly distinguish metastatic prostate cancer (MET) from clinically localized prostate cancer (PCA). Figure 20b shows a DNA microarray analysis of prostate cancer

that reveals upregulation of EZH2 in metastatic prostate cancer. Figure 20c shows RT-PCR analysis of the EZH2 transcript in prostate tissue and cell lines. Figure 20d shows increased expression of EZH2 protein in prostate cancer.

Figure 21 shows that EZH2 protein levels correlate with the lethal progression and aggressiveness of prostate cancer. Figure 21a shows tissue microarray analysis of EZH2 expression. The mean EZH2 protein expression for the indicated prostate tissues is summarized using error bars with 95% confidence intervals. Figure 21b shows a Kaplan-Meier analysis demonstrating that patients with clinically localized prostate cancers that have high EZH2 expression (Moderate/Strong staining) have a greater risk for prostate cancer recurrence after prostatectomy (log rank test, p= 0.03).

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Figure 22 shows the role of EZH2 in prostate cell proliferation. Figure 22a shows an immunoblot analysis of RNA interference using siRNA duplexes targeting the EZH2 sequence in prostate cells. Figure 22b shows that RNA interference of EZH2 decreases cell proliferation as assessed by cell counting assay. Figure 22c shows that RNA interference of EZH2 inhibits cell proliferation as assessed by WST assay. Figure 22d shows that RNA interference of EZH2 induces G2/M arrest of prostate cells.

Figure 23 shows that EZH2 functions as a transcriptional repressor in prostate cells. Figure 23a shows a schematic diagram of EZH2 constructs used in transfection/transcriptome analysis. ER, modified ligand binding domain of estrogen receptor. H-1 and H-2, homology domains 1 and 2 which share similarity between EZH2 and E(z). CYS, cysteine-rich domain. SET, SET domain. TAG, myc-epitope tag. NLS, nuclear localization signal. Figure 23b shows confirmation of expression of EZH2 constructs used in a. An anti-myc antibody was used. Figure 23c shows a cluster diagram of genes that are significantly repressed by EZH2 overexpression. Figure 23d shows SAM analysis of gene expression profiles of EZH2 transfected cells compared against EZH2 .SET transfected cells. Figure 23e shows a model for potential functional interactions of EZH2 as elucidated by transcriptome analysis and placed in the context of previously reported interactions. +, induction. -, repression.

Figure 24 shows the detection of AMACR in PCA cell lines.

Figure 25 shows the detection of AMACR protein in serum by quantitation of microarray data.

Figure 26 shows an immunoblot analysis of serum from patients with either negative or positive PSA antigen.

Figure 27 shows an immunoblot analysis of the presence of AMACR in urine samples from patients with bladder cancer (females) or bladder cancer and increased PSA (males).

Figure 28 shows representative data of a humoral response by protein microarray analysis.

Figure 29 shows immunoblot analysis of the humoral response of AMACR.

Figure 29a shows an immunoblot analysis of the humoral response to AMACR. Figure 29b shows a control experiment where the humoral response was blocked.

Figure 30 shows GP73 Transcript levels in prostate cancer. Figure 30a shows the level of GP73 in individual samples after microarray analysis. Figure 30b shows the result of GP73 transcripts determined by DNA microarray analysis from 76 prostate samples grouped according to sample type and averaged.

Figure 31 shows that GP73 protein is upregulated in prostate cancer. Figure 31a shows Western blot analysis of GP73 protein in prostate cancer. Figure 31b shows an immunoblot analysis of the Golgi resident protein Golgin 97.

Figure 32 shows immunoblot analysis of normal and prostate cancer epithelial cells.

Figure 33 shows the cDNA expression of select annexin gene family members.

Figure 34 shows a heat map representation of annexin family gene expression across four prostate cancer profiling studies. Over and under expression at the transcript level are represented by shades of red and green, respectively. Gray shading indicates that insufficient data was available. Each square represents an individual tissue sample.

Figure 35 shows the expression of CtBP proteins in PCA specimens.

Figure 36 shows tissue microarray analysis of CtBP in prostate cancer that suggests mis-localization during prostate cancer progression.

Figure 37 shows the sub-cellular fractionation of LNCaP cells.

Figure 38 shows a Kaplan-Meier Analysis of prostate cancer tissue microarray

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GENERAL DESCRIPTION

Exploring the molecular circuitry that differentiates indolent PCA from aggressive PCA has the potential to lead to the discovery of prognostic markers and novel therapeutic targets. Insight into the mechanisms of prostate carcinogenesis is also gleaned by such a global molecular approach. Similar to breast cancer (Lopez-Otin and Diamandis, Endor. Rev., 19:365 [1998]), PCA develops in a complex milieu of genetic and environmental factors in which steroid hormone signaling plays a central role. The primary precursor lesion of PCA, high-grade prostatic intraepithelial neoplasia (HG-PIN), has several characteristics similar to other early invasive carcinomas (i.e., chromosomal abnormalities and cytologic features). Loss of specific chromosomal 10 regions (e.g., 8p21, 10q, 13q, 17p) along with losses and mutations of tumor suppressor genes such as Nkx3.1, PTEN, Rb, and p53 have been implicated in the initiation and progression of prostate cancer (Abate-Shen and Shen, supra). With the emergence of global profiling strategies, a systematic analysis of genes involved in PCA is now possible. DNA microarray technology is revolutionizing the way fundamental biological 15 questions are addressed in the post-genomic era. Rather than the traditional approach of focusing on one gene at a time, genomic-scale methodologies allow for a global perspective to be achieved. The power of this approach lies in its ability to comparatively analyze genome-wide patterns of mRNA expression (Brown and Botstein, Nat. Gent., 20 21:33 [1999]). Obtaining large-scale gene expression profiles of tumors allows for the identification of subsets of genes that function as prognostic disease markers or biologic predictors of therapeutic response (Emmert-Buck et al., Am. J. Pathol., 156:1109 [2000]). Golub et al. used DNA arrays in the molecular classification of acute leukemias (Golub et al., Science 286:531 [1999], demonstrating the feasibility of using microarrays for identifying new cancer classes (class discovery) and for assigning tumors to known 25 classes (class prediction). Using a similar approach, Alizadeh et al. showed that diffuse large B-cell lymphoma could be dissected into two prognostic categories by gene expression profiling (Alizadeh et al., Nature 403:503 [2000]). They provided evidence that lymphomas possessing a gene expression signature characteristic of germinal center B cells had a more favorable prognosis than those expressing genes characteristic of 30 activated peripheral B-cells. Similar large-scale classifications of breast cancer and

melanoma have been undertaken, and as with the other studies, molecular classification was the primary focus (Alizadeh et al., supra).

Accordingly, the present invention provides an analysis of gene expression profiles in benign and malignant prostate tissue. Three candidate genes, AMACR, hepsin and pim-1, identified by DNA microarray analysis of PCA, were characterized at the protein level using PCA tissue microarrays. Analysis of the differential gene expression profiles of normal and neoplastic prostate has led to the identification of a select set of genes that define a molecular signature for PCA. The expression profiling experiments of the present invention demonstrate a role for multiple, collaborative gene expression alterations which ultimately manifest as the neoplastic phenotype. By making direct comparative hybridizations of normal and neoplastic tissues, genes that molecularly distinguish benign tissue from malignant are identified.

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α-Methylacyl-CoA Racemase (AMACR) is an enzyme that plays an important role in bile acid biosynthesis and β-oxidation of branched-chain fatty acids (Ferdinandusse et al., J. Lipid Res., 41:1890 [2000]; Kotti et al., J. Biol.Chem., 275:20887 [2000]). Mutations of the AMACR gene have been shown to cause adultonset sensory motor neuropathy (Ferdinandusse et al., Nat. Genet., 24:188 [2000]). In diagnostically challenging prostate biopsy cases, pathologists often employ the basal cell markers 34βE12 or p63, which stain the basal cell layer of benign glands that is not present in malignant glands. Thus, in many biopsy specimens, the pathologist must rely on absence of staining to make the final diagnosis of prostate cancer. Experiments conducted during the development of the present invention identified AMACR as a marker expressed in cancerous biopsy tissue. Thus, the clinical utility of AMACR in prostate needle biopsies is large. For example, at the University of Michigan Medical Center, approximately 400 prostate needle biopsies are performed per year and approximately 20% require the use of a basal-cell specific marker to evaluate difficult lesions, characterized by a small amount of atypical glands. Accordingly, it is contemplated that in combination with basal cell specific markers, such as $34\beta E12$ or p63, screening for AMACR expression by the methods of the present invention results in fewer cases diagnosed as "atypical without a definitive diagnosis."

Identification of the over-expression of AMACR in prostate cancer has clinical utility beyond diagnostic uses. Experiments conducted during the development of the present invention revealed that the only non-cancerous tissue to expresses significant levels of AMACR protein is the human liver. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism in not necessary to practice the present invention. Nonetheless, it is contemplated that AMACR activity is required for prostate cancer growth and by virtue of its specificity serves as a therapeutic target.

Additional experiments conducted during the course of development of the present invention investigated AMACR expression in different groups of prostate cancer, including the aspect of neo-adjuvant hormonal withdrawal in localized disease. AMACR expression was found to be hormone independent in cell culture experiments. PSA, a gene known to be regulated by androgens, demonstrated hormone related alterations in expression under the same conditions. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that these findings provide evidence that AMACR is not regulated by the androgen pathway. It is further contemplated that the decreased AMACR expression in hormone refractory tissue allows the use of AMACR as a biomarker for hormone resistance. It is also contemplated that, given the fact that hormone treatment in the mean of hormonal withdrawal did not affect AMACR expression in the cell culture, that some other mechanism than the androgen pathway is responsible for AMACR downregulation in the integrity of cancer tissue.

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The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention.

Nonetheless, it is contemplated that, alternatively, AMACR is over expressed in the development of cancer, perhaps playing an important role in providing energy for the neoplastic cells. However, as the tumors become de-differentiated, they no longer require these sources of energy. It is contemplated that poorly differentiated tumors may take over other pathways to accomplish this same activity of branched fatty acid oxidation. There is no association with the proliferative rate of the tumor cells and AMACR expression.

AMACR expression was also examined in other cancers. Examination of other tumors demonstrated that colon cancer has the highest AMACR expression. As colorectal cancers are not known to be hormonally regulated, the fact that dedifferentiation and decreased AMACR expression were correlated in PCA further supports the hypothesis that de-differentiation leads to decreased AMACR expression in the hormone refractory metastatic PCA. Hormone treatment is also a front line therapy in metastatic prostate cancer but is known to loose efficacy, selecting out hormone insensitive clones. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that this phenomenon explains the observation that strong hormone treatment effect is consistent with decreased AMACR expression due to selection of potentially more de-differentiated cells.

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The AMACR gene product is an enzyme, which plays an important role in bile acid biosynthesis and beta-oxidation of branched-chain fatty acids (Kotti et al., J. Biol. Chem. 275:20887 [2000]; Ferdinandusse et al., J Lipid Res 42:137 [2001]). AMACR over expression occurs in tumors with a high percentage of lipids such as PCA and colorectal cancer. The relationship between fatty acid consumption and cancer is a controversial subject in the development of PCA and colorectal cancer (Moyad, Curr Opin Urol 11:457 [2001]; Willett, Oncologist 5:393 [2000]). An essential role for AMACR in the oxidation of bile acid intermediates has been demonstrated. AMACR encodes an enzyme which catalyzes the racemization of alpha-methyl branched carboxylic coenzyme A thioesters and is localized in peroxisomes and mitochondria (Schmitz et al., Eur J Biochem 231:815 [1995]). The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that, as AMACR is involved in the metabolism of lipids, that this leads to alterations in the oxidant balance of a cell. It is further contemplated that these changes are associated with DNA damage, malignant transformation, and other parameters of cell disturbance.

Additional experiments conducted during the course of development of the present invention demonstrated that AMACR mRNA and protein product are over expressed in a number of adenocarcinomas, including colorectal, prostate, breast, and

ovarian and melanoma. Adenocarcinoma from the colorectum and prostate demonstrated consistent AMACR over expression (92% and 83% of tumor, respectively). Thus, AMACR is of use in the diagnosis of colonic neoplasia. For example, in some embodiments of the present invention, AMACR is used in the diagnosis of dysplasia. Specifically, in the setting of inflammatory bowel disease (IBD), where the identification of dysplasia may be diagnostically challenging, one evaluates putative lesions for their AMACR protein expression intensity. In some embodiments, this is performed in conjunction with the analysis of the adenomatous polyposis coli gene, since mutations in this gene are also believed to occur early in the development of colorectal neoplasia (Kinzler and Vogelstein, Cell 87:159 [1996]; Tsao and Shibata, Am J Pathol 145: 531 [1994]).

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Colonic adenomas (Kinzler and Vogelstein, *supra*; Tsao and Shibata, *supra*) and high-grade PIN (McNeal and Bostwick, Hum Pathol 17:64 [1986]; McNeal *et al.*, Lancet 1:60 [1986]) are well know precursors of invasive colonic and prostate cancer, respectively. Experiments conducted during the course of development of the present invention demonstrated that AMACR is over expressed in colorectal adenomas (75%) and high-grade PIN (64%). Further supporting AMACR expression in early neoplastic lesions was the presence of focal AMACR expression in some atrophic prostate lesions. Some atrophic lesions (*i.e.*, proliferative inflammatory atrophy and postatrophic hyperplasia) have recently been recognized as proliferative in nature with molecular alterations suggestive of early neoplastic changes (De Marzo *et al.*, Am J Pathol 155:1985 [1999]; Shah *et al.*, Am J Pathol 158:1767 [2001]). Some morphologically benign prostate glands were also observed to have focal moderate AMACR staining. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that AMACR may have a role in the early steps of cancer development.

Several cancers that are associated with AMACR over expression, including colorectal, prostate and breast cancer, have been linked to high-fat diet. The exact mechanism how high-fat diet contributes to tumorigenesis in these organ systems is unknown, but emerging evidence suggest that peroxisome proliferator activated receptor (PPAR) mediated pathway plays a critical role (Debril *et al.*, J. Mol. Med. 79:30 [2001]).

Diet fatty acids have been shown to function as peroxisome proliferators and bind to and activate PPARs (Zomer et al., J. Lipid Res. 41:1801 [2000]), a family of nuclear receptor transcriptional factors. Activation of PPAR mediated pathways in turn control cell proliferation and differentiation. In addition, it can also alter the cellular oxidant balance (Yeldandi et al., Mutat. Res. 448:159 [2000]). The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that these effects act in concert to contribute to the tumorigenesis of several cancers. This hypothesis is supported by the findings that peroxisome proliferators, when given to mice, enhance the development colon adenomatous polyps in mice (Saez et al., Nat. Med. 4:1058 [1998]). In addition, PPARs are expressed in several prostate cancer cell lines and their ligands, and peroxisome proliferators, when added to culture, affect the growth of these cell lines (Shappell et al., Cancer Res. 61:497 [2001]; Mueller et al., PNAS 97:10990 [2000]). A phase II clinical trial also showed that troglitazone, a PPARy activator, could stabilize PSA level in patients with prostate cancer (Kubota et al., Cancer Res. 58:3344 [1998]; Hisatake et al., Cancer Res. 60:5494 [2000]).

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AMACR is an involved in the β -oxidation of pristanic acid (Ferdinandusse *et al.*, J. Lipid. Res. 41:1890 [2000]). Pristanic acid can function as a PPAR α activator and promote cell growth (Zomer *et al.*, J. Lipid Res. 41:1801 [2000]). The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that hyperfunctioning of β -oxidation pathway leads to exhaustion of reducing molecules and alters the cellular oxidant status (Yeldandi *et al.*, Mutat. Res. 448:159 [2000]).

The present invention further provides methods of targeting AMACR as a therapeutic target in cancer treatment. Over expressed in high percentage of colorectal, prostate, breast and melanoma, but not in adjacent normal tissues, AMACR is targeted using antibody or enzyme inhibitors. Toxicity is expected not to be a major concern because individuals with congenital absence of this enzyme have no or insignificant clinical manifestations (Clayton et al., Biochem. Soc. Trans. 29:298 [2001]).

Experiments conducted during the course of development of the present invention further demonstrated that AMACR is present in the serum of prostate cancer patients. In

addition, a humoral response to AMACR was identified based on the presence of antibodies to AMACR in the serum of prostate cancer patients.

Annexins are a group of structurally related calcium-binding proteins, which have a domain that binds to phospholipids and an amino terminal domain that determines specificity (Smith et al., Trends. Genet. 10:241 [1994]; Mailliard et al., J Biol. Chem. 271:719 [1996]). The annexins are involved in regulation of membrane trafficking, cellular adhesion and possible tumorigenesis. Experiments conducted during the course of development of the present invention used cDNA microarrays to study the expression patterns of multiple annexin family members in a wide range of prostate tissue samples in order to determine their role in PCA progression. Meta-analysis of gene expression data was employed to help further validate the cDNA expression array findings. Finally, high-density tissue microarrays were used to assess annexin protein expression levels by immunohistochemistry.

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Eight annexins were evaluated for their mRNA expression levels in benign prostatic tissue, localized hormone naïve PCA and metastatic hormone refractory PCA samples. Five annexins (1,2,4,7,and 11) demonstrated a progressive down regulation at the transcript level going from benign prostatic tissue to localized PCA to hormone refractory PCA. In order to validate the cDNA expression array finding of these 5 annexin family members, a meta-analysis was performed, which confirmed that when looking across 4 studies where at least two studies reported results, annexin 1,2,4, and 6 were significantly down regulated in localized PCA samples when compared to benign prostatic tissue. Therefore the meta-analysis confirmed results on annexin 1, 2, and 4. In these examples, summary statistics across all datasets found these annexins to be significantly down regulated at the cDNA level. However, not all of the 4 studies had significant down-regulation. Annexin 4, for example, was significantly down regulated in two of four studies but the resultant summary statistic, which also takes into account the number of samples evaluated, was statistically significant. Annexins 7,8, and 13 were not found to be significantly under expressed. As demonstrated in figure 1, annexin 7 does decrease significantly when comparing localized PCA and metastatic PCA.

The protein expression levels of all above five annexins tested were statistically significantly decreased in hormone refractory PCA samples when compared to either

localized PCA or benign prostate tissue. Four of 5 annexins also demonstrated a decrease in protein expression in clinically localized PCA as compared to benign prostate tissue. However, in none of these cases was the protein expression found to be significantly decreased. This second validation method at the protein level confirmed the cDNA expression array data for annexin 1,2,4, 7, and 11.

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Based on gene expression array data described herein, localized PCA cells down regulate their mRNA levels of annexins but maintained the corresponding protein expression levels. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that post-translational alteration may compensate for decrease mRNA, producing enough protein to maintain levels seen with benign samples. Since annexins play an important role in maintaining cellular adhesion, once the cells eventually lose this ability, tumor progression may occur. Therefore, as one might anticipate, annexin expression levels decreased significantly in the advanced hormone refractory PCA samples. This was confirmed at the protein level by significant decreases as demonstrated by immunohistochemistry.

A sequential down-regulation of annexins in both transcriptional and translational levels in metastatic PCA samples was observed. Annexin I, also called lipocortin, has been described as a phospholipase A2 inhibitor, and served as a substrate of epidermal growth factor receptor (Pepinsky et al., Nature 321:81 [1986]; Wallner et al., Nature 320:77 [1986]). The significant reduction of protein level has been shown in esophageal and prostate tumor cells (Paweletz et al., Cancer Res. 60:6293 [2000]). Annexin 2, also called p36, appears an efficient substrate of protein kinase C and Src pp60 (Hubaishy et al., Biochemistry 34:14527 [1995]). Annexin 4, called endonexin, regulates Cl- flux by mediating calmodulin kinase II (CaMKII) activity (Chan et al., J. Biol. Chem. 269:32464 [1994]). Annexin 7, synexin, is involved in Duchenne's muscular dystrophy (Selbert et al. Exp. Cell. Res. 222:199 [1996]). Its gene is located on human chromosome 10q21, and its protein expression was decreased in hormone refractory tumor cells. In conclusion, the results of experiments conducted during the course of development of the present invention suggest that down regulation of several annexin family members may play a role in the development of the lethal PCA phenotype.

Additional experiments conducted during the course of development of the present invention identified additional markers that exhibited altered (e.g., increased or decreased) expression in prostate cancer. Additional markers include, but are not limited to, EZH2, Annexins 1, 2, 4, 7, and 11, CTBP 1 and 2, GP73, ABCC5 (MDR5), ASNS, TOP2A, and Vav2. In particular, EZH2 was identified as a marker that was overexpressed in prostate cancer, and in particular, in metastatic prostate cancer. EZH2 was further identified as being correlated with clinical failure (e.g., increased PSA levels). In addition, siRNA inhibition of EZH2 resulted in a decrease in cell proliferation of a prostate cancer cell line.

The present invention thus identifies markers and targets for diagnostic and therapeutic agents in a variety of cancers.

DEFINITIONS

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To facilitate an understanding of the present invention, a number of terms and phrases are defined below:

The term "epitope" as used herein refers to that portion of an antigen that makes contact with a particular antibody.

When a protein or fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to a given region or three-dimensional structure on the protein; these regions or structures are referred to as "antigenic determinants". An antigenic determinant may compete with the intact antigen (i.e., the "immunogen" used to elicit the immune response) for binding to an antibody.

The terms "specific binding" or "specifically binding" when used in reference to the interaction of an antibody and a protein or peptide means that the interaction is dependent upon the presence of a particular structure (i.e., the antigenic determinant or epitope) on the protein; in other words the antibody is recognizing and binding to a specific protein structure rather than to proteins in general. For example, if an antibody is specific for epitope "A," the presence of a protein containing epitope A (or free, unlabelled A) in a reaction containing labeled "A" and the antibody will reduce the amount of labeled A bound to the antibody.

As used herein, the terms "non-specific binding" and "background binding" when used in reference to the interaction of an antibody and a protein or peptide refer to an interaction that is not dependent on the presence of a particular structure (i.e., the antibody is binding to proteins in general rather that a particular structure such as an epitope).

As used herein, the term "subject" refers to any animal (e.g., a mammal), including, but not limited to, humans, non-human primates, rodents, and the like, which is to be the recipient of a particular treatment. Typically, the terms "subject" and "patient" are used interchangeably herein in reference to a human subject.

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As used herein, the term "subject suspected of having cancer" refers to a subject that presents one or more symptoms indicative of a cancer (e.g., a noticeable lump or mass) or is being screened for a cancer (e.g., during a routine physical). A subject suspected of having cancer may also have one or more risk factors. A subject suspected of having cancer has generally not been tested for cancer. However, a "subject suspected of having cancer" encompasses an individual who has received an initial diagnosis (e.g., a CT scan showing a mass or increased PSA level) but for whom the stage of cancer is not known. The term further includes people who once had cancer (e.g., an individual in remission).

As used herein, the term "subject at risk for cancer" refers to a subject with one or more risk factors for developing a specific cancer. Risk factors include, but are not limited to, gender, age, genetic predisposition, environmental expose, previous incidents of cancer, preexisting non-cancer diseases, and lifestyle.

As used herein, the term "characterizing cancer in subject" refers to the identification of one or more properties of a cancer sample in a subject, including but not limited to, the presence of benign, pre-cancerous or cancerous tissue, the stage of the cancer, and the subject's prognosis. Cancers may be characterized by the identification of the expression of one or more cancer marker genes, including but not limited to, the cancer markers disclosed herein.

As used herein, the term "characterizing prostate tissue in a subject" refers to the identification of one or more properties of a prostate tissue sample (e.g., including but not limited to, the presence of cancerous tissue, the presence of pre-cancerous tissue that is

likely to become cancerous, and the presence of cancerous tissue that is likely to metastasize). In some embodiments, tissues are characterized by the identification of the expression of one or more cancer marker genes, including but not limited to, the cancer markers disclosed herein.

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As used herein, the term "cancer marker genes" refers to a gene whose expression level, alone or in combination with other genes, is correlated with cancer or prognosis of cancer. The correlation may relate to either an increased or decreased expression of the gene. For example, the expression of the gene may be indicative of cancer, or lack of expression of the gene may be correlated with poor prognosis in a cancer patient. Cancer marker expression may be characterized using any suitable method, including but not limited to, those described in illustrative Examples 1-15 below.

As used herein, the term "a reagent that specifically detects expression levels" refers to reagents used to detect the expression of one or more genes (e.g., including but not limited to, the cancer markers of the present invention). Examples of suitable reagents include but are not limited to, nucleic acid probes capable of specifically hybridizing to the gene of interest, PCR primers capable of specifically amplifying the gene of interest, and antibodies capable of specifically binding to proteins expressed by the gene of interest. Other non-limiting examples can be found in the description and examples below.

As used herein, the term "detecting a decreased or increased expression relative to non-cancerous prostate control" refers to measuring the level of expression of a gene (e.g., the level of mRNA or protein) relative to the level in a non-cancerous prostate control sample. Gene expression can be measured using any suitable method, including but not limited to, those described herein.

As used herein, the term "detecting a change in gene expression (e.g., hepsin, pim-1, or AMACR) in said prostate cell sample in the presence of said test compound relative to the absence of said test compound" refers to measuring an altered level of expression (e.g., increased or decreased) in the presence of a test compound relative to the absence of the test compound. Gene expression can be measured using any suitable method, including but not limited to, those described in Examples 1-5 below.

As used herein, the term "instructions for using said kit for detecting cancer in said subject" includes instructions for using the reagents contained in the kit for the detection and characterization of cancer in a sample from a subject. In some embodiments, the instructions further comprise the statement of intended use required by the U.S. Food and Drug Administration (FDA) in labeling in vitro diagnostic products. The FDA classifies in vitro diagnostics as medical devices and requires that they be approved through the 510(k) procedure. Information required in an application under 510(k) includes: 1) The in vitro diagnostic product name, including the trade or proprietary name, the common or usual name, and the classification name of the device; 2) The intended use of the product; 3) The establishment registration number, if 10 applicable, of the owner or operator submitting the 510(k) submission; the class in which the in vitro diagnostic product was placed under section 513 of the FD&C Act, if known, its appropriate panel, or, if the owner or operator determines that the device has not been classified under such section, a statement of that determination and the basis for the determination that the in vitro diagnostic product is not so classified; 4) Proposed labels, 15 labeling and advertisements sufficient to describe the in vitro diagnostic product, its intended use, and directions for use. Where applicable, photographs or engineering drawings should be supplied; 5) A statement indicating that the device is similar to and/or different from other in vitro diagnostic products of comparable type in commercial 20 distribution in the U.S., accompanied by data to support the statement; 6) A 510(k) summary of the safety and effectiveness data upon which the substantial equivalence determination is based; or a statement that the 510(k) safety and effectiveness information supporting the FDA finding of substantial equivalence will be made available to any person within 30 days of a written request; 7) A statement that the submitter believes, to the best of their knowledge, that all data and information submitted 25 in the premarket notification are truthful and accurate and that no material fact has been omitted; 8) Any additional information regarding the in vitro diagnostic product requested that is necessary for the FDA to make a substantial equivalency determination. Additional information is available at the Internet web page of the U.S. FDA.

As used herein, the term "prostate cancer expression profile map" refers to a presentation of expression levels of genes in a particular type of prostate tissue (e.g.,

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primary, metastatic, and pre-cancerous prostate tissues). The map may be presented as a graphical representation (e.g., on paper or on a computer screen), a physical representation (e.g., a gel or array) or a digital representation stored in computer memory. Each map corresponds to a particular type of prostate tissue (e.g., primary, metastatic, and pre-cancerous) and thus provides a template for comparison to a patient sample. In preferred embodiments, maps are generated from pooled samples comprising tissue samples from a plurality of patients with the same type of tissue.

As used herein, the terms "computer memory" and "computer memory device" refer to any storage media readable by a computer processor. Examples of computer memory include, but are not limited to, RAM, ROM, computer chips, digital video disc (DVDs), compact discs (CDs), hard disk drives (HDD), and magnetic tape.

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As used herein, the term "computer readable medium" refers to any device or system for storing and providing information (e.g., data and instructions) to a computer processor. Examples of computer readable media include, but are not limited to, DVDs, CDs, hard disk drives, magnetic tape and servers for streaming media over networks.

As used herein, the terms "processor" and "central processing unit" or "CPU" are used interchangeably and refer to a device that is able to read a program from a computer memory (e.g., ROM or other computer memory) and perform a set of steps according to the program.

As used herein, the term "stage of cancer" refers to a qualitative or quantitative assessment of the level of advancement of a cancer. Criteria used to determine the stage of a cancer include, but are not limited to, the size of the tumor, whether the tumor has spread to other parts of the body and where the cancer has spread (e.g., within the same organ or region of the body or to another organ).

As used herein, the term "providing a prognosis" refers to providing information regarding the impact of the presence of cancer (e.g., as determined by the diagnostic methods of the present invention) on a subject's future health (e.g., expected morbidity or mortality, the likelihood of getting cancer, and the risk of metastasis).

As used herein, the term "prostate specific antigen failure" refers to the development of high prostate specific antigen levels in a patient following prostate cancer therapy (e.g., surgery). See Examples 3 and 4 for examples of how prostate specific

antigen failure is determined. As used herein, the term "risk of developing prostate specific antigen failure" refers to a subject's relative risk (e.g., the percent chance or a relative score) of developing prostate specific antigen failure following prostate cancer therapy.

As used herein, the term "post surgical tumor tissue" refers to cancerous tissue (e.g., prostate tissue) that has been removed from a subject (e.g., during surgery).

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As used herein, the term "subject diagnosed with a cancer" refers to a subject who has been tested and found to have cancerous cells. The cancer may be diagnosed using any suitable method, including but not limited to, biopsy, x-ray, blood test, and the diagnostic methods of the present invention.

As used herein, the term "initial diagnosis" refers to results of initial cancer diagnosis (e.g. the presence or absence of cancerous cells). An initial diagnosis does not include information about the stage of the cancer of the risk of prostate specific antigen failure.

As used herein, the term "biopsy tissue" refers to a sample of tissue (e.g., prostate tissue) that is removed from a subject for the purpose of determining if the sample contains cancerous tissue. In some embodiment, biopsy tissue is obtained because a subject is suspected of having cancer. The biopsy tissue is then examined (e.g., by microscopy) for the presence or absence of cancer.

As used herein, the term "inconclusive biopsy tissue" refers to biopsy tissue for which histological examination has not determined the presence or absence of cancer.

As used herein, the term "basal cell marker" refers to a marker (e.g., an antibody) that binds to proteins present in the basal cell layer of benign prostate glands. Exemplary basal cell markers include, but are not limited to, 34βE12 and p63 (See e.g., O'Malley et al., Virchows Arch. Pathol. Anat. Histopathol., 417:191 [1990]; Wojno et al., Am. J. Surg. Pathol., 19:251 [1995]; Googe et al., Am. J. Clin. Pathol., 107:219 [1997]; Parsons et al., Urology 58:619; and Signoretti et al., Am. J. Pathol., 157:1769 [2000]).

As used herein, the term "non-human animals" refers to all non-human animals including, but are not limited to, vertebrates such as rodents, non-human primates, ovines, bovines, ruminants, lagomorphs, porcines, caprines, equines, canines, felines, aves, etc.

As used herein, the term "gene transfer system" refers to any means of delivering a composition comprising a nucleic acid sequence to a cell or tissue. For example, gene transfer systems include, but are not limited to, vectors (e.g., retroviral, adenoviral, adeno-associated viral, and other nucleic acid-based delivery systems), microinjection of naked nucleic acid, polymer-based delivery systems (e.g., liposome-based and metallic particle-based systems), biolistic injection, and the like. As used herein, the term "viral gene transfer system" refers to gene transfer systems comprising viral elements (e.g., intact viruses, modified viruses and viral components such as nucleic acids or proteins) to facilitate delivery of the sample to a desired cell or tissue. As used herein, the term "adenovirus gene transfer system" refers to gene transfer systems comprising intact or altered viruses belonging to the family Adenoviridae.

As used herein, the term "site-specific recombination target sequences" refers to nucleic acid sequences that provide recognition sequences for recombination factors and the location where recombination takes place.

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As used herein, the term "nucleic acid molecule" refers to any nucleic acid 15 containing molecule, including but not limited to, DNA or RNA. The term encompasses sequences that include any of the known base analogs of DNA and RNA including, but not limited to, 4-acetylcytosine, 8-hydroxy-N6-methyladenosine, aziridinylcytosine, pseudoisocytosine, 5-(carboxyhydroxylmethyl) uracil, 5-fluorouracil, 5-bromouracil, 5carboxymethylaminomethyl-2-thiouracil, 5-carboxymethylaminomethyluracil, 20 dihydrouracil, inosine, N6-isopentenyladenine, 1-methyladenine, 1-methylpseudouracil, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-methyladenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarbonylmethyluracil, 5-methoxyuracil, 25 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid, oxybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, N-uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid, pseudouracil, queosine, 2-thiocytosine, and 2.6-diaminopurine. 30

The term "gene" refers to a nucleic acid (e.g., DNA) sequence that comprises coding sequences necessary for the production of a polypeptide, precursor, or RNA (e.g., rRNA, tRNA). The polypeptide can be encoded by a full length coding sequence or by any portion of the coding sequence so long as the desired activity or functional properties (e.g., enzymatic activity, ligand binding, signal transduction, immunogenicity, etc.) of the full-length or fragment are retained. The term also encompasses the coding region of a structural gene and the sequences located adjacent to the coding region on both the 5' and 3' ends for a distance of about 1 kb or more on either end such that the gene corresponds to the length of the full-length mRNA. Sequences located 5' of the coding region and present on the mRNA are referred to as 5' non-translated sequences. Sequences located 3' or downstream of the coding region and present on the mRNA are referred to as 3' nontranslated sequences. The term "gene" encompasses both cDNA and genomic forms of a gene. A genomic form or clone of a gene contains the coding region interrupted with non-coding sequences termed "introns" or "intervening regions" or "intervening sequences." Introns are segments of a gene that are transcribed into nuclear RNA (hnRNA); introns may contain regulatory elements such as enhancers. Introns are removed or "spliced out" from the nuclear or primary transcript; introns therefore are absent in the messenger RNA (mRNA) transcript. The mRNA functions during translation to specify the sequence or order of amino acids in a nascent polypeptide.

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As used herein, the term "heterologous gene" refers to a gene that is not in its natural environment. For example, a heterologous gene includes a gene from one species introduced into another species. A heterologous gene also includes a gene native to an organism that has been altered in some way (e.g., mutated, added in multiple copies, linked to non-native regulatory sequences, etc). Heterologous genes are distinguished from endogenous genes in that the heterologous gene sequences are typically joined to DNA sequences that are not found naturally associated with the gene sequences in the chromosome or are associated with portions of the chromosome not found in nature (e.g., genes expressed in loci where the gene is not normally expressed).

As used herein, the term "gene expression" refers to the process of converting genetic information encoded in a gene into RNA (e.g., mRNA, rRNA, tRNA, or snRNA) through "transcription" of the gene (i.e., via the enzymatic action of an RNA

polymerase), and for protein encoding genes, into protein through "translation" of mRNA. Gene expression can be regulated at many stages in the process. "Upregulation" or "activation" refers to regulation that increases the production of gene expression products (i.e., RNA or protein), while "down-regulation" or "repression" refers to regulation that decrease production. Molecules (e.g., transcription factors) that are involved in up-regulation or down-regulation are often called "activators" and "repressors," respectively.

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In addition to containing introns, genomic forms of a gene may also include sequences located on both the 5' and 3' end of the sequences that are present on the RNA transcript. These sequences are referred to as "flanking" sequences or regions (these flanking sequences are located 5' or 3' to the non-translated sequences present on the mRNA transcript). The 5' flanking region may contain regulatory sequences such as promoters and enhancers that control or influence the transcription of the gene. The 3' flanking region may contain sequences that direct the termination of transcription, post-transcriptional cleavage and polyadenylation.

The term "wild-type" refers to a gene or gene product isolated from a naturally occurring source. A wild-type gene is that which is most frequently observed in a population and is thus arbitrarily designed the "normal" or "wild-type" form of the gene. In contrast, the term "modified" or "mutant" refers to a gene or gene product that displays modifications in sequence and or functional properties (i.e., altered characteristics) when compared to the wild-type gene or gene product. It is noted that naturally occurring mutants can be isolated; these are identified by the fact that they have altered characteristics (including altered nucleic acid sequences) when compared to the wild-type gene or gene product.

As used herein, the terms "nucleic acid molecule encoding," "DNA sequence encoding," and "DNA encoding" refer to the order or sequence of deoxyribonucleotides along a strand of deoxyribonucleic acid. The order of these deoxyribonucleotides determines the order of amino acids along the polypeptide (protein) chain. The DNA sequence thus codes for the amino acid sequence.

As used herein, the terms "an oligonucleotide having a nucleotide sequence encoding a gene" and "polynucleotide having a nucleotide sequence encoding a gene,"

means a nucleic acid sequence comprising the coding region of a gene or in other words the nucleic acid sequence that encodes a gene product. The coding region may be present in a cDNA, genomic DNA or RNA form. When present in a DNA form, the oligonucleotide or polynucleotide may be single-stranded (*i.e.*, the sense strand) or double-stranded. Suitable control elements such as enhancers/promoters, splice junctions, polyadenylation signals, etc. may be placed in close proximity to the coding region of the gene if needed to permit proper initiation of transcription and/or correct processing of the primary RNA transcript. Alternatively, the coding region utilized in the expression vectors of the present invention may contain endogenous enhancers/promoters, splice junctions, intervening sequences, polyadenylation signals, etc. or a combination of both endogenous and exogenous control elements.

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As used herein, the term "oligonucleotide," refers to a short length of single-stranded polynucleotide chain. Oligonucleotides are typically less than 200 residues long (e.g., between 15 and 100), however, as used herein, the term is also intended to encompass longer polynucleotide chains. Oligonucleotides are often referred to by their length. For example a 24 residue oligonucleotide is referred to as a "24-mer". Oligonucleotides can form secondary and tertiary structures by self-hybridizing or by hybridizing to other polynucleotides. Such structures can include, but are not limited to, duplexes, hairpins, cruciforms, bends, and triplexes.

As used herein, the terms "complementary" or "complementarity" are used in reference to polynucleotides (i.e., a sequence of nucleotides) related by the base-pairing rules. For example, for the sequence "A-G-T," is complementary to the sequence "T-C-A." Complementarity may be "partial," in which only some of the nucleic acids' bases are matched according to the base pairing rules. Or, there may be "complete" or "total" complementarity between the nucleic acids. The degree of complementarity between nucleic acid strands has significant effects on the efficiency and strength of hybridization between nucleic acid strands. This is of particular importance in amplification reactions, as well as detection methods that depend upon binding between nucleic acids.

The term "homology" refers to a degree of complementarity. There may be partial homology or complete homology (i.e., identity). A partially complementary sequence is a nucleic acid molecule that at least partially inhibits a completely

complementary nucleic acid molecule from hybridizing to a target nucleic acid is "substantially homologous." The inhibition of hybridization of the completely complementary sequence to the target sequence may be examined using a hybridization assay (Southern or Northern blot, solution hybridization and the like) under conditions of low stringency. A substantially homologous sequence or probe will compete for and inhibit the binding (i.e., the hybridization) of a completely homologous nucleic acid molecule to a target under conditions of low stringency. This is not to say that conditions of low stringency are such that non-specific binding is permitted; low stringency conditions require that the binding of two sequences to one another be a specific (i.e., selective) interaction. The absence of non-specific binding may be tested by the use of a second target that is substantially non-complementary (e.g., less than about 30% identity); in the absence of non-specific binding the probe will not hybridize to the second non-complementary target.

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When used in reference to a double-stranded nucleic acid sequence such as a cDNA or genomic clone, the term "substantially homologous" refers to any probe that can hybridize to either or both strands of the double-stranded nucleic acid sequence under conditions of low stringency as described above.

A gene may produce multiple RNA species that are generated by differential splicing of the primary RNA transcript. cDNAs that are splice variants of the same gene will contain regions of sequence identity or complete homology (representing the presence of the same exon or portion of the same exon on both cDNAs) and regions of complete non-identity (for example, representing the presence of exon "A" on cDNA 1 wherein cDNA 2 contains exon "B" instead). Because the two cDNAs contain regions of sequence identity they will both hybridize to a probe derived from the entire gene or portions of the gene containing sequences found on both cDNAs; the two splice variants are therefore substantially homologous to such a probe and to each other.

When used in reference to a single-stranded nucleic acid sequence, the term "substantially homologous" refers to any probe that can hybridize (i.e., it is the complement of) the single-stranded nucleic acid sequence under conditions of low stringency as described above.

WO 03/012067 PCT/US02/24567.

As used herein, the term "hybridization" is used in reference to the pairing of complementary nucleic acids. Hybridization and the strength of hybridization (i.e., the strength of the association between the nucleic acids) is impacted by such factors as the degree of complementary between the nucleic acids, stringency of the conditions involved, the T_m of the formed hybrid, and the G:C ratio within the nucleic acids. A single molecule that contains pairing of complementary nucleic acids within its structure is said to be "self-hybridized."

As used herein, the term " T_m " is used in reference to the "melting temperature." The melting temperature is the temperature at which a population of double-stranded nucleic acid molecules becomes half dissociated into single strands. The equation for calculating the T_m of nucleic acids is well known in the art. As indicated by standard references, a simple estimate of the T_m value may be calculated by the equation: $T_m = 81.5 + 0.41(\% G + C)$, when a nucleic acid is in aqueous solution at 1 M NaCl (See e.g., Anderson and Young, Quantitative Filter Hybridization, in Nucleic Acid Hybridization [1985]). Other references include more sophisticated computations that take structural as well as sequence characteristics into account for the calculation of T_m .

As used herein the term "stringency" is used in reference to the conditions of temperature, ionic strength, and the presence of other compounds such as organic solvents, under which nucleic acid hybridizations are conducted. Under "low stringency conditions" a nucleic acid sequence of interest will hybridize to its exact complement, sequences with single base mismatches, closely related sequences (e.g., sequences with 90% or greater homology), and sequences having only partial homology (e.g., sequences with 50-90% homology). Under 'medium stringency conditions," a nucleic acid sequence of interest will hybridize only to its exact complement, sequences with single base mismatches, and closely relation sequences (e.g., 90% or greater homology). Under "high stringency conditions," a nucleic acid sequence of interest will hybridize only to its exact complement, and (depending on conditions such a temperature) sequences with single base mismatches. In other words, under conditions of high stringency the temperature can be raised so as to exclude hybridization to sequences with single base mismatches.

"High stringency conditions" when used in reference to nucleic acid hybridization comprise conditions equivalent to binding or hybridization at 42°C in a solution consisting of 5X SSPE (43.8 g/l NaCl, 6.9 g/l NaH₂PO₄ H₂O and 1.85 g/l EDTA, pH adjusted to 7.4 with NaOH), 0.5% SDS, 5X Denhardt's reagent and 100 μg/ml denatured salmon sperm DNA followed by washing in a solution comprising 0.1X SSPE, 1.0% SDS at 42°C when a probe of about 500 nucleotides in length is employed.

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"Medium stringency conditions" when used in reference to nucleic acid hybridization comprise conditions equivalent to binding or hybridization at 42°C in a solution consisting of 5X SSPE (43.8 g/l NaCl, 6.9 g/l NaH₂PO₄ H₂O and 1.85 g/l EDTA, pH adjusted to 7.4 with NaOH), 0.5% SDS, 5X Denhardt's reagent and 100 μg/ml denatured salmon sperm DNA followed by washing in a solution comprising 1.0X SSPE, 1.0% SDS at 42°C when a probe of about 500 nucleotides in length is employed.

"Low stringency conditions" comprise conditions equivalent to binding or hybridization at 42°C in a solution consisting of 5X SSPE (43.8 g/l NaCl, 6.9 g/l NaH₂PO₄ H₂O and 1.85 g/l EDTA, pH adjusted to 7.4 with NaOH), 0.1% SDS, 5X Denhardt's reagent [50X Denhardt's contains per 500 ml: 5 g Ficoll (Type 400, Pharamcia), 5 g BSA (Fraction V; Sigma)] and 100 μg/ml denatured salmon sperm DNA followed by washing in a solution comprising 5X SSPE, 0.1% SDS at 42°C when a probe of about 500 nucleotides in length is employed.

The art knows well that numerous equivalent conditions may be employed to comprise low stringency conditions; factors such as the length and nature (DNA, RNA, base composition) of the probe and nature of the target (DNA, RNA, base composition, present in solution or immobilized, etc.) and the concentration of the salts and other components (e.g., the presence or absence of formamide, dextran sulfate, polyethylene glycol) are considered and the hybridization solution may be varied to generate conditions of low stringency hybridization different from, but equivalent to, the above listed conditions. In addition, the art knows conditions that promote hybridization under conditions of high stringency (e.g., increasing the temperature of the hybridization and/or wash steps, the use of formamide in the hybridization solution, etc.) (see definition above for "stringency").

"Amplification" is a special case of nucleic acid replication involving template specificity. It is to be contrasted with non-specific template replication (i.e., replication that is template-dependent but not dependent on a specific template). Template specificity is here distinguished from fidelity of replication (i.e., synthesis of the proper polynucleotide sequence) and nucleotide (ribo- or deoxyribo-) specificity. Template specificity is frequently described in terms of "target" specificity. Target sequences are "targets" in the sense that they are sought to be sorted out from other nucleic acid. Amplification techniques have been designed primarily for this sorting out.

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Template specificity is achieved in most amplification techniques by the choice of enzyme. Amplification enzymes are enzymes that, under conditions they are used, will process only specific sequences of nucleic acid in a heterogeneous mixture of nucleic acid. For example, in the case of QB replicase, MDV-1 RNA is the specific template for the replicase (Kacian et al., Proc. Natl. Acad. Sci. USA 69:3038 [1972]). Other nucleic acids will not be replicated by this amplification enzyme. Similarly, in the case of T7 RNA polymerase, this amplification enzyme has a stringent specificity for its own promoters (Chamberlin et al., Nature 228:227 [1970]). In the case of T4 DNA ligase, the enzyme will not ligate the two oligonucleotides or polynucleotides, where there is a mismatch between the oligonucleotide or polynucleotide substrate and the template at the ligation junction (Wu and Wallace, Genomics 4:560 [1989]). Finally, Taq and Pfu polymerases, by virtue of their ability to function at high temperature, are found to display high specificity for the sequences bounded and thus defined by the primers; the high temperature results in thermodynamic conditions that favor primer hybridization with the target sequences and not hybridization with non-target sequences (H.A. Erlich (ed.), PCR Technology, Stockton Press [1989]).

As used herein, the term "amplifiable nucleic acid" is used in reference to nucleic acids that may be amplified by any amplification method. It is contemplated that "amplifiable nucleic acid" will usually comprise "sample template."

As used herein, the term "sample template" refers to nucleic acid originating from a sample that is analyzed for the presence of "target." In contrast, "background template" is used in reference to nucleic acid other than sample template that may or may not be present in a sample. Background template is most often inadvertent. It may be the result

of carryover, or it may be due to the presence of nucleic acid contaminants sought to be purified away from the sample. For example, nucleic acids from organisms other than those to be detected may be present as background in a test sample.

As used herein, the term "primer" refers to an oligonucleotide, whether occurring naturally as in a purified restriction digest or produced synthetically, that is capable of acting as a point of initiation of synthesis when placed under conditions in which synthesis of a primer extension product that is complementary to a nucleic acid strand is induced, (*i.e.*, in the presence of nucleotides and an inducing agent such as DNA polymerase and at a suitable temperature and pH). The primer is preferably single stranded for maximum efficiency in amplification, but may alternatively be double stranded. If double stranded, the primer is first treated to separate its strands before being used to prepare extension products. Preferably, the primer is an oligodeoxyribonucleotide. The primer must be sufficiently long to prime the synthesis of extension products in the presence of the inducing agent. The exact lengths of the primers will depend on many factors, including temperature, source of primer and the use of the method.

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As used herein, the term "probe" refers to an oligonucleotide (i.e., a sequence of nucleotides), whether occurring naturally as in a purified restriction digest or produced synthetically, recombinantly or by PCR amplification, that is capable of hybridizing to at least a portion of another oligonucleotide of interest. A probe may be single-stranded or double-stranded. Probes are useful in the detection, identification and isolation of particular gene sequences. It is contemplated that any probe used in the present invention will be labeled with any "reporter molecule," so that is detectable in any detection system, including, but not limited to enzyme (e.g., ELISA, as well as enzyme-based histochemical assays), fluorescent, radioactive, and luminescent systems. It is not intended that the present invention be limited to any particular detection system or label.

As used herein the term "portion" when in reference to a nucleotide sequence (as in "a portion of a given nucleotide sequence") refers to fragments of that sequence. The fragments may range in size from four nucleotides to the entire nucleotide sequence minus one nucleotide (10 nucleotides, 20, 30, 40, 50, 100, 200, etc.).

As used herein, the term "target," refers to the region of nucleic acid bounded by the primers. Thus, the "target" is sought to be sorted out from other nucleic acid sequences. A "segment" is defined as a region of nucleic acid within the target sequence.

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As used herein, the term "polymerase chain reaction" ("PCR") refers to the method of K.B. Mullis U.S. Patent Nos. 4,683,195 4,683,202, and 4,965,188, hereby incorporated by reference, which describe a method for increasing the concentration of a segment of a target sequence in a mixture of genomic DNA without cloning or purification. This process for amplifying the target sequence consists of introducing a large excess of two oligonucleotide primers to the DNA mixture containing the desired target sequence, followed by a precise sequence of thermal cycling in the presence of a DNA polymerase. The two primers are complementary to their respective strands of the double stranded target sequence. To effect amplification, the mixture is denatured and the primers then annealed to their complementary sequences within the target molecule. Following annealing, the primers are extended with a polymerase so as to form a new pair of complementary strands. The steps of denaturation, primer annealing and polymerase extension can be repeated many times (i.e., denaturation, annealing and extension constitute one "cycle"; there can be numerous "cycles") to obtain a high concentration of an amplified segment of the desired target sequence. The length of the amplified segment of the desired target sequence is determined by the relative positions of the primers with respect to each other, and therefore, this length is a controllable parameter. By virtue of the repeating aspect of the process, the method is referred to as the "polymerase chain reaction" (hereinafter "PCR"). Because the desired amplified segments of the target sequence become the predominant sequences (in terms of concentration) in the mixture, they are said to be "PCR amplified".

With PCR, it is possible to amplify a single copy of a specific target sequence in genomic DNA to a level detectable by several different methodologies (e.g., hybridization with a labeled probe; incorporation of biotinylated primers followed by avidin-enzyme conjugate detection; incorporation of ³²P-labeled deoxynucleotide triphosphates, such as dCTP or dATP, into the amplified segment). In addition to genomic DNA, any oligonucleotide or polynucleotide sequence can be amplified with the appropriate set of primer molecules. In particular, the amplified segments created by the

PCR process are, themselves, efficient templates for subsequent PCR amplifications.

As used herein, the terms "PCR product," "PCR fragment," and "amplification product" refer to the resultant mixture of compounds after two or more cycles of the PCR steps of denaturation, annealing and extension are complete. These terms encompass the case where there has been amplification of one or more segments of one or more target sequences.

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As used herein, the term "amplification reagents" refers to those reagents (deoxyribonucleotide triphosphates, buffer, etc.), needed for amplification except for primers, nucleic acid template and the amplification enzyme. Typically, amplification reagents along with other reaction components are placed and contained in a reaction vessel (test tube, microwell, etc.).

As used herein, the terms "restriction endonucleases" and "restriction enzymes" refer to bacterial enzymes, each of which cut double-stranded DNA at or near a specific nucleotide sequence.

The terms "in operable combination," "in operable order," and "operably linked" as used herein refer to the linkage of nucleic acid sequences in such a manner that a nucleic acid molecule capable of directing the transcription of a given gene and/or the synthesis of a desired protein molecule is produced. The term also refers to the linkage of amino acid sequences in such a manner so that a functional protein is produced.

The term "isolated" when used in relation to a nucleic acid, as in "an isolated oligonucleotide" or "isolated polynucleotide" refers to a nucleic acid sequence that is identified and separated from at least one component or contaminant with which it is ordinarily associated in its natural source. Isolated nucleic acid is such present in a form or setting that is different from that in which it is found in nature. In contrast, non-isolated nucleic acids as nucleic acids such as DNA and RNA found in the state they exist in nature. For example, a given DNA sequence (e.g., a gene) is found on the host cell chromosome in proximity to neighboring genes; RNA sequences, such as a specific mRNA sequence encoding a specific protein, are found in the cell as a mixture with numerous other mRNAs that encode a multitude of proteins. However, isolated nucleic acid encoding a given protein includes, by way of example, such nucleic acid in cells ordinarily expressing the given protein where the nucleic acid is in a chromosomal

location different from that of natural cells, or is otherwise flanked by a different nucleic acid sequence than that found in nature. The isolated nucleic acid, oligonucleotide, or polynucleotide may be present in single-stranded or double-stranded form. When an isolated nucleic acid, oligonucleotide or polynucleotide is to be utilized to express a protein, the oligonucleotide or polynucleotide will contain at a minimum the sense or coding strand (i.e., the oligonucleotide or polynucleotide may be single-stranded), but may contain both the sense and anti-sense strands (i.e., the oligonucleotide or polynucleotide may be double-stranded).

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As used herein, the term "purified" or "to purify" refers to the removal of components (e.g., contaminants) from a sample. For example, antibodies are purified by removal of contaminating non-immunoglobulin proteins; they are also purified by the removal of immunoglobulin that does not bind to the target molecule. The removal of non-immunoglobulin proteins and/or the removal of immunoglobulins that do not bind to the target molecule results in an increase in the percent of target-reactive immunoglobulins in the sample. In another example, recombinant polypeptides are expressed in bacterial host cells and the polypeptides are purified by the removal of host cell proteins; the percent of recombinant polypeptides is thereby increased in the sample.

"Amino acid sequence" and terms such as "polypeptide" or "protein" are not meant to limit the amino acid sequence to the complete, native amino acid sequence associated with the recited protein molecule.

The term "native protein" as used herein to indicate that a protein does not contain amino acid residues encoded by vector sequences; that is, the native protein contains only those amino acids found in the protein as it occurs in nature. A native protein may be produced by recombinant means or may be isolated from a naturally occurring source.

As used herein the term "portion" when in reference to a protein (as in "a portion of a given protein") refers to fragments of that protein. The fragments may range in size from four amino acid residues to the entire amino acid sequence minus one amino acid.

The term "Southern blot," refers to the analysis of DNA on agarose or acrylamide gels to fractionate the DNA according to size followed by transfer of the DNA from the gel to a solid support, such as nitrocellulose or a nylon membrane. The immobilized DNA is then probed with a labeled probe to detect DNA species complementary to the

probe used. The DNA may be cleaved with restriction enzymes prior to electrophoresis. Following electrophoresis, the DNA may be partially depurinated and denatured prior to or during transfer to the solid support. Southern blots are a standard tool of molecular biologists (J. Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, NY, pp 9.31-9.58 [1989]).

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The term "Northern blot," as used herein refers to the analysis of RNA by electrophoresis of RNA on agarose gels to fractionate the RNA according to size followed by transfer of the RNA from the gel to a solid support, such as nitrocellulose or a nylon membrane. The immobilized RNA is then probed with a labeled probe to detect RNA species complementary to the probe used. Northern blots are a standard tool of molecular biologists (J. Sambrook, et al., supra, pp 7.39-7.52 [1989]).

The term "Western blot" refers to the analysis of protein(s) (or polypeptides) immobilized onto a support such as nitrocellulose or a membrane. The proteins are run on acrylamide gels to separate the proteins, followed by transfer of the protein from the gel to a solid support, such as nitrocellulose or a nylon membrane. The immobilized proteins are then exposed to antibodies with reactivity against an antigen of interest. The binding of the antibodies may be detected by various methods, including the use of radiolabeled antibodies.

The term "transgene" as used herein refers to a foreign gene that is placed into an organism by, for example, introducing the foreign gene into newly fertilized eggs or early embryos. The term "foreign gene" refers to any nucleic acid (e.g., gene sequence) that is introduced into the genome of an animal by experimental manipulations and may include gene sequences found in that animal so long as the introduced gene does not reside in the same location as does the naturally occurring gene.

As used herein, the term "vector" is used in reference to nucleic acid molecules that transfer DNA segment(s) from one cell to another. The term "vehicle" is sometimes used interchangeably with "vector." Vectors are often derived from plasmids, bacteriophages, or plant or animal viruses.

The term "expression vector" as used herein refers to a recombinant DNA molecule containing a desired coding sequence and appropriate nucleic acid sequences necessary for the expression of the operably linked coding sequence in a particular host

organism. Nucleic acid sequences necessary for expression in prokaryotes usually include a promoter, an operator (optional), and a ribosome binding site, often along with other sequences. Eukaryotic cells are known to utilize promoters, enhancers, and termination and polyadenylation signals.

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The terms "overexpression" and "overexpressing" and grammatical equivalents, are used in reference to levels of mRNA to indicate a level of expression approximately 3-fold higher (or greater) than that observed in a given tissue in a control or non-transgenic animal. Levels of mRNA are measured using any of a number of techniques known to those skilled in the art including, but not limited to Northern blot analysis. Appropriate controls are included on the Northern blot to control for differences in the amount of RNA loaded from each tissue analyzed (e.g., the amount of 28S rRNA, an abundant RNA transcript present at essentially the same amount in all tissues, present in each sample can be used as a means of normalizing or standardizing the mRNA-specific signal observed on Northern blots). The amount of mRNA present in the band corresponding in size to the correctly spliced transgene RNA is quantified; other minor species of RNA which hybridize to the transgene probe are not considered in the quantification of the expression of the transgenic mRNA.

The term "transfection" as used herein refers to the introduction of foreign DNA into eukaryotic cells. Transfection may be accomplished by a variety of means known to the art including calcium phosphate-DNA co-precipitation, DEAE-dextran-mediated transfection, polybrene-mediated transfection, electroporation, microinjection, liposome fusion, lipofection, protoplast fusion, retroviral infection, and biolistics.

The term "calcium phosphate co-precipitation" refers to a technique for the introduction of nucleic acids into a cell. The uptake of nucleic acids by cells is enhanced when the nucleic acid is presented as a calcium phosphate-nucleic acid co-precipitate. The original technique of Graham and van der Eb (Graham and van der Eb, Virol., 52:456 [1973]), has been modified by several groups to optimize conditions for particular types of cells. The art is well aware of these numerous modifications.

The term "stable transfection" or "stably transfected" refers to the introduction and integration of foreign DNA into the genome of the transfected cell. The term "stable

transfectant" refers to a cell that has stably integrated foreign DNA into the genomic DNA.

The term "transient transfection" or "transiently transfected" refers to the introduction of foreign DNA into a cell where the foreign DNA fails to integrate into the genome of the transfected cell. The foreign DNA persists in the nucleus of the transfected cell for several days. During this time the foreign DNA is subject to the regulatory controls that govern the expression of endogenous genes in the chromosomes. The term "transient transfectant" refers to cells that have taken up foreign DNA but have failed to integrate this DNA.

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As used herein, the term "selectable marker" refers to the use of a gene that encodes an enzymatic activity that confers the ability to grow in medium lacking what would otherwise be an essential nutrient (e.g. the HIS3 gene in yeast cells); in addition, a selectable marker may confer resistance to an antibiotic or drug upon the cell in which the selectable marker is expressed. Selectable markers may be "dominant"; a dominant selectable marker encodes an enzymatic activity that can be detected in any eukaryotic cell line. Examples of dominant selectable markers include the bacterial aminoglycoside 3' phosphotransferase gene (also referred to as the neo gene) that confers resistance to the drug G418 in mammalian cells, the bacterial hygromycin G phosphotransferase (hyg) gene that confers resistance to the antibiotic hygromycin and the bacterial xanthineguanine phosphoribosyl transferase gene (also referred to as the gpt gene) that confers the ability to grow in the presence of mycophenolic acid. Other selectable markers are not dominant in that their use must be in conjunction with a cell line that lacks the relevant enzyme activity. Examples of non-dominant selectable markers include the thymidine kinase (tk) gene that is used in conjunction with tk - cell lines, the CAD gene that is used in conjunction with CAD-deficient cells and the mammalian hypoxanthine-guanine phosphoribosyl transferase (hprt) gene that is used in conjunction with hprt - cell lines. A review of the use of selectable markers in mammalian cell lines is provided in Sambrook, J. et al., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory Press, New York (1989) pp.16.9-16.15.

As used herein, the term "cell culture" refers to any in vitro culture of cells.

Included within this term are continuous cell lines (e.g., with an immortal phenotype),

primary cell cultures, transformed cell lines, finite cell lines (e.g., non-transformed cells), and any other cell population maintained in vitro.

As used, the term "eukaryote" refers to organisms distinguishable from "prokaryotes." It is intended that the term encompass all organisms with cells that exhibit the usual characteristics of eukaryotes, such as the presence of a true nucleus bounded by a nuclear membrane, within which lie the chromosomes, the presence of membrane-bound organelles, and other characteristics commonly observed in eukaryotic organisms. Thus, the term includes, but is not limited to such organisms as fungi, protozoa, and animals (e.g., humans).

As used herein, the term "in vitro" refers to an artificial environment and to processes or reactions that occur within an artificial environment. In vitro environments can consist of, but are not limited to, test tubes and cell culture. The term "in vivo" refers to the natural environment (e.g., an animal or a cell) and to processes or reaction that occur within a natural environment.

The terms "test compound" and "candidate compound" refer to any chemical entity, pharmaceutical, drug, and the like that is a candidate for use to treat or prevent a disease, illness, sickness, or disorder of bodily function (e.g., cancer). Test compounds comprise both known and potential therapeutic compounds. A test compound can be determined to be therapeutic by screening using the screening methods of the present invention. In some embodiments of the present invention, test compounds include antisense compounds.

As used herein, the term "sample" is used in its broadest sense. In one sense, it is meant to include a specimen or culture obtained from any source, as well as biological and environmental samples. Biological samples may be obtained from animals (including humans) and encompass fluids, solids, tissues, and gases. Biological samples include blood products, such as plasma, serum and the like. Environmental samples include environmental material such as surface matter, soil, water, crystals and industrial samples. Such examples are not however to be construed as limiting the sample types applicable to the present invention.

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DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to compositions and methods for cancer diagnostics, including but not limited to, cancer markers. In particular, the present invention provides gene expression profiles associated with prostate cancers. Accordingly, the present invention provides method of characterizing prostate tissues, kits for the detection of markers, as well as drug screening and therapeutic applications.

I. Markers for Prostate Cancer

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The present invention provides markers whose expression is specifically altered in cancerous prostate tissues. Such markers find use in the diagnosis and characterization of prostate cancer.

A. Identification of Markers

Experiments conducted during the development of the present invention resulted in the identification of genes whose expression level was altered (e.g., increased or decreased) in PCA. The methods utilized glass slide cDNA microarrays that included approximately 5000 known, named genes, 4400 ESTs, and 500 control elements, as well as normal and cancerous prostate tissue. Differentially expressed genes were divided into functional clusters. The expression of relevant genes was confirmed using Western blot analysis. Protein expression in prostate tissues was measured for several genes of interest.

The methods of the present invention (See e.g., Example 2) were used to identify clusters of genes that were up or down regulated in PCA, benign prostate tissue, precancerous tissue, and normal prostate. From these clusters, two genes, hepsin and pim-1 were identified as genes that were of particular relevance. Immunohistochemistry (See e.g., Example 4) was used to characterize the presence of hepsin and pim-1 proteins in prostate tissue. Hepsin was found to stain strongly in pre-cancerous tissue (HG-PIN). In addition, hepsin was found to stain less strongly in PCA tissues of men found to have an increased risk of metastasis as measured by PSA failure (increased PSA following surgery), thus confirming the diagnostic utility of hepsin. In addition, deceased expression of pim-1 in PCA tissue was also found to be associated with increased risk of

PSA failure. Accordingly, in some embodiments, the present invention provides methods of detecting and characterizing prostate tissues.

The methods of the present invention identified a further gene alpha-methyl-CoA racemase (AMACR) that was found to be expressed in PCA, but not benign prostate tissue (See e.g., Example 5). AMACR was found to be present in the serum and urine of prostate or bladder cancer patients. In addition, a humoral response to AMACR was identified. In still further embodiments, the methods of the present invention were used to characterize the EZH2 gene. EZH2 was found to be up-regulated in metastatic prostate cancer. The inhibition of EZH2 expression in prostate cells inhibited cell proliferation in vitro, as well as inducing transcriptional repression of a variety of genes. The methods of the present invention further identified CtBP1 and CTBP2, as well as that GP73 as being over-expressed in metastatic prostate cancer relative to localized prostate cancer and benign tissue.

In still further embodiments, the methods of the present invention identified annexins 1, 2, 4, 7 and 11 as being significantly decreased in hormone refractory PCA when compared to localized hormone naïve Pca. Tissue microarray analysis revealed a significant decrease in protein expression for annexins 1, 2, 4, 7 and 11 in hormone refractory PCA as compared to localized Pca. No significant differences were detected between the clinically localized PCA and non-cancerous prostate tissues.

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B. Detection of Markers

In some embodiments, the present invention provides methods for detection of expression of cancer markers (e.g., prostate cancer markers). In preferred embodiments, expression is measured directly (e.g., at the RNA or protein level). In some embodiments, expression is detected in tissue samples (e.g., biopsy tissue). In other embodiments, expression is detected in bodily fluids (e.g., including but not limited to, plasma, serum, whole blood, mucus, and urine). The present invention further provides panels and kits for the detection of markers. In preferred embodiments, the presence of a cancer marker is used to provide a prognosis to a subject. For example, the detection of hepsin or pim-1 in prostate tissues is indicative of a cancer that is likely to metastasize and the expression of hepsin is indicative of a pre-cancerous tissue that is likely to

become cancerous. In addition, the expression of AMACR is indicative of cancerous tissue. The information provided is also used to direct the course of treatment. For example, if a subject is found to have a marker indicative of a highly metastasizing tumor, additional therapies (e.g., hormonal or radiation therapies) can be started at a earlier point when they are more likely to be effective (e.g., before metastasis). In addition, if a subject is found to have a tumor that is not responsive to hormonal therapy, the expense and inconvenience of such therapies can be avoided.

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The present invention is not limited to the markers described above. Any suitable marker that correlates with cancer or the progression of cancer may be utilized, including but not limited to, those described in the illustrative examples below (e.g., FKBP5, 10 FASN, FOLH1, TNFSF10, PCM1, S100A11, IGFBP3, SLUG, GSTM3, ATF2, RAB5A, IL1R2, ITGB4, CCND2, EDNRB, APP, THROMBOSPONDIN 1, ANNEXIN A1, EPHA1, NCK1, MAPK6, SGK, HEVIN, MEIS2, MYLK, FZD7, CAVEOLIN 2, TACC1, ARHB, PSG9, GSTM1, KERATIN 5, TIMP2, GELSOLIN, ITM2C, GSTM5, VINCULIN, FHL1, GSTP1, MEIS1, ETS2, PPP2CB, CATHEPSIN B, CATHEPSIN H, 15 COL1A2, RIG, VIMENTIN, MOESIN, MCAM, FIBRONECTIN 1, NBL1, ANNEXIN A4, ANEXIN A11, IL1R1, IGFBP5, CYSTATIN C, COL15A1, ADAMTS1, SKI, EGR1, FOSB, CFLAR, JUN, YWHAB, NRAS, C7, SCYA2, ITGA1, LUMICAN, C1S, C4BPA, COL3A1, FAT, MMECD10, CLUSTERIN, PLA2G2A, MADh4, SEPP1, RAB2, PP1CB, MPDZ, PRKCL2, CTBP1, CTBP2, MAP3K10, TBXA2F, MTA1, 20 RAP2, TRAP1, TFCP2, E2EPF, UBCH10, TASTIN, EZH2, FLS353, MYBL2, LIMK1, GP73, VAV2, TOP2A, ASNS, CTBP, AMACR, ABCC5 (MDR5), and TRAF4. Additional markers are also contemplated to be within the scope of the present invention. Any suitable method may be utilized to identify and characterize cancer markers suitable for use in the methods of the present invention, including but not limited to, those 25 described in illustrative Examples 1-15 below. For example, in some embodiments, markers identified as being up or down-regulated in PCA using the gene expression microarray methods of the present invention are further characterized using tissue microarray, immunohistochemistry, Northern blot analysis, siRNA or antisense RNA inhibition, mutation analysis, investigation of expression with clinical outcome, as well as 30 other methods disclosed herein.

In some embodiments, the present invention provides a panel for the analysis of a plurality of markers. The panel allows for the simultaneous analysis of multiple markers correlating with carcinogenesis and/or metastasis. For example, a panel may include markers identified as correlating with cancerous tissue, metastatic cancer, localized cancer that is likely to metastasize, pre-cancerous tissue that is likely to become cancerous, and pre-cancerous tissue that is not likely to become cancerous. Depending on the subject, panels may be analyzed alone or in combination in order to provide the best possible diagnosis and prognosis. Markers for inclusion on a panel are selected by screening for their predictive value using any suitable method, including but not limited to, those described in the illustrative examples below.

In other embodiments, the present invention provides an expression profile map comprising expression profiles of cancers of various stages or prognoses (e.g., likelihood of future metastasis). Such maps can be used for comparison with patient samples. In some embodiments comparisons are made using the method described in Example 2. However, the present invention is not limited to the method described in Example 2. Any suitable method may be utilized, including but not limited to, by computer comparison of digitized data. The comparison data is used to provide diagnoses and/or prognoses to patients.

1. Detection of RNA

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In some preferred embodiments, detection of prostate cancer markers (e.g., including but not limited to, those disclosed herein) is detected by measuring the expression of corresponding mRNA in a tissue sample (e.g., prostate tissue). mRNA expression may be measured by any suitable method, including but not limited to, those disclosed below.

In some embodiments, RNA is detection by Northern blot analysis. Northern blot analysis involves the separation of RNA and hybridization of a complementary labeled probe. An exemplary method for Northern blot analysis is provided in Example 3.

In other embodiments, RNA expression is detected by enzymatic cleavage of specific structures (INVADER assay, Third Wave Technologies; *See e.g.*, U.S. Patent Nos. 5,846,717, 6,090,543; 6,001,567; 5,985,557; and 5,994,069; each of which is herein

incorporated by reference). The INVADER assay detects specific nucleic acid (e.g., RNA) sequences by using structure-specific enzymes to cleave a complex formed by the hybridization of overlapping oligonucleotide probes.

In still further embodiments, RNA (or corresponding cDNA) is detected by hybridization to a oligonucleotide probe). A variety of hybridization assays using a variety of technologies for hybridization and detection are available. For example, in some embodiments, TaqMan assay (PE Biosystems, Foster City, CA; See e.g., U.S. Patent Nos. 5,962,233 and 5,538,848, each of which is herein incorporated by reference) is utilized. The assay is performed during a PCR reaction. The TaqMan assay exploits the 5'-3' exonuclease activity of the AMPLITAQ GOLD DNA polymerase. A probe consisting of an oligonucleotide with a 5'-reporter dye (e.g., a fluorescent dye) and a 3'-quencher dye is included in the PCR reaction. During PCR, if the probe is bound to its target, the 5'-3' nucleolytic activity of the AMPLITAQ GOLD polymerase cleaves the probe between the reporter and the quencher dye. The separation of the reporter dye from the quencher dye results in an increase of fluorescence. The signal accumulates with each cycle of PCR and can be monitored with a fluorimeter.

In yet other embodiments, reverse-transcriptase PCR (RT-PCR) is used to detect the expression of RNA. In RT-PCR, RNA is enzymatically converted to complementary DNA or "cDNA" using a reverse transcriptase enzyme. The cDNA is then used as a template for a PCR reaction. PCR products can be detected by any suitable method, including but not limited to, gel electrophoresis and staining with a DNA specific stain or hybridization to a labeled probe. In some embodiments, the quantitative reverse transcriptase PCR with standardized mixtures of competitive templates method described in U.S. Patents 5,639,606, 5,643,765, and 5,876,978 (each of which is herein incorporated by reference) is utilized.

2. Detection of Protein

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In other embodiments, gene expression of cancer markers is detected by measuring the expression of the corresponding protein or polypeptide. Protein expression may be detected by any suitable method. In some embodiments, proteins are detected by the immunohistochemistry method of Example 4. In other embodiments, proteins are

detected by their binding to an antibody raised against the protein. The generation of antibodies is described below.

Antibody binding is detected by techniques known in the art (e.g., radioimmunoassay, ELISA (enzyme-linked immunosorbant assay), "sandwich" immunoassays, immunoradiometric assays, gel diffusion precipitation reactions, immunodiffusion assays, in situ immunoassays (e.g., using colloidal gold, enzyme or radioisotope labels, for example), Western blots, precipitation reactions, agglutination assays (e.g., gel agglutination assays, hemagglutination assays, etc.), complement fixation assays, immunofluorescence assays, protein A assays, and immunoelectrophoresis assays, etc.

In one embodiment, antibody binding is detected by detecting a label on the primary antibody. In another embodiment, the primary antibody is detected by detecting binding of a secondary antibody or reagent to the primary antibody. In a further embodiment, the secondary antibody is labeled. Many methods are known in the art for detecting binding in an immunoassay and are within the scope of the present invention.

In some embodiments, an automated detection assay is utilized. Methods for the automation of immunoassays include those described in U.S. Patents 5,885,530, 4,981,785, 6,159,750, and 5,358,691, each of which is herein incorporated by reference. In some embodiments, the analysis and presentation of results is also automated. For example, in some embodiments, software that generates a prognosis based on the presence or absence of a series of proteins corresponding to cancer markers is utilized.

In other embodiments, the immunoassay described in U.S. Patents 5,599,677 and 5,672,480; each of which is herein incorporated by reference.

25 3. Data Analysis

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In some embodiments, a computer-based analysis program is used to translate the raw data generated by the detection assay (e.g., the presence, absence, or amount of a given marker or markers) into data of predictive value for a clinician. The clinician can access the predictive data using any suitable means. Thus, in some preferred embodiments, the present invention provides the further benefit that the clinician, who is not likely to be trained in genetics or molecular biology, need not understand the raw

data. The data is presented directly to the clinician in its most useful form. The clinician is then able to immediately utilize the information in order to optimize the care of the subject.

The present invention contemplates any method capable of receiving, processing, and transmitting the information to and from laboratories conducting the assays, information provides, medical personal, and subjects. For example, in some embodiments of the present invention, a sample (e.g., a biopsy or a serum or urine sample) is obtained from a subject and submitted to a profiling service (e.g., clinical lab at a medical facility, genomic profiling business, etc.), located in any part of the world (e.g., in a country different than the country where the subject resides or where the information is ultimately used) to generate raw data. Where the sample comprises a tissue or other biological sample, the subject may visit a medical center to have the sample obtained and sent to the profiling center, or subjects may collect the sample themselves (e.g., a urine sample) and directly send it to a profiling center. Where the sample comprises previously determined biological information, the information may be directly sent to the profiling service by the subject (e.g., an information card containing the information may be scanned by a computer and the data transmitted to a computer of the profiling center using an electronic communication systems). Once received by the profiling service, the sample is processed and a profile is produced (i.e., expression data), specific for the diagnostic or prognostic information desired for the subject.

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The profile data is then prepared in a format suitable for interpretation by a treating clinician. For example, rather than providing raw expression data, the prepared format may represent a diagnosis or risk assessment (e.g., likelihood of metastasis or PSA failure) for the subject, along with recommendations for particular treatment options. The data may be displayed to the clinician by any suitable method. For example, in some embodiments, the profiling service generates a report that can be printed for the clinician (e.g., at the point of care) or displayed to the clinician on a computer monitor.

In some embodiments, the information is first analyzed at the point of care or at a regional facility. The raw data is then sent to a central processing facility for further analysis and/or to convert the raw data to information useful for a clinician or patient.

The central processing facility provides the advantage of privacy (all data is stored in a

central facility with uniform security protocols), speed, and uniformity of data analysis. The central processing facility can then control the fate of the data following treatment of the subject. For example, using an electronic communication system, the central facility can provide data to the clinician, the subject, or researchers.

In some embodiments, the subject is able to directly access the data using the electronic communication system. The subject may chose further intervention or counseling based on the results. In some embodiments, the data is used for research use. For example, the data may be used to further optimize the inclusion or elimination of markers as useful indicators of a particular condition or stage of disease.

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4. Kits

In yet other embodiments, the present invention provides kits for the detection and characterization of prostate cancer. In some embodiments, the kits contain antibodies specific for a cancer marker, in addition to detection reagents and buffers. In other embodiments, the kits contain reagents specific for the detection of mRNA or cDNA (e.g., oligonucleotide probes or primers). In preferred embodiments, the kits contain all of the components necessary to perform a detection assay, including all controls, directions for performing assays, and any necessary software for analysis and presentation of results.

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5. In vivo Imaging

In some embodiments, in vivo imaging techniques are used to visualize the expression of cancer markers in an animal (e.g., a human or non-human mammal). For example, in some embodiments, cancer marker mRNA or protein is labeled using an labeled antibody specific for the cancer marker. A specifically bound and labeled antibody can be detected in an individual using an in vivo imaging method, including, but not limited to, radionuclide imaging, positron emission tomography, computerized axial tomography, X-ray or magnetic resonance imaging method, fluorescence detection, and chemiluminescent detection. Methods for generating antibodies to the cancer markers of the present invention are described below.

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The in vivo imaging methods of the present invention are useful in the diagnosis of cancers that express the cancer markers of the present invention (e.g., prostate cancer). In vivo imaging is used to visualize the presence of a marker indicative of the cancer. Such techniques allow for diagnosis without the use of an unpleasant biopsy. The in vivo imaging methods of the present invention are also useful for providing prognoses to cancer patients. For example, the presence of a marker indicative of cancers likely to metastasize can be detected. The in vivo imaging methods of the present invention can further be used to detect metastatic cancers in other parts of the body.

In some embodiments, reagents (e.g., antibodies) specific for the cancer markers of the present invention are fluorescently labeled. The labeled antibodies are introduced into a subject (e.g., orally or parenterally). Fluorescently labeled antibodies are detected using any suitable method (e.g., using the apparatus described in U.S. Patent 6,198,107, herein incorporated by reference).

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In other embodiments, antibodies are radioactively labeled. The use of antibodies 15 for in vivo diagnosis is well known in the art. Sumerdon et al., (Nucl. Med. Biol 17:247-254 [1990] have described an optimized antibody-chelator for the radioimmunoscintographic imaging of tumors using Indium-111 as the label, Griffin et al., (J Clin Onc 9:631-640 [1991]) have described the use of this agent in detecting tumors in patients suspected of having recurrent colorectal cancer. The use of similar 20 agents with paramagnetic ions as labels for magnetic resonance imaging is known in the art (Lauffer, Magnetic Resonance in Medicine 22:339-342 [1991]). The label used will depend on the imaging modality chosen. Radioactive labels such as Indium-111, Technetium-99m, or Iodine-131 can be used for planar scans or single photon emission computed tomography (SPECT). Positron emitting labels such as Fluorine-19 can also 25 be used for positron emission tomography (PET). For MRI, paramagnetic ions such as Gadolinium (III) or Manganese (II) can be used.

Radioactive metals with half-lives ranging from 1 hour to 3.5 days are available for conjugation to antibodies, such as scandium-47 (3.5 days) gallium-67 (2.8 days), gallium-68 (68 minutes), technetium-99m (6 hours), and indium-111 (3.2 days), of which gallium-67, technetium-99m, and indium-111 are preferable for gamma camera imaging, gallium-68 is preferable for positron emission tomography.

A useful method of labeling antibodies with such radiometals is by means of a bifunctional chelating agent, such as diethylenetriaminepentaacetic acid (DTPA), as described, for example, by Khaw et al. (Science 209:295 [1980]) for In-111 and Tc-99m, and by Scheinberg et al. (Science 215:1511 [1982]). Other chelating agents may also be used, but the 1-(p-carboxymethoxybenzyl)EDTA and the carboxycarbonic anhydride of DTPA are advantageous because their use permits conjugation without affecting the antibody's immunoreactivity substantially.

Another method for coupling DPTA to proteins is by use of the cyclic anhydride of DTPA, as described by Hnatowich *et al.* (Int. J. Appl. Radiat. Isot. 33:327 [1982]) for labeling of albumin with In-111, but which can be adapted for labeling of antibodies. A suitable method of labeling antibodies with Tc-99m which does not use chelation with DPTA is the pretinning method of Crockford *et al.*, (U.S. Pat. No. 4,323,546, herein incorporated by reference).

A preferred method of labeling immunoglobulins with Tc-99m is that described by Wong et al. (Int. J. Appl. Radiat. Isot., 29:251 [1978]) for plasma protein, and recently applied successfully by Wong et al. (J. Nucl. Med., 23:229 [1981]) for labeling antibodies.

In the case of the radiometals conjugated to the specific antibody, it is likewise desirable to introduce as high a proportion of the radiolabel as possible into the antibody molecule without destroying its immunospecificity. A further improvement may be achieved by effecting radiolabeling in the presence of the specific cancer marker of the present invention, to insure that the antigen binding site on the antibody will be protected. The antigen is separated after labeling.

In still further embodiments, in vivo biophotonic imaging (Xenogen, Almeda, CA) is utilized for in vivo imaging. This real-time in vivo imaging utilizes luciferase. The luciferase gene is incorporated into cells, microorganisms, and animals (e.g., as a fusion protein with a cancer marker of the present invention). When active, it leads to a reaction that emits light. A CCD camera and software is used to capture the image and analyze it.

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II. Antibodies

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The present invention provides isolated antibodies. In preferred embodiments, the present invention provides monoclonal antibodies that specifically bind to an isolated polypeptide comprised of at least five amino acid residues of the cancer markers described herein (e.g., hepsin, pim-1, AMACR, EZH2, CTBP). These antibodies find use in the diagnostic methods described herein.

An antibody against a protein of the present invention may be any monoclonal or polyclonal antibody, as long as it can recognize the protein. Antibodies can be produced by using a protein of the present invention as the antigen according to a conventional antibody or antiserum preparation process.

The present invention contemplates the use of both monoclonal and polyclonal antibodies. Any suitable method may be used to generate the antibodies used in the methods and compositions of the present invention, including but not limited to, those disclosed herein. For example, for preparation of a monoclonal antibody, protein, as such, or together with a suitable carrier or diluent is administered to an animal (e.g., a mammal) under conditions that permit the production of antibodies. For enhancing the antibody production capability, complete or incomplete Freund's adjuvant may be administered. Normally, the protein is administered once every 2 weeks to 6 weeks, in total, about 2 times to about 10 times. Animals suitable for use in such methods include, but are not limited to, primates, rabbits, dogs, guinea pigs, mice, rats, sheep, goats, etc.

For preparing monoclonal antibody-producing cells, an individual animal whose antibody titer has been confirmed (e.g., a mouse) is selected, and 2 days to 5 days after the final immunization, its spleen or lymph node is harvested and antibody-producing cells contained therein are fused with myeloma cells to prepare the desired monoclonal antibody producer hybridoma. Measurement of the antibody titer in antiserum can be carried out, for example, by reacting the labeled protein, as described hereinafter and antiserum and then measuring the activity of the labeling agent bound to the antibody. The cell fusion can be carried out according to known methods, for example, the method described by Koehler and Milstein (Nature 256:495 [1975]). As a fusion promoter, for example, polyethylene glycol (PEG) or Sendai virus (HVJ), preferably PEG is used.

Examples of myeloma cells include NS-1, P3U1, SP2/0, AP-1 and the like. The proportion of the number of antibody producer cells (spleen cells) and the number of myeloma cells to be used is preferably about 1:1 to about 20:1. PEG (preferably PEG 1000-PEG 6000) is preferably added in concentration of about 10% to about 80%. Cell fusion can be carried out efficiently by incubating a mixture of both cells at about 20°C to about 40°C, preferably about 30°C to about 37°C for about 1 minute to 10 minutes.

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Various methods may be used for screening for a hybridoma producing the antibody (e.g., against a tumor antigen or autoantibody of the present invention). For example, where a supernatant of the hybridoma is added to a solid phase (e.g., microplate) to which antibody is adsorbed directly or together with a carrier and then an anti-immunoglobulin antibody (if mouse cells are used in cell fusion, anti-mouse immunoglobulin antibody is used) or Protein A labeled with a radioactive substance or an enzyme is added to detect the monoclonal antibody against the protein bound to the solid phase. Alternately, a supernatant of the hybridoma is added to a solid phase to which an anti-immunoglobulin antibody or Protein A is adsorbed and then the protein labeled with a radioactive substance or an enzyme is added to detect the monoclonal antibody against the protein bound to the solid phase.

Selection of the monoclonal antibody can be carried out according to any known method or its modification. Normally, a medium for animal cells to which HAT (hypoxanthine, aminopterin, thymidine) are added is employed. Any selection and growth medium can be employed as long as the hybridoma can grow. For example, RPMI 1640 medium containing 1% to 20%, preferably 10% to 20% fetal bovine serum, GIT medium containing 1% to 10% fetal bovine serum, a serum free medium for cultivation of a hybridoma (SFM-101, Nissui Seiyaku) and the like can be used. Normally, the cultivation is carried out at 20°C to 40°C, preferably 37°C for about 5 days to 3 weeks, preferably 1 week to 2 weeks under about 5% CO₂ gas. The antibody titer of the supernatant of a hybridoma culture can be measured according to the same manner as described above with respect to the antibody titer of the anti-protein in the antiserum.

Separation and purification of a monoclonal antibody (e.g., against a cancer marker of the present invention) can be carried out according to the same manner as those of conventional polyclonal antibodies such as separation and purification of

immunoglobulins, for example, salting-out, alcoholic precipitation, isoelectric point precipitation, electrophoresis, adsorption and desorption with ion exchangers (e.g., DEAE), ultracentrifugation, gel filtration, or a specific purification method wherein only an antibody is collected with an active adsorbent such as an antigen-binding solid phase, Protein A or Protein G and dissociating the binding to obtain the antibody.

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Polyclonal antibodies may be prepared by any known method or modifications of these methods including obtaining antibodies from patients. For example, a complex of an immunogen (an antigen against the protein) and a carrier protein is prepared and an animal is immunized by the complex according to the same manner as that described with respect to the above monoclonal antibody preparation. A material containing the antibody against is recovered from the immunized animal and the antibody is separated and purified.

As to the complex of the immunogen and the carrier protein to be used for immunization of an animal, any carrier protein and any mixing proportion of the carrier and a hapten can be employed as long as an antibody against the hapten, which is crosslinked on the carrier and used for immunization, is produced efficiently. For example, bovine serum albumin, bovine cycloglobulin, keyhole limpet hemocyanin, etc. may be coupled to an hapten in a weight ratio of about 0.1 part to about 20 parts, preferably, about 1 part to about 5 parts per 1 part of the hapten.

In addition, various condensing agents can be used for coupling of a hapten and a carrier. For example, glutaraldehyde, carbodiimide, maleimide activated ester, activated ester reagents containing thiol group or dithiopyridyl group, and the like find use with the present invention. The condensation product as such or together with a suitable carrier or diluent is administered to a site of an animal that permits the antibody production. For enhancing the antibody production capability, complete or incomplete Freund's adjuvant may be administered. Normally, the protein is administered once every 2 weeks to 6 weeks, in total, about 3 times to about 10 times.

The polyclonal antibody is recovered from blood, ascites and the like, of an animal immunized by the above method. The antibody titer in the antiserum can be measured according to the same manner as that described above with respect to the supernatant of the hybridoma culture. Separation and purification of the antibody can be

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carried out according to the same separation and purification method of immunoglobulin as that described with respect to the above monoclonal antibody.

The protein used herein as the immunogen is not limited to any particular type of immunogen. For example, a cancer marker of the present invention (further including a gene having a nucleotide sequence partly altered) can be used as the immunogen. Further, fragments of the protein may be used. Fragments may be obtained by any methods including, but not limited to expressing a fragment of the gene, enzymatic processing of the protein, chemical synthesis, and the like.

Drug Screening 10 Ш.

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In some embodiments, the present invention provides drug screening assays (e.g., to screen for anticancer drugs). The screening methods of the present invention utilize cancer markers identified using the methods of the present invention (e.g., including but not limited to, hepsin, pim-1, AMACR, EZH2, and CTBP). For example, in some embodiments, the present invention provides methods of screening for compound that alter (e.g., increase or decrease) the expression of cancer marker genes. In some embodiments, candidate compounds are antisense agents (e.g., oligonucleotides) directed against cancer markers. See Section IV below for a discussion of antisense therapy. In other embodiments, candidate compounds are antibodies that specifically bind to a cancer marker of the present invention.

In one screening method, candidate compounds are evaluated for their ability to alter cancer marker expression by contacting a compound with a cell expressing a cancer marker and then assaying for the effect of the candidate compounds on expression. In some embodiments, the effect of candidate compounds on expression of a cancer marker gene is assayed for by detecting the level of cancer marker mRNA expressed by the cell. mRNA expression can be detected by any suitable method. In other embodiments, the effect of candidate compounds on expression of cancer marker genes is assayed by measuring the level of polypeptide encoded by the cancer markers. The level of polypeptide expressed can be measured using any suitable method, including but not limited to, those disclosed herein.

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Specifically, the present invention provides screening methods for identifying modulators, *i.e.*, candidate or test compounds or agents (*e.g.*, proteins, peptides, peptidomimetics, peptoids, small molecules or other drugs) which bind to cancer markers of the present invention, have an inhibitory (or stimulatory) effect on, for example, cancer marker expression or cancer markers activity, or have a stimulatory or inhibitory effect on, for example, the expression or activity of a cancer marker substrate. Compounds thus identified can be used to modulate the activity of target gene products (*e.g.*, cancer marker genes) either directly or indirectly in a therapeutic protocol, to elaborate the biological function of the target gene product, or to identify compounds that disrupt normal target gene interactions. Compounds which inhibit the activity or expression of cancer markers are useful in the treatment of proliferative disorders, *e.g.*, cancer, particularly metastatic (*e.g.*, androgen independent) prostate cancer.

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In one embodiment, the invention provides assays for screening candidate or test compounds that are substrates of a cancer markers protein or polypeptide or a biologically active portion thereof. In another embodiment, the invention provides assays for screening candidate or test compounds that bind to or modulate the activity of a cancer marker protein or polypeptide or a biologically active portion thereof.

The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including biological libraries; peptoid libraries (libraries of molecules having the functionalities of peptides, but with a novel, non-peptide backbone, which are resistant to enzymatic degradation but which nevertheless remain bioactive; see, e.g., Zuckennann et al., J. Med. Chem. 37: 2678-85 [1994]); spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the 'one-bead one-compound' library method; and synthetic library methods using affinity chromatography selection. The biological library and peptoid library approaches are preferred for use with peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam (1997) Anticancer Drug Des. 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt et al., Proc. Natl. Acad. Sci. U.S.A. 90:6909 [1993]; Erb et

al., Proc. Nad. Acad. Sci. USA 91:11422 [1994]; Zuckermann et al., J. Med. Chem. 37:2678 [1994]; Cho et al., Science 261:1303 [1993]; Carrell et al., Angew. Chem. Int. Ed. Engl. 33:2059 [1994]; Carell et al., Angew. Chem. Int. Ed. Engl. 33:2061 [1994]; and Gallop et al., J. Med. Chem. 37:1233 [1994].

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Libraries of compounds may be presented in solution (e.g., Houghten, Biotechniques 13:412-421 [1992]), or on beads (Lam, Nature 354:82-84 [1991]), chips (Fodor, Nature 364:555-556 [1993]), bacteria or spores (U.S. Patent No. 5,223,409; herein incorporated by reference), plasmids (Cull et al., Proc. Nad. Acad. Sci. USA 89:18651869 [1992]) or on phage (Scott and Smith, Science 249:386-390 [1990]; Devlin Science 249:404-406 [1990]; Cwirla et al., Proc. Natl. Acad. Sci. 87:6378-6382 [1990]; Felici, J. Mol. Biol. 222:301 [1991]).

In one embodiment, an assay is a cell-based assay in which a cell that expresses a cancer marker protein or biologically active portion thereof is contacted with a test compound, and the ability of the test compound to the modulate cancer marker's activity is determined. Determining the ability of the test compound to modulate cancer marker activity can be accomplished by monitoring, for example, changes in enzymatic activity. The cell, for example, can be of mammalian origin.

The ability of the test compound to modulate cancer marker binding to a compound, e.g., a cancer marker substrate, can also be evaluated. This can be accomplished, for example, by coupling the compound, e.g., the substrate, with a radioisotope or enzymatic label such that binding of the compound, e.g., the substrate, to a cancer marker can be determined by detecting the labeled compound, e.g., substrate, in a complex.

Alternatively, the cancer marker is coupled with a radioisotope or enzymatic label to monitor the ability of a test compound to modulate cancer marker binding to a cancer markers substrate in a complex. For example, compounds (e.g., substrates) can be labeled with ¹²⁵I, ³⁵S ¹⁴C or ³H, either directly or indirectly, and the radioisotope detected by direct counting of radioemmission or by scintillation counting. Alternatively, compounds can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by determination of conversion of an appropriate substrate to product.

The ability of a compound (e.g., a cancer marker substrate) to interact with a cancer marker with or without the labeling of any of the interactants can be evaluated. For example, a microphysiometer can be used to detect the interaction of a compound with a cancer marker without the labeling of either the compound or the cancer marker (McConnell et al. Science 257:1906-1912 [1992]). As used herein, a "microphysiometer" (e.g., Cytosensor) is an analytical instrument that measures the rate at which a cell acidifies its environment using a light-addressable potentiometric sensor (LAPS). Changes in this acidification rate can be used as an indicator of the interaction between a compound and cancer markers.

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In yet another embodiment, a cell-free assay is provided in which a cancer marker protein or biologically active portion thereof is contacted with a test compound and the ability of the test compound to bind to the cancer marker protein or biologically active portion thereof is evaluated. Preferred biologically active portions of the cancer markers proteins to be used in assays of the present invention include fragments that participate in interactions with substrates or other proteins, *e.g.*, fragments with high surface probability scores.

Cell-free assays involve preparing a reaction mixture of the target gene protein and the test compound under conditions and for a time sufficient to allow the two components to interact and bind, thus forming a complex that can be removed and/or detected.

The interaction between two molecules can also be detected, e.g., using fluorescence energy transfer (FRET) (see, for example, Lakowicz et al., U.S. Patent No. 5,631,169; Stavrianopoulos et al., U.S. Patent No. 4,968,103; each of which is herein incorporated by reference). A fluorophore label is selected such that a first donor molecule's emitted fluorescent energy will be absorbed by a fluorescent label on a second, 'acceptor' molecule, which in turn is able to fluoresce due to the absorbed energy.

Alternately, the 'donor' protein molecule may simply utilize the natural fluorescent energy of tryptophan residues. Labels are chosen that emit different wavelengths of light, such that the 'acceptor' molecule label may be differentiated from that of the 'donor'. Since the efficiency of energy transfer between the labels is related to the distance separating the molecules, the spatial relationship between the molecules can

be assessed. In a situation in which binding occurs between the molecules, the fluorescent emission of the 'acceptor' molecule label in 1 5 the assay should be maximal. An FRET binding event can be conveniently measured through standard fluorometric detection means well known in the art (e.g., using a fluorimeter).

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In another embodiment, determining the ability of the cancer markers protein to bind to a target molecule can be accomplished using real-time Biomolecular Interaction Analysis (BIA) (see, e.g., Sjolander and Urbaniczky, Anal. Chem. 63:2338-2345 [1991] and Szabo et al. Curr. Opin. Struct. Biol. 5:699-705 [1995]). "Surface plasmon resonance" or "BIA" detects biospecific interactions in real time, without labeling any of the interactants (e.g., BlAcore). Changes in the mass at the binding surface (indicative of a binding event) result in alterations of the refractive index of light near the surface (the optical phenomenon of surface plasmon resonance (SPR)), resulting in a detectable signal that can be used as an indication of real-time reactions between biological molecules.

In one embodiment, the target gene product or the test substance is anchored onto a solid phase. The target gene product/test compound complexes anchored on the solid phase can be detected at the end of the reaction. Preferably, the target gene product can be anchored onto a solid surface, and the test compound, (which is not anchored), can be labeled, either directly or indirectly, with detectable labels discussed herein.

It may be desirable to immobilize cancer markers, an anti-cancer marker antibody or its target molecule to facilitate separation of complexed from non-complexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to a cancer marker protein, or interaction of a cancer marker protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtiter plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided which adds a domain that allows one or both of the proteins to be bound to a matrix. For example, glutathione-S-transferase-cancer marker fusion proteins or glutathione-S-transferase/target fusion proteins can be adsorbed onto glutathione Sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione-derivatized microtiter plates, which are then combined with the test compound or the test compound and either the non-adsorbed target protein or cancer

marker protein, and the mixture incubated under conditions conducive for complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtiter plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above.

Alternatively, the complexes can be dissociated from the matrix, and the level of cancer markers binding or activity determined using standard techniques. Other techniques for immobilizing either cancer markers protein or a target molecule on matrices include using conjugation of biotin and streptavidin. Biotinylated cancer marker protein or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, EL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical).

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In order to conduct the assay, the non-immobilized component is added to the coated surface containing the anchored component. After the reaction is complete, unreacted components are removed (e.g., by washing) under conditions such that any complexes formed will remain immobilized on the solid surface. The detection of complexes anchored on the solid surface can be accomplished in a number of ways. Where the previously non-immobilized component is pre-labeled, the detection of label immobilized on the surface indicates that complexes were formed. Where the previously non-immobilized component is not pre-labeled, an indirect label can be used to detect complexes anchored on the surface; e.g., using a labeled antibody specific for the immobilized component (the antibody, in turn, can be directly labeled or indirectly labeled with, e.g., a labeled anti-IgG antibody).

This assay is performed utilizing antibodies reactive with cancer marker protein or target molecules but which do not interfere with binding of the cancer markers protein to its target molecule. Such antibodies can be derivatized to the wells of the plate, and unbound target or cancer markers protein trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the cancer marker protein or target molecule, as well as enzyme-linked

assays which rely on detecting an enzymatic activity associated with the cancer marker protein or target molecule.

Alternatively, cell free assays can be conducted in a liquid phase. In such an assay, the reaction products are separated from unreacted components, by any of a number of standard techniques, including, but not limited to: differential centrifugation (see, for example, Rivas and Minton, Trends Biochem Sci 18:284-7 [1993]); chromatography (gel filtration chromatography, ion-exchange chromatography); electrophoresis (see, e.g., Ausubel et al., eds. Current Protocols in Molecular Biology 1999, J. Wiley: New York.); and immunoprecipitation (see, for example, Ausubel et al., eds. Current Protocols in Molecular Biology 1999, J. Wiley: New York). Such resins and chromatographic techniques are known to one skilled in the art (See e.g., Heegaard J. Mol. Recognit 11:141-8 [1998]; Hageand Tweed J. Chromatogr. Biomed. Sci. App1 699:499-525 [1997]). Further, fluorescence energy transfer may also be conveniently utilized, as described herein, to detect binding without further purification of the complex from solution.

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The assay can include contacting the cancer markers protein or biologically active portion thereof with a known compound that binds the cancer marker to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a cancer marker protein, wherein determining the ability of the test compound to interact with a cancer marker protein includes determining the ability of the test compound to preferentially bind to cancer markers or biologically active portion thereof, or to modulate the activity of a target molecule, as compared to the known compound.

To the extent that cancer markers can, *in vivo*, interact with one or more cellular or extracellular macromolecules, such as proteins, inhibitors of such an interaction are useful. A homogeneous assay can be used can be used to identify inhibitors.

For example, a preformed complex of the target gene product and the interactive cellular or extracellular binding partner product is prepared such that either the target gene products or their binding partners are labeled, but the signal generated by the label is quenched due to complex formation (see, e.g., U.S. Patent No. 4,109,496, herein incorporated by reference, that utilizes this approach for immunoassays). The addition of

a test substance that competes with and displaces one of the species from the preformed complex will result in the generation of a signal above background. In this way, test substances that disrupt target gene product-binding partner interaction can be identified. Alternatively, cancer markers protein can be used as a "bait protein" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent No. 5,283,317; Zervos et al., Cell 72:223-232 [1993]; Madura et al., J. Biol. Chem. 268.12046-12054 [1993]; Bartel et al., Biotechniques 14:920-924 [1993]; Iwabuchi et al., Oncogene 8:1693-1696 [1993]; and Brent W0 94/10300; each of which is herein incorporated by reference), to identify other proteins, that bind to or interact with cancer markers ("cancer marker-binding proteins" or "cancer marker-bp") and are involved in cancer marker activity. Such cancer marker-bps can be activators or inhibitors of signals by the cancer marker proteins or targets as, for example, downstream elements of a cancer markers-mediated signaling pathway.

Modulators of cancer markers expression can also be identified. For example, a cell or cell free mixture is contacted with a candidate compound and the expression of cancer marker mRNA or protein evaluated relative to the level of expression of cancer marker mRNA or protein in the absence of the candidate compound. When expression of cancer marker mRNA or protein is greater in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of cancer marker mRNA or protein expression. Alternatively, when expression of cancer marker mRNA or protein is less (i.e., statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of cancer marker mRNA or protein expression. The level of cancer markers mRNA or protein expression can be determined by methods described herein for detecting cancer markers mRNA or protein.

A modulating agent can be identified using a cell-based or a cell free assay, and the ability of the agent to modulate the activity of a cancer markers protein can be confirmed *in vivo*, *e.g.*, in an animal such as an animal model for a disease (*e.g.*, an animal with prostate cancer or metastatic prostate cancer; or an animal harboring a xenograft of a prostate cancer from an animal (*e.g.*, human) or cells from a cancer resulting from metastasis of a prostate cancer (*e.g.*, to a lymph node, bone, or liver), or cells from a prostate cancer cell line.

This invention further pertains to novel agents identified by the above-described screening assays (See e.g., below description of cancer therapies). Accordingly, it is within the scope of this invention to further use an agent identified as described herein (e.g., a cancer marker modulating agent, an antisense cancer marker nucleic acid molecule, a siRNA molecule, a cancer marker specific antibody, or a cancer marker-binding partner) in an appropriate animal model (such as those described herein) to determine the efficacy, toxicity, side effects, or mechanism of action, of treatment with such an agent. Furthermore, novel agents identified by the above-described screening assays can be, e.g., used for treatments as described herein.

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IV. Cancer Therapies

In some embodiments, the present invention provides therapies for cancer (e.g., prostate cancer). In some embodiments, therapies target cancer markers (e.g., including but not limited to, hepsin, pim-1, AMACR, EZH2, and CTBP).

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A. Antisense Therapies

In some embodiments, the present invention targets the expression of cancer markers. For example, in some embodiments, the present invention employs compositions comprising oligomeric antisense compounds, particularly oligonucleotides (e.g., those identified in the drug screening methods described above), for use in modulating the function of nucleic acid molecules encoding cancer markers of the present invention, ultimately modulating the amount of cancer marker expressed. This is accomplished by providing antisense compounds that specifically hybridize with one or more nucleic acids encoding cancer markers of the present invention. The specific hybridization of an oligomeric compound with its target nucleic acid interferes with the normal function of the nucleic acid. This modulation of function of a target nucleic acid by compounds that specifically hybridize to it is generally referred to as "antisense." The functions of DNA to be interfered with include replication and transcription. The functions of RNA to be interfered with include all vital functions such as, for example, translocation of the RNA to the site of protein translation, translation of protein from the RNA, splicing of the RNA to yield one or more mRNA species, and catalytic activity that

may be engaged in or facilitated by the RNA. The overall effect of such interference with target nucleic acid function is modulation of the expression of cancer markers of the present invention. In the context of the present invention, "modulation" means either an increase (stimulation) or a decrease (inhibition) in the expression of a gene. For example, expression may be inhibited to potentially prevent tumor proliferation.

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It is preferred to target specific nucleic acids for antisense. "Targeting" an antisense compound to a particular nucleic acid, in the context of the present invention, is a multistep process. The process usually begins with the identification of a nucleic acid sequence whose function is to be modulated. This may be, for example, a cellular gene (or mRNA transcribed from the gene) whose expression is associated with a particular disorder or disease state, or a nucleic acid molecule from an infectious agent. In the present invention, the target is a nucleic acid molecule encoding a cancer marker of the present invention. The targeting process also includes determination of a site or sites within this gene for the antisense interaction to occur such that the desired effect, e.g., detection or modulation of expression of the protein, will result. Within the context of the present invention, a preferred intragenic site is the region encompassing the translation initiation or termination codon of the open reading frame (ORF) of the gene. Since the translation initiation codon is typically 5'-AUG (in transcribed mRNA molecules; 5'-ATG in the corresponding DNA molecule), the translation initiation codon is also referred to as the "AUG codon," the "start codon" or the "AUG start codon". A minority of genes have a translation initiation codon having the RNA sequence 5'-GUG, 5'-UUG or 5'-CUG, and 5'-AUA, 5'-ACG and 5'-CUG have been shown to function in vivo. Thus, the terms "translation initiation codon" and "start codon" can encompass many codon sequences, even though the initiator amino acid in each instance is typically methionine (in eukaryotes) or formylmethionine (in prokaryotes). Eukaryotic and prokaryotic genes may have two or more alternative start codons, any one of which may be preferentially utilized for translation initiation in a particular cell type or tissue, or under a particular set of conditions. In the context of the present invention, "start codon" and "translation initiation codon" refer to the codon or codons that are used in vivo to initiate translation of an mRNA molecule transcribed from a gene encoding a tumor antigen of the present invention, regardless of the sequence(s) of such codons.

Translation termination codon (or "stop codon") of a gene may have one of three sequences (i.e., 5'-UAA, 5'-UAG and 5'-UGA; the corresponding DNA sequences are 5'-TAA, 5'-TAG and 5'-TGA, respectively). The terms "start codon region" and "translation initiation codon region" refer to a portion of such an mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation initiation codon. Similarly, the terms "stop codon region" and "translation termination codon region" refer to a portion of such an mRNA or gene that encompasses from about 25 to about 50 contiguous nucleotides in either direction (i.e., 5' or 3') from a translation termination codon.

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The open reading frame (ORF) or "coding region," which refers to the region between the translation initiation codon and the translation termination codon, is also a region that may be targeted effectively. Other target regions include the 5' untranslated region (5' UTR), referring to the portion of an mRNA in the 5' direction from the translation initiation codon, and thus including nucleotides between the 5' cap site and the translation initiation codon of an mRNA or corresponding nucleotides on the gene, and the 3' untranslated region (3' UTR), referring to the portion of an mRNA in the 3' direction from the translation termination codon, and thus including nucleotides between the translation termination codon and 3' end of an mRNA or corresponding nucleotides on the gene. The 5' cap of an mRNA comprises an N7-methylated guanosine residue joined to the 5'-most residue of the mRNA via a 5'-5' triphosphate linkage. The 5' cap region of an mRNA is considered to include the 5' cap structure itself as well as the first 50 nucleotides adjacent to the cap. The cap region may also be a preferred target region.

Although some eukaryotic mRNA transcripts are directly translated, many contain one or more regions, known as "introns," that are excised from a transcript before it is translated. The remaining (and therefore translated) regions are known as "exons" and are spliced together to form a continuous mRNA sequence. mRNA splice sites (i.e., intron-exon junctions) may also be preferred target regions, and are particularly useful in situations where aberrant splicing is implicated in disease, or where an overproduction of a particular mRNA splice product is implicated in disease. Aberrant fusion junctions due to rearrangements or deletions are also preferred targets. It has also been found that

introns can also be effective, and therefore preferred, target regions for antisense compounds targeted, for example, to DNA or pre-mRNA.

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In some embodiments, target sites for antisense inhibition are identified using commercially available software programs (e.g., Biognostik, Gottingen, Germany; SysArris Software, Bangalore, India; Antisense Research Group, University of Liverpool, Liverpool, England; GeneTrove, Carlsbad, CA). In other embodiments, target sites for antisense inhibition are identified using the accessible site method described in U.S. Patent WO0198537A2, herein incorporated by reference.

Once one or more target sites have been identified, oligonucleotides are chosen that are sufficiently complementary to the target (i.e., hybridize sufficiently well and with sufficient specificity) to give the desired effect. For example, in preferred embodiments of the present invention, antisense oligonucleotides are targeted to or near the start codon.

In the context of this invention, "hybridization," with respect to antisense compositions and methods, means hydrogen bonding, which may be Watson-Crick, Hoogsteen or reversed Hoogsteen hydrogen bonding, between complementary nucleoside or nucleotide bases. For example, adenine and thymine are complementary nucleobases that pair through the formation of hydrogen bonds. It is understood that the sequence of an antisense compound need not be 100% complementary to that of its target nucleic acid to be specifically hybridizable. An antisense compound is specifically hybridizable when binding of the compound to the target DNA or RNA molecule interferes with the normal function of the target DNA or RNA to cause a loss of utility, and there is a sufficient degree of complementarity to avoid non-specific binding of the antisense compound to non-target sequences under conditions in which specific binding is desired (i.e., under physiological conditions in the case of in vivo assays or therapeutic treatment, and in the case of in vitro assays, under conditions in which the assays are performed).

Antisense compounds are commonly used as research reagents and diagnostics. For example, antisense oligonucleotides, which are able to inhibit gene expression with specificity, can be used to elucidate the function of particular genes. Antisense compounds are also used, for example, to distinguish between functions of various members of a biological pathway.

The specificity and sensitivity of antisense is also applied for therapeutic uses. For example, antisense oligonucleotides have been employed as therapeutic moieties in the treatment of disease states in animals and man. Antisense oligonucleotides have been safely and effectively administered to humans and numerous clinical trials are presently underway. It is thus established that oligonucleotides are useful therapeutic modalities that can be configured to be useful in treatment regimes for treatment of cells, tissues, and animals, especially humans.

While antisense oligonucleotides are a preferred form of antisense compound, the present invention comprehends other oligomeric antisense compounds, including but not limited to oligonucleotide mimetics such as are described below. The antisense compounds in accordance with this invention preferably comprise from about 8 to about 30 nucleobases (i.e., from about 8 to about 30 linked bases), although both longer and shorter sequences may find use with the present invention. Particularly preferred antisense compounds are antisense oligonucleotides, even more preferably those comprising from about 12 to about 25 nucleobases.

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Specific examples of preferred antisense compounds useful with the present invention include oligonucleotides containing modified backbones or non-natural internucleoside linkages. As defined in this specification, oligonucleotides having modified backbones include those that retain a phosphorus atom in the backbone and those that do not have a phosphorus atom in the backbone. For the purposes of this specification, modified oligonucleotides that do not have a phosphorus atom in their internucleoside backbone can also be considered to be oligonucleosides.

Preferred modified oligonucleotide backbones include, for example, phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkylphosphotriesters, methyl and other alkyl phosphonates including 3'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates including 3'-amino phosphoramidate and aminoalkylphosphoramidates, thionophosphoramidates, thionoalkylphosphonates, thionoalkylphosphotriesters, and boranophosphates having normal 3'-5' linkages, 2'-5' linked analogs of these, and those having inverted polarity wherein the adjacent pairs of nucleoside units are linked 3'-5' to 5'-3' or 2'-5' to 5'-2'. Various salts, mixed salts and free acid forms are also included.

Preferred modified oligonucleotide backbones that do not include a phosphorus atom therein have backbones that are formed by short chain alkyl or cycloalkyl internucleoside linkages, mixed heteroatom and alkyl or cycloalkyl internucleoside linkages, or one or more short chain heteroatomic or heterocyclic internucleoside linkages. These include those having morpholino linkages (formed in part from the sugar portion of a nucleoside); siloxane backbones; sulfide, sulfoxide and sulfone backbones; formacetyl and thioformacetyl backbones; methylene formacetyl and thioformacetyl backbones; sulfamate backbones; methyleneimino and methylenehydrazino backbones; sulfonate and sulfonamide backbones; amide backbones; and others having mixed N, O, S and CH₂ component parts.

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In other preferred oligonucleotide mimetics, both the sugar and the internucleoside linkage (*i.e.*, the backbone) of the nucleotide units are replaced with novel groups. The base units are maintained for hybridization with an appropriate nucleic acid target compound. One such oligomeric compound, an oligonucleotide mimetic that has been shown to have excellent hybridization properties, is referred to as a peptide nucleic acid (PNA). In PNA compounds, the sugar-backbone of an oligonucleotide is replaced with an amide containing backbone, in particular an aminoethylglycine backbone. The nucleobases are retained and are bound directly or indirectly to aza nitrogen atoms of the amide portion of the backbone. Representative United States patents that teach the preparation of PNA compounds include, but are not limited to, U.S. Pat. Nos.: 5,539,082; 5,714,331; and 5,719,262, each of which is herein incorporated by reference. Further teaching of PNA compounds can be found in Nielsen et al., Science 254:1497 (1991).

Most preferred embodiments of the invention are oligonucleotides with phosphorothioate backbones and oligonucleosides with heteroatom backbones, and in particular --CH₂, --NH--O--CH₂--, --CH₂--N(CH₃)--O--CH₂-- [known as a methylene (methylimino) or MMI backbone], --CH₂--O--N(CH₃)--CH₂--, --CH₂--N(CH₃)--CH₂--, and --O--N(CH₃)--CH₂-- [wherein the native phosphodiester backbone is represented as --O--P--O--CH₂--] of the above referenced U.S. Pat. No. 5,489,677, and the amide backbones of the above referenced U.S. Pat. No.

5,602,240. Also preferred are oligonucleotides having morpholino backbone structures of the above-referenced U.S. Pat. No. 5,034,506.

Modified oligonucleotides may also contain one or more substituted sugar moieties. Preferred oligonucleotides comprise one of the following at the 2' position: OH; F; O-, S-, or N-alkyl; O-, S-, or N-alkenyl; O-, S- or N-alkynyl; or O-alkyl-O-alkyl, wherein the alkyl, alkenyl and alkynyl may be substituted or unsubstituted C1 to C10 alkyl or C_2 to C_{10} alkenyl and alkynyl. Particularly preferred are $O[(CH_2)_nO]_mCH_3$, O(CH₂)_nOCH₃, O(CH₂)_nNH₂, O(CH₂)_nCH₃, O(CH₂)_nONH₂, and O(CH₂)_nON[(CH₂)_nCH₃)]₂, where n and m are from 1 to about 10. Other preferred oligonucleotides comprise one of the following at the 2' position: C₁ to C₁₀ lower alkyl, 10 substituted lower alkyl, alkaryl, aralkyl, O-alkaryl or O-aralkyl, SH, SCH3, OCN, Cl, Br, CN, CF3, OCF3, SOCH3, SO2CH3, ONO2, NO2, N3, NH2, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalkylamino, substituted silyl, an RNA cleaving group, a reporter group, an intercalator, a group for improving the pharmacokinetic 15 properties of an oligonucleotide, or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. A preferred modification includes 2'-methoxyethoxy (2'-O--CH2CH2OCH3, also known as 2'-O-(2-methoxyethyl) or 2'-MOE) (Martin et al., Helv. Chim. Acta 78:486 [1995]) i.e., an alkoxyalkoxy group. A further preferred modification includes 20 2'-dimethylaminooxyethoxy (i.e., a O(CH₂)₂ON(CH₃)₂ group), also known as 2'-DMAOE, and 2'-dimethylaminoethoxyethoxy (also known in the art as 2'-O-dimethylaminoethoxyethyl or 2'-DMAEOE), i.e., 2'-O--CH₂--O--CH₂--N(CH₂)₂.

Other preferred modifications include 2'-methoxy(2'-O--CH₃),
2'-aminopropoxy(2'-OCH₂CH₂CH₂NH₂) and 2'-fluoro (2'-F). Similar modifications
may also be made at other positions on the oligonucleotide, particularly the 3' position of
the sugar on the 3' terminal nucleotide or in 2'-5' linked oligonucleotides and the 5'
position of 5' terminal nucleotide. Oligonucleotides may also have sugar mimetics such
as cyclobutyl moieties in place of the pentofuranosyl sugar.

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Oligonucleotides may also include nucleobase (often referred to in the art simply as "base") modifications or substitutions. As used herein, "unmodified" or "natural" nucleobases include the purine bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U). Modified nucleobases include other synthetic and natural nucleobases such as 5-methylcytosine (5-me-C), 5-hydroxymethyl 5 cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 10 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine. Further nucleobases include those disclosed in U.S. Pat. No. 3,687,808. 15 Certain of these nucleobases are particularly useful for increasing the binding affinity of the oligomeric compounds of the invention. These include 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine. 5-methylcytosine substitutions have been shown to increase nucleic acid duplex stability by 0.6-1.2. degree °C and are presently preferred base substitutions, even more particularly when combined 20 with 2'-O-methoxyethyl sugar modifications.

Another modification of the oligonucleotides of the present invention involves chemically linking to the oligonucleotide one or more moieties or conjugates that enhance the activity, cellular distribution or cellular uptake of the oligonucleotide. Such moieties include but are not limited to lipid moieties such as a cholesterol moiety, cholic acid, a thioether, (e.g., hexyl-S-tritylthiol), a thiocholesterol, an aliphatic chain, (e.g., dodecandiol or undecyl residues), a phospholipid, (e.g., di-hexadecyl-rac-glycerol or triethylammonium 1,2-di-O-hexadecyl-rac-glycero-3-H-phosphonate), a polyamine or a polyethylene glycol chain or adamantane acetic acid, a palmityl moiety, or an octadecylamine or hexylamino-carbonyl-oxycholesterol moiety.

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One skilled in the relevant art knows well how to generate oligonucleotides containing the above-described modifications. The present invention is not limited to the antisensce oligonucleotides described above. Any suitable modification or substitution may be utilized.

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It is not necessary for all positions in a given compound to be uniformly modified, and in fact more than one of the aforementioned modifications may be incorporated in a single compound or even at a single nucleoside within an oligonucleotide. The present invention also includes antisense compounds that are chimeric compounds. "Chimeric" antisense compounds or "chimeras," in the context of the present invention, are antisense compounds, particularly oligonucleotides, which contain two or more chemically distinct regions, each made up of at least one monomer unit, i.e., a nucleotide in the case of an oligonucleotide compound. These oligonucleotides typically contain at least one region wherein the oligonucleotide is modified so as to confer upon the oligonucleotide increased resistance to nuclease degradation, increased cellular uptake, and/or increased binding affinity for the target nucleic acid. An additional region of the oligonucleotide may serve as a substrate for enzymes capable of cleaving RNA:DNA or RNA:RNA hybrids. By way of example, RNaseH is a cellular endonuclease that cleaves the RNA strand of an RNA:DNA duplex. Activation of RNase H, therefore, results in cleavage of the RNA target, thereby greatly enhancing the efficiency of oligonucleotide inhibition of gene expression. Consequently, comparable results can often be obtained with shorter oligonucleotides when chimeric oligonucleotides are used, compared to phosphorothioate deoxyoligonucleotides hybridizing to the same target region. Cleavage of the RNA target can be routinely detected by gel electrophoresis and, if necessary, associated nucleic acid hybridization techniques known in the art.

Chimeric antisense compounds of the present invention may be formed as composite structures of two or more oligonucleotides, modified oligonucleotides, oligonucleosides and/or oligonucleotide mimetics as described above.

The present invention also includes pharmaceutical compositions and formulations that include the antisense compounds of the present invention as described below.

B. Genetic Therapies

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The present invention contemplates the use of any genetic manipulation for use in modulating the expression of cancer markers of the present invention. Examples of genetic manipulation include, but are not limited to, gene knockout (e.g., removing the cancer marker gene from the chromosome using, for example, recombination), expression of antisense constructs with or without inducible promoters, and the like. Delivery of nucleic acid construct to cells in vitro or in vivo may be conducted using any suitable method. A suitable method is one that introduces the nucleic acid construct into the cell such that the desired event occurs (e.g., expression of an antisense construct).

Introduction of molecules carrying genetic information into cells is achieved by any of various methods including, but not limited to, directed injection of naked DNA constructs, bombardment with gold particles loaded with said constructs, and macromolecule mediated gene transfer using, for example, liposomes, biopolymers, and the like. Preferred methods use gene delivery vehicles derived from viruses, including, but not limited to, adenoviruses, retroviruses, vaccinia viruses, and adeno-associated viruses. Because of the higher efficiency as compared to retroviruses, vectors derived from adenoviruses are the preferred gene delivery vehicles for transferring nucleic acid molecules into host cells *in vivo*. Adenoviral vectors have been shown to provide very efficient *in vivo* gene transfer into a variety of solid tumors in animal models and into human solid tumor xenografts in immune-deficient mice. Examples of adenoviral vectors and methods for gene transfer are described in PCT publications WO 00/12738 and WO 00/09675 and U.S. Pat. Appl. Nos. 6,033,908, 6,019,978, 6,001,557, 5,994,132, 5,994,128, 5,994,106, 5,981,225, 5,885,808, 5,872,154, 5,830,730, and 5,824,544, each of which is herein incorporated by reference in its entirety.

Vectors may be administered to subject in a variety of ways. For example, in some embodiments of the present invention, vectors are administered into tumors or tissue associated with tumors using direct injection. In other embodiments, administration is via the blood or lymphatic circulation (See e.g., PCT publication 99/02685 herein incorporated by reference in its entirety). Exemplary dose levels of adenoviral vector are preferably 10⁸ to 10¹¹ vector particles added to the perfusate.

C. Antibody Therapy

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In some embodiments, the present invention provides antibodies that target prostate tumors that express a cancer marker of the present invention (e.g., hepsin, pim-1, EZH2, Annexin, CTBP, GP73, and AMACR). Any suitable antibody (e.g., monoclonal, polyclonal, or synthetic) may be utilized in the therapeutic methods disclosed herein. In preferred embodiments, the antibodies used for cancer therapy are humanized antibodies. Methods for humanizing antibodies are well known in the art (See e.g., U.S. Patents 6,180,370, 5,585,089, 6,054,297, and 5,565,332; each of which is herein incorporated by reference).

In some embodiments, the therapeutic antibodies comprise an antibody generated against a cancer marker of the present invention (e.g., hepsin, pim-1, EZH2, Annexin, CTBP, GP73, and AMACR), wherein the antibody is conjugated to a cytotoxic agent. In such embodiments, a tumor specific therapeutic agent is generated that does not target normal cells, thus reducing many of the detrimental side effects of traditional chemotherapy. For certain applications, it is envisioned that the therapeutic agents will be pharmacologic agents that will serve as useful agents for attachment to antibodies, particularly cytotoxic or otherwise anticellular agents having the ability to kill or suppress the growth or cell division of endothelial cells. The present invention contemplates the use of any pharmacologic agent that can be conjugated to an antibody, and delivered in active form. Exemplary anticellular agents include chemotherapeutic agents, radioisotopes, and cytotoxins. The therapeutic antibodies of the present invention may include a variety of cytotoxic moieties, including but not limited to, radioactive isotopes (e.g., iodine-131, iodine-123, technicium-99m, indium-111, rhenium-188, rhenium-186, gallium-67, copper-67, yttrium-90, iodine-125 or astatine-211), hormones such as a steroid, antimetabolites such as cytosines (e.g., arabinoside, fluorouracil, methotrexate or aminopterin; an anthracycline; mitomycin C), vinca alkaloids (e.g., demecolcine; etoposide; mithramycin), and antitumor alkylating agent such as chlorambucil or melphalan. Other embodiments may include agents such as a coagulant, a cytokine, growth factor, bacterial endotoxin or the lipid A moiety of bacterial endotoxin. For example, in some embodiments, therapeutic agents will include plant-, fungus- or bacteria-derived toxin, such as an A chain toxins, a ribosome inactivating protein, a-

sarcin, aspergillin, restrictocin, a ribonuclease, diphtheria toxin or pseudomonas exotoxin, to mention just a few examples. In some preferred embodiments, deglycosylated ricin A chain is utilized.

In any event, it is proposed that agents such as these may, if desired, be successfully conjugated to an antibody, in a manner that will allow their targeting, internalization, release or presentation to blood components at the site of the targeted tumor cells as required using known conjugation technology (See, e.g., Ghose et al., Methods Enzymol., 93:280 [1983]).

For example, in some embodiments the present invention provides immunotoxins targeted a cancer marker of the present invention (e.g., hepsin, pim-1, EZH2, Annexin, CTBP, GP73, and AMACR). Immunotoxins are conjugates of a specific targeting agent typically a tumor-directed antibody or fragment, with a cytotoxic agent, such as a toxin moiety. The targeting agent directs the toxin to, and thereby selectively kills, cells carrying the targeted antigen. In some embodiments, therapeutic antibodies employ crosslinkers that provide high in vivo stability (Thorpe et al., Cancer Res., 48:6396 [1988]).

In other embodiments, particularly those involving treatment of solid tumors, antibodies are designed to have a cytotoxic or otherwise anticellular effect against the tumor vasculature, by suppressing the growth or cell division of the vascular endothelial cells. This attack is intended to lead to a tumor-localized vascular collapse, depriving the tumor cells, particularly those tumor cells distal of the vasculature, of oxygen and nutrients, ultimately leading to cell death and tumor necrosis.

In preferred embodiments, antibody based therapeutics are formulated as pharmaceutical compositions as described below. In preferred embodiments, administration of an antibody composition of the present invention results in a measurable decrease in cancer (e.g., decrease or elimination of tumor).

D. Pharmaceutical Compositions

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The present invention further provides pharmaceutical compositions (e.g., comprising the antisense or antibody compounds described above). The pharmaceutical compositions of the present invention may be administered in a number of ways

depending upon whether local or systemic treatment is desired and upon the area to be treated. Administration may be topical (including ophthalmic and to mucous membranes including vaginal and rectal delivery), pulmonary (e.g., by inhalation or insufflation of powders or aerosols, including by nebulizer; intratracheal, intranasal, epidermal and transdermal), oral or parenteral. Parenteral administration includes intravenous, intraarterial, subcutaneous, intraperitoneal or intramuscular injection or infusion; or intracranial, e.g., intrathecal or intraventricular, administration.

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Pharmaceutical compositions and formulations for topical administration may include transdermal patches, ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable.

Compositions and formulations for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets or tablets. Thickeners, flavoring agents, diluents, emulsifiers, dispersing aids or binders may be desirable.

Compositions and formulations for parenteral, intrathecal or intraventricular administration may include sterile aqueous solutions that may also contain buffers, diluents and other suitable additives such as, but not limited to, penetration enhancers, carrier compounds and other pharmaceutically acceptable carriers or excipients.

Pharmaceutical compositions of the present invention include, but are not limited to, solutions, emulsions, and liposome-containing formulations. These compositions may be generated from a variety of components that include, but are not limited to, preformed liquids, self-emulsifying solids and self-emulsifying semisolids.

The pharmaceutical formulations of the present invention, which may conveniently be presented in unit dosage form, may be prepared according to conventional techniques well known in the pharmaceutical industry. Such techniques include the step of bringing into association the active ingredients with the pharmaceutical carrier(s) or excipient(s). In general the formulations are prepared by uniformly and intimately bringing into association the active ingredients with liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

The compositions of the present invention may be formulated into any of many possible dosage forms such as, but not limited to, tablets, capsules, liquid syrups, soft gels, suppositories, and enemas. The compositions of the present invention may also be formulated as suspensions in aqueous, non-aqueous or mixed media. Aqueous suspensions may further contain substances that increase the viscosity of the suspension including, for example, sodium carboxymethylcellulose, sorbitol and/or dextran. The suspension may also contain stabilizers.

In one embodiment of the present invention the pharmaceutical compositions may be formulated and used as foams. Pharmaceutical foams include formulations such as, but not limited to, emulsions, microemulsions, creams, jellies and liposomes. While basically similar in nature these formulations vary in the components and the consistency of the final product.

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Agents that enhance uptake of oligonucleotides at the cellular level may also be added to the pharmaceutical and other compositions of the present invention. For example, cationic lipids, such as lipofectin (U.S. Pat. No. 5,705,188), cationic glycerol derivatives, and polycationic molecules, such as polylysine (WO 97/30731), also enhance the cellular uptake of oligonucleotides.

The compositions of the present invention may additionally contain other adjunct components conventionally found in pharmaceutical compositions. Thus, for example, the compositions may contain additional, compatible, pharmaceutically-active materials such as, for example, antipruritics, astringents, local anesthetics or anti-inflammatory agents, or may contain additional materials useful in physically formulating various dosage forms of the compositions of the present invention, such as dyes, flavoring agents, preservatives, antioxidants, opacifiers, thickening agents and stabilizers. However, such materials, when added, should not unduly interfere with the biological activities of the components of the compositions of the present invention. The formulations can be sterilized and, if desired, mixed with auxiliary agents, e.g., lubricants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure, buffers, colorings, flavorings and/or aromatic substances and the like which do not deleteriously interact with the nucleic acid(s) of the formulation.

Certain embodiments of the invention provide pharmaceutical compositions containing (a) one or more antisense compounds and (b) one or more other chemotherapeutic agents that function by a non-antisense mechanism. Examples of such chemotherapeutic agents include, but are not limited to, anticancer drugs such as daunorubicin, dactinomycin, doxorubicin, bleomycin, mitomycin, nitrogen mustard, chlorambucil, melphalan, cyclophosphamide, 6-mercaptopurine, 6-thioguanine, cytarabine (CA), 5-fluorouracil (5-FU), floxuridine (5-FUdR), methotrexate (MTX), colchicine, vincristine, vinblastine, etoposide, teniposide, cisplatin and diethylstilbestrol (DES). Anti-inflammatory drugs, including but not limited to nonsteroidal anti-inflammatory drugs and corticosteroids, and antiviral drugs, including but not limited to ribivirin, vidarabine, acyclovir and ganciclovir, may also be combined in compositions of the invention. Other non-antisense chemotherapeutic agents are also within the scope of this invention. Two or more combined compounds may be used together or sequentially.

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Dosing is dependent on severity and responsiveness of the disease state to be treated, with the course of treatment lasting from several days to several months, or until a cure is effected or a diminution of the disease state is achieved. Optimal dosing schedules can be calculated from measurements of drug accumulation in the body of the patient. The administering physician can easily determine optimum dosages, dosing methodologies and repetition rates. Optimum dosages may vary depending on the relative potency of individual oligonucleotides, and can generally be estimated based on EC50s found to be effective in in vitro and in vivo animal models or based on the examples described herein. In general, dosage is from 0.01 µg to 100 g per kg of body weight, and may be given once or more daily, weekly, monthly or yearly. The treating physician can estimate repetition rates for dosing based on measured residence times and concentrations of the drug in bodily fluids or tissues. Following successful treatment, it may be desirable to have the subject undergo maintenance therapy to prevent the recurrence of the disease state, wherein the oligonucleotide is administered in maintenance doses, ranging from 0.01 µg to 100 g per kg of body weight, once or more daily, to once every 20 years.

V. Transgenic Animals Expressing Cancer Marker Genes

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The present invention contemplates the generation of transgenic animals comprising an exogenous cancer marker gene of the present invention or mutants and variants thereof (e.g., truncations or single nucleotide polymorphisms). In preferred embodiments, the transgenic animal displays an altered phenotype (e.g., increased or decreased presence of markers) as compared to wild-type animals. Methods for analyzing the presence or absence of such phenotypes include but are not limited to, those disclosed herein. In some preferred embodiments, the transgenic animals further display an increased or decreased growth of tumors or evidence of cancer.

The transgenic animals of the present invention find use in drug (e.g., cancer therapy) screens. In some embodiments, test compounds (e.g., a drug that is suspected of being useful to treat cancer) and control compounds (e.g., a placebo) are administered to the transgenic animals and the control animals and the effects evaluated.

The transgenic animals can be generated via a variety of methods. In some embodiments, embryonal cells at various developmental stages are used to introduce transgenes for the production of transgenic animals. Different methods are used depending on the stage of development of the embryonal cell. The zygote is the best target for micro-injection. In the mouse, the male pronucleus reaches the size of approximately 20 micrometers in diameter that allows reproducible injection of 1-2 picoliters (pl) of DNA solution. The use of zygotes as a target for gene transfer has a major advantage in that in most cases the injected DNA will be incorporated into the host genome before the first cleavage (Brinster *et al.*, Proc. Natl. Acad. Sci. USA 82:4438-4442 [1985]). As a consequence, all cells of the transgenic non-human animal will carry the incorporated transgene. This will in general also be reflected in the efficient transmission of the transgene to offspring of the founder since 50% of the germ cells will harbor the transgene. U.S. Patent No. 4,873,191 describes a method for the micro-injection of zygotes; the disclosure of this patent is incorporated herein in its entirety.

In other embodiments, retroviral infection is used to introduce transgenes into a non-human animal. In some embodiments, the retroviral vector is utilized to transfect oocytes by injecting the retroviral vector into the perivitelline space of the oocyte (U.S. Pat. No. 6,080,912, incorporated herein by reference). In other embodiments, the

developing non-human embryo can be cultured in vitro to the blastocyst stage. During this time, the blastomeres can be targets for retroviral infection (Janenich, Proc. Natl. Acad. Sci. USA 73:1260 [1976]). Efficient infection of the blastomeres is obtained by enzymatic treatment to remove the zona pellucida (Hogan et al., in Manipulating the Mouse Embryo, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. [1986]). 5 The viral vector system used to introduce the transgene is typically a replication-defective retrovirus carrying the transgene (Jahner et al., Proc. Natl. Acad Sci. USA 82:6927 [1985]). Transfection is easily and efficiently obtained by culturing the blastomeres on a monolayer of virus-producing cells (Stewart, et al., EMBO J., 6:383 [1987]). Alternatively, infection can be performed at a later stage. Virus or virus-producing cells 10 can be injected into the blastocoele (Jahner et al., Nature 298:623 [1982]). Most of the founders will be mosaic for the transgene since incorporation occurs only in a subset of cells that form the transgenic animal. Further, the founder may contain various retroviral insertions of the transgene at different positions in the genome that generally will segregate in the offspring. In addition, it is also possible to introduce transgenes into the 15 germline, albeit with low efficiency, by intrauterine retroviral infection of the midgestation embryo (Jahner et al., supra [1982]). Additional means of using retroviruses or retroviral vectors to create transgenic animals known to the art involve the micro-injection of retroviral particles or mitomycin C-treated cells producing retrovirus into the perivitelline space of fertilized eggs or early embryos (PCT International . 20 Application WO 90/08832 [1990], and Haskell and Bowen, Mol. Reprod. Dev., 40:386 [1995]).

In other embodiments, the transgene is introduced into embryonic stem cells and the transfected stem cells are utilized to form an embryo. ES cells are obtained by culturing pre-implantation embryos in vitro under appropriate conditions (Evans et al., Nature 292:154 [1981]; Bradley et al., Nature 309:255 [1984]; Gossler et al., Proc. Acad. Sci. USA 83:9065 [1986]; and Robertson et al., Nature 322:445 [1986]). Transgenes can be efficiently introduced into the ES cells by DNA transfection by a variety of methods known to the art including calcium phosphate co-precipitation, protoplast or spheroplast fusion, lipofection and DEAE-dextran-mediated transfection. Transgenes may also be introduced into ES cells by retrovirus-mediated transduction or by micro-injection. Such

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transfected ES cells can thereafter colonize an embryo following their introduction into the blastocoel of a blastocyst-stage embryo and contribute to the germ line of the resulting chimeric animal (for review, *See*, Jaenisch, Science 240:1468 [1988]). Prior to the introduction of transfected ES cells into the blastocoel, the transfected ES cells may be subjected to various selection protocols to enrich for ES cells which have integrated the transgene assuming that the transgene provides a means for such selection.

Alternatively, the polymerase chain reaction may be used to screen for ES cells that have integrated the transgene. This technique obviates the need for growth of the transfected ES cells under appropriate selective conditions prior to transfer into the blastocoel.

In still other embodiments, homologous recombination is utilized to knock-out gene function or create deletion mutants (e.g., truncation mutants). Methods for homologous recombination are described in U.S. Pat. No. 5,614,396, incorporated herein by reference.

15 EXPERIMENTAL

The following examples are provided in order to demonstrate and further illustrate certain preferred embodiments and aspects of the present invention and are not to be construed as limiting the scope thereof.

In the experimental disclosure which follows, the following abbreviations apply: N (normal); M (molar); mM (millimolar); μ M (micromolar); mol (moles); mmol (millimoles); μ mol (micromoles); nmol (nanomoles); pmol (picomoles); g (grams); mg (milligrams); μ g (micrograms); ng (nanograms); l or L (liters); ml (milliliters); μ l (microliters); cm (centimeters); mm (millimeters); μ m (micrometers); nm (nanometers); and C (degrees Centigrade).

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Example 1

Preparation of Total RNA and Reference Pools

The prostate surgical specimens were obtained from The University of Michigan Specialized Research Program in Prostate Cancer (S.P.O.R.E.) Tumor Bank with Institutional Review Board approval. Tumors samples were derived from patients with clinically localized and advanced hormone refractory prostate cancer. Table 1 shows the

samples used in the present studies. All patients were operated on between 1993 and 1998 for clinically localized prostate cancer as determined by preoperative PSA, digital-rectal examination, and prostate needle biopsy. In addition, a subset of patients received bone and CAT scans to evaluate the possibility of metastatic spread. All patients received radical prostatectomy as a monotherapy (i.e., no hormonal or radiation therapy). The advanced prostate tumors were collected from a series of 12 rapid autopsies performed at the University of Michigan on men who died of hormone refractory prostate cancer. In brief, the majority of these patients had either widely metastatic prostate cancer which was treated with hormonal therapy followed by chemotherapy, or patients who presented with clinically localized disease which progressed and were then treated with both hormonal and chemotherapy. The majority of cases had multiple metastatic lesions to numerous sites. All autopsies were performed within 4-6 hours after death. The clinical and pathologic findings of these cases have recently been reported (Rubin et al., Clin. Cancer Res., 6:1038 [2000]). All samples used for the tissue microarray study were fixed in 10% formalin.

Tissues were homogenized using a polytron homogenizer (Brinkman Instruments) in Trizol (Gibco BRL) and the total RNA was isolated according to the standard Trizol protocol. The total RNA obtained was further subjected to an additional round of phenol chloroform extraction, precipitated and resuspended in RNAse free water. Total RNA was quantitated by spectrophotmetric (260/280nm) absorbance and integrity judged by denaturing- formaldehyde agarose gel electrophoresis. Total RNA from four normal tissues was combined in equal concentrations to obtain the reference pool. The human prostate total RNA used in the commercial reference pool was obtained from Clontech, Inc.

Table 1			
Prostate Samples			
ID	PSA level	Tissue	Gleason Score

BPH-201	6.2	Prostate	NA ;
BPH-202	3.9	Prostate	NA
BPH-203	3.9	Prostate	NA
BPH-204	4.6	Prostate	NA .
BPH-205	4.6	Prostate	NA
BPH-206	4.6	Prostate	NA
BPH-207	4.8	Prostate	NA
BPH-208	13.6	Prostate	NA
BPH-209	9.8	Prostate	NA
BPH-210	4.6	Prostate	NA
BPH-211	2.6	Prostate	NA
BPH-212	7.1	Prostate	NA
BPH-214		Prostate	NA
BPH-215	5.4	Prostate	NA
Prostatitis	9.8	Prostate	NA
NAP-101	4.6	Prostate	NA
NAP-102	9.8	Prostate	NA

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NAP-104	7	Prostate	NA
NAP-105	0.09	Prostate	NA
NAP-107	4.7	Prostate	NA
PCA-401	5.2	Prostate	4+4
PCA-402	22	Prostate	4+3
PCA-403	4.7	Prostate	3+3
PCA-404	8.5	Prostate	3+3
PCA-405	4.6	Prostate	3+3
PCA-406	7.8	Prostate	3+3
PCA-407	7.8	Prostate	3+3
PCA-408	5.4	Prostate	3+3
PCA-409	7	Prostate	3+3
PCA-410	44.6	Prostate	4+4
PCA-414		Prostate	3+4
PCA-416	24.1	Prostate	4+4
PCA-417	12.4	Prostate	4+4
PCA-420		Prostate	3+3

WO 03/012067

PCA-421	13.6	Prostate	3+4
MET-301		Lung	NA
MET-302		Liver	NA
MET-303		Liver	NA
MET-304		Stomach	NA
MET-305		Adrenal	NA
MET-306		Prostate	NA
MET-307		Lymph Node	NA
MET-308		Lymph Node	NA
MET-309		Lymph Node	NA
MET-310		Liver	NA
MET-311		Soft tissue	NA .
MET-312		Liver	NA
MET-313		Soft tissue	NA
MET-314		Soft tissue	NA .
MET-315 .		Soft tissue	NA :
MET-316		Soft tissue	NA

MET-317	Liver	NA
MET-318	bone	NA .
MET-319	bone	NA
MET-320	bone	NA

Table 1. Samples employed in the study. Designating PSA level in ng/mL, Organ sources and Gleason scores. Normal adjacent prostate (NAP), Benign prostatic hyperplasia (BPH), Localized prostate cancer (PCA) and Hormone refractory metastatic prostate cancer (MET). NA refers to "not applicable".

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Example 2

Microarray Analysis

This example describes the use of microarray analysis to identify genes that demonstrate an altered level of expression in cancerous or benign prostate tissues.

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A. Experimental Methods

Microarray analysis of gene expression was conducted essentially as described by the Brown and Derisi Labs (available at the Internet site www.microarrays.org). The sequence-verified cDNA clones on the human cDNA microarray are available from the web site of Research Genetics. Based on the latest Unigene build, the 10K human cDNA microarray used covers approximately 5520 known, named genes and 4464 ESTs. All chips have various control elements that include human, rat, and yeast genomic DNAs, SSC, yeast genes and "housekeeping genes," among others. In addition, 500 cancer- and apoptosis- related cDNAs from Research Genetics were used to serve as independent controls for clone tracking and function as duplicates for quality control. Three metastatic prostate cancer cell lines: DU-145, LnCAP, and PC3 were also profiled for gene expression.

Fluorescently labeled (Cy5) cDNA was prepared from total RNA from each experimental sample. The two reference samples used in this study were labeled using a second distinguishable fluorescent dye (Cy3) and included a pool of normal adjacent prostate tissue (NAP) from four patients (distinct from those used in the experimental samples) and a commercial pool of normal prostate tissues (CP). In addition to minimizing patient-to-patient variation, comparisons against pools of normal prostate tissue facilitate the discovery of genes that molecularly distinguish prostate neoplasms. The two reference pools are different in that one is comprised of normal adjacent prostate tissue, which may be influenced by paracrine effects mediated by PCA, and furthermore is exposed to the same environmental and genetic factors as the adjacent PCA. By contrast, the CP pool is derived from 19 individuals with no known prostate pathology and also represents a renewable commercially available reference resource.

Purified PCR products, generated using the clone inserts as template, were spotted onto poly-L- lysine coated microscope slides using an Omnigrid robotic arrayer (GeneMachines, CA) equipped with quill-type pins (Majer Scientific, AZ). One full print run generated approximately 100 DNA microarrays. Protocols for printing and post-processing of arrays are well known in the art.

B. Data analysis

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Primary analysis was done using the Genepix software package. Images of scanned microarrays were gridded and linked to a gene print list. Initially, data was viewed as a scatter plot of Cy3 versus Cy5 intensities. Cy3 to Cy5 ratios were determined for the individual genes along with various other quality control parameters (e.g., intensity over local background). The Genepix software analysis package flags spots as absent based on spot characteristics (refer to the web site of Axon Instruments, Inc.). Bad spots or areas of the array with obvious defects were manually flagged. Spots with small diameters (< 50 microns) and spots with low signal strengths <350 fluorescence intensity units over local background in the more intense channel were discarded. Flagged spots were not included in subsequent analyses. Data were scaled such that the average median ratio value for all spots was normalized to 1.0 per array.

These files were then imported into a Microsoft Access database. The data for the required experiments were extracted from the database in a single table format with each row representing an array element, each column a hybridization and each cell the observed normalized median of ratios for the array element of the appropriate hybridization. Prior to clustering, the normalized median of ratio values of the genes were log base 2 transformed and filtered for presence across arrays and selected for expression levels and patterns depending on the experimental set as stated. Average linkage hierarchial clustering of an uncentered Pearson correlation similarity matrix was applied using the program Cluster (Eisen et al., PNAS 95:14863 [1998]), and the results were analyzed and figures generated with the program TreeView. TreeView and Cluster are available from Michael Eisen's lab at the Lawrence Berkeley National Lab.

C. Results

Over forty 10K human cDNA microarrays were used to assess gene expression in four clinical states of prostate-derived tissues in relation to two distinct reference pools of normal specimens. Figure 1 provides an overview of the variation in gene expression across the different tissue specimens analyzed. A hierarchical clustering algorithm was employed to group genes and experimental samples based on similarities of gene expression patterns over all the genes and samples tested, respectively.

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1. Expression Dendrograms

Relationships between the experimental samples are summarized as dendrograms (Figure 1a), in which the pattern and length of the branches reflect the relatedness of the samples. Figure 1a shows dendrograms that reveal the variation in gene expression pattern between experimental samples with the two references employed. Individual samples in each group are indicated by the branches of the same color whereby METs are in dark blue, localized PCAs in orange, NAPs in light blue, BPHs in gray, and cell lines in pink. Asterisk (*) indicates a sample that was initially documented as BPH, but was later confirmed to have 5% cancer tissue. The details of metastatic samples used in this study are as follows: MET 301, from Lung; MET 302 and 303 from liver; MET 304, from stomach; MET 305 from adrenal gland; MET 306 from prostate; and MET 307 was

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from lymph node. Hierarchical clustering of the data identified distinct patterns of gene expression between the various groups analyzed. Each row represents a single gene with 1520 genes depicted in b, and 1006 genes depicted in c. The results represent the ratio of hybridization of fluorescent cDNA probes prepared from each experimental mRNA to the respective reference pools. These ratios are a measure of relative gene expression in each experimental sample and are depicted according to the color scale at the bottom left. Red and green colors in the matrix represent genes that are up- and down-regulated, respectively, relative to the reference pool employed. Black lines in the matrix represent transcript levels that are unchanged, while gray lines signify technically inadequate or missing data (NP, not present). Color saturation reflects the magnitude of the ratio relative to the median for each set of samples.

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Figure 1b shows a cluster diagram of the various sample groups compared against normal adjacent prostate pool as reference. Data obtained in the expression profiling experiments included CBCR-t Index number, Clone, ID, Unigene Cluster Ids, Accession ID, NID, gene symbol, and name fields for each gene used in the array. The name field 15 · contains genes having partial or complete homology based on homology searches. In addition, the data contains the numerical difference in expression levels compared to the reference pool for each gene. Prior to hierarchical average-linkage clustering, the data was filtered for at least a 2-fold change in expression ratio and ratio measurements present in 50% of the samples. By this method, 1520 genes were selected from the NAP reference data set. Indicated by vertical bars on the left (b1 to b6) of Figure 1b are regions identified with characteristic gene expression signatures. Clusters b1 and b5 show genes up-regulated in localized PCA but not in metastatic PCA. Clusters b2 and b4 highlight genes down-regulated in metastatic PCA and the cell lines DU145 and LnCAP. Cluster b3 identifies genes down-regulated in both localized PCA and metastatic PCA. Cluster b6 highlights genes that are primarily up-regulated in metastatic PCA alone. Portions of Clusters b4 and b6 are shown enlarged with selected genes shown using Human genome organization (HUGO) gene nomenclature.

Figure 1c shows a cluster diagram of the various sample groups compared against the commercial prostate pool reference. Prior to hierarchical average-linkage clustering, the data was filtered for at least a 3-fold change in expression ratio and ratio

measurements present in 75% of the samples resulting in a total of 1006 genes. Regions with distinct patterns (c1-c6) are indicated by vertical bars to the right of Figure 1c. Cluster c1 depicts genes down-regulated in both localized PCA and metastatic PCA. Cluster c2 represents genes down-regulated only in metastatic PCA. Cluster c3 shows genes that are highly represented in the commercial pool. Cluster c4 highlights genes that are up-regulated in localized PCA and in metastatic PCA. Cluster c5 represent genes with a low representation in the commercial pool. Cluster c6, represents genes that are down-regulated in metastatic PCA but are up-regulated in all other samples used.

Benign conditions of the prostate such as BPH and NAP cluster separately from malignant PCA cell lines or tissues, regardless of the reference pool used. Within the PCA cluster, it is also evident that metastatic PCA and clinically localized PCA formed distinct subgroups. Similarly, in the "benign" grouping, BPH tended to distinguish itself from NAP. Interestingly, one of the "BPH" samples initially clustered with the localized PCA group. Upon further histopathologic review, however, it was discovered that this sample contained a small focus of neoplastic tissue (~5%), thus accounting for its initial misclassification (now designated PCA+BPH in Figure 1a).

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Eisen matrix formats (Eisen et al., supra) of the variation in gene expression are also presented (Figure 1 b and 1c). With a global perspective of the data, it is apparent that metastatic PCA dominates the analysis and has the greatest variation in gene expression of the samples tested. Bars on the left or right of each matrix represent clusters of coordinately expressed genes highlighting interrelationships between specimens. For example, Clusters b3 and c1 represent genes down-regulated in both localized and metastatic PCA (Figures 1b and 1c). By contrast, Clusters b6 and b4 highlight genes that are specifically up- and down- regulated in metastatic PCA, respectively (Figure 1b). IGFBP-5, DAN1, FAT tumor suppressor and RAB5A are examples of genes that are down-regulated specifically in metastatic PCA and also have a proposed role in oncogenesis ("magnified" regions, Figure 1b). Similarly, cancer-related genes that are up-regulated in metastatic PCA include MTA-1 (metastasis-associated 1), MYBL2, and FLS353 (preferentially expressed in colorectal cancer). Many genes in this "met-specific" cluster are shared by both the metastatic PCA tissue and the two PCA cell lines DU145 and LnCAP.

Data was also obtained from the expression profiling of additional prostate tissue specimens profiled against a commercial prostate reference pool (CPP). A total of 53 prostate specimens were profiled against the commercial pool. They include 4 normal adjacent prostate tissue (NAP), 14 benign prostatic hyperplasia (BPH), 1 prostatitis, 14 localized prostate cancer (PCA) and 20 hormone refractory metastatic PCA (MET). Prior to hierarchial average-linkage clustering, the data was filtered for at least 3-fold change in Cy5/Cy3 ratios and measurements present in 75% of the samples. By this method 1325 genes were selected. The data expands on Figure 1c with an additional 40 samples, which include all from Figure 1b, and also includes 28 additional prostate specimens.

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2. Focused Clusters

Data was next assessed by examining functional groups of known, named genes. Cancer-related functional clusters were arbitrarily defined including cell growth/cell death, cell adhesion, anti-protease/protease, free radical scavengers, inflammation/immunity, phosphatase/kinase, transcription, and miscellaneous (Figures 2 and 6).

One of several available methods of gene selection was used to create a more limited set of genes for future exploration. In one method, t-statistics (based on MET/PCA vs. benign) were computed for each gene. The cell line samples were excluded from the analysis. Also, genes and ESTs that had data missing from 20% of samples were excluded from analysis. The t-statistics were ranked in two ways. First, they were ranked by absolute magnitude, which takes into account the inter-sample variability in expression ratios. Second, they were ranked by the magnitude of the numerator of the test statistic, which is based on the biological difference in expression ratios and designated as "effect size" (for MET/PCA vs. benign). A scatterplot of the genes with the 200 largest effect sizes and 200 largest t-statistics was then plotted (See Figure 7). Figure 7 shows gene selection based on computed t-statistics for each gene. Two groups were used in the analysis: PCA/MET and benign (NAP/BPH). Figure 7a shows analysis of NAP pool data. Figure 7b shows analysis of CP pool data. Selected genes are named and 200 genes for each data set are shown. Gene selection based on

each method is shown. Selected gene names or symbols (as specified by Human genome organization (HUGO) gene nomenclature) are shown.

Genes that made both lists were also looked at separately in order to identify potential candidate genes. Implementing this methodology on both reference pool data sets (NAP and CP) yielded genes that included hepsin, pim-1, IM/ENIGMA, TIMP2, hevin, rig, and thrombospondin-1, among others. Several genes identified using gene selection methods are described in more detail in the context of "functional" clusters described in Figure 2.

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Figure 2 shows the differential expression of functional clusters of select genes in prostate cancer. Gene names or symbols (as specified by Human genome organization (HUGO) gene nomenclature) are shown. The same convention for representing changes in transcript levels was used as in Figure 1. The sample order from Figure 1 was preserved for clarity.

Figure 8 shows a focused cluster of PCA-related genes. The same convention for representing changes in transcript levels was used as in Figure 1. This cluster of 231 genes was generated by selecting for a 3.5-fold variation in at least 2 of any class, and ratio measurements present in 75% of the samples. Classes included: PCA vs. NAP, MET vs. NAP, PCA vs. CP and MET vs. CP.

The reliability of the hierarchical clustering results was assessed using three separate methods: that of Calinski and Harabasz (1974), Hartigan (1975) and Krzanowski and Lai (1985). The number of "stable" clusters estimated by all these methods is two. In the CP pool data set, that would elicit a valid benign cluster (NAP and BPH) and a malignant cluster (PCA and MET).

Many of the genes identified in these "focused" clusters have been implicated directly or indirectly as cancer biomarkers or mediators of carcinogenesis. Several have been shown to be dysregulated in PCA. For example, the tumor suppressor gene PTEN was down-regulated, while the proto-oncogene myc was up-regulated in the microarray analysis of PCA (Figure 2) (Abate-Shen and Shen, *supra*). Likewise, decreased expression of E-cadherin and increased expression of fatty acid synthase, both of which have been shown to be dysregulated in PCA were observed (Tomita *et al.*, Cancer Res., 60:3650 [2000] and Shurbaji *et al.*, Hum. Pathol., 27:917 [1996]). In addition to

uncharacterized expressed sequence tags (ESTs), there are numerous genes that were identified by the screen but not previously known to be associated with PCA. It is contemplated that they find use as cancer markers.

Exemplary nucleic acid sequences for some of the genes identified in focused clusters are shown in Figures 9 and 10. The present invention is not limited to the particular nucleic acid sequences described in Figures 9 and 10. One skilled in the art recognizes that additional variants, homologs, and mutants of the described sequences find use in the practice of the present invention.

3. Comparison Between NAP and CP pools

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A direct comparison between the NAP and CP pool was also made and notable gene expression differences were readily apparent. Figure 5 shows a comparison between the NAP and CP pools. The same convention for representing changes in transcript levels was used as in Figure 1. The cluster was obtained by selecting for genes with at least a 2.5-fold variation in any two of the samples of each class, namely the normal tissues versus the NAP pool and normal tissue versus the CP pool at a 50% filter. Of the genes analyzed 59 were selected with this criteria. Genes that were found to be up-regulated in the NAP pool in comparison with CP pool included connective tissue growth factor, EGR-1 (Early Growth Response 1), matrilysin (MMP7), CFLAR/I-FLICE (caspase 8 and FADD-like apoptosis regulator), lumican, serum glucocorticoid regulated kinase, lens epithelium derived growth factor, PAI1 (plasminogen activator inhibitor type I), JUN and FOS B, among others. Vascular endothelial growth factor (VEGF), growth arrest specific (GAS1), cholecystokinin (CCK), amiloride binding protein (ABP1) were among the down-regulated genes in the normal adjacent prostate pool when compared to the commercial pool. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that the gene expression differences between normal prostate adjacent to PCA (NAP) and normal prostate tissue from individuals without prostate pathology (CP) may be attributable to a "field effect" induced by PCA itself.

Example 3

Northern Blot Analysis

Thirty micrograms of total RNA was resolved by denaturing formaldehyde agarose gel and transferred onto Hybond membrane (Amersham) by a capillary transfer set up. Hybridizations were performed by the method described by Church and Gilbert, 1984. Signal was visualized and quantitated by phosphorimager. For relative fold estimation, the ratio between the signals obtained from hepsin and GAPDH probes was calculated.

Selected genes identified by microarray analysis were corroborated by Northern
analysis. For example, hevin, 4 1/2 LIM domain protein and gelsolin were shown to be
3.2-, 3.2- and 1.9- fold down-regulated, respectively by microarray and 8.8-, 4.5-, and
3.5- fold down-regulated by Northern analysis. Similarly, hepsin was
4.3-fold up-regulated by microarray and 11.3- fold up-regulated by Northern analysis
(Figure 3a). As hepsin is a cell-surface serine protease with transcript expression
precisely restricted to localized and metastatic PCA, its expression was examined in more
detail at the protein level (See Example 4 below).

Example 4

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Tissue Analysis

This example describes the analysis of protein expression in normal and cancerous prostate tissues.

A. Tissue microarray construction.

Kononen et al. have described a method for evaluating tumor tissues in large

numbers on a single glass slide (Kononen et al., Nat. Med., 4:844 [1998]). These highdensity tissue microarrays allow for analysis of up to 1,000 tissue samples on a single
slide. These slides can be evaluated by routine light microscopy on hematoxylin and
eosin (H&E) prepared and immunohistochemically stained slides. Thus, candidate
cancer biomarkers, identified by gene expression methodologies, can be evaluated at the
protein level over a large number of clinically stratified tumor specimens.

Prostate tissues used in microarray analysis included 4 BPH, 8 NAP, 1 commercial pool of normal prostate tissue (from 19 individuals), 1 prostatitis, 11 localized PCA, and 7 metastatic PCA specimens. High-density tissue microarrays (TMA) were assembled using a manual tissue puncher/array (Beecher Instruments, Silver Springs, MD) as previously described (Kononen et al., Nat. Med., 4:844 [1998]; Perrone et al., J. Natl. Cancer Inst., 92:937 [2000]). The instrument consists of thin-walled stainless steel needles with an inner diameter of approximately 600 µm and stylet used to transfer and empty the needle contents. The assembly is held in an X-Y position guide that is manually adjusted by digital micrometers. Small biopsies are retrieved from selected regions of donor tissue and are precisely arrayed in a new paraffin block. Tissue cores were 0.6 mm in diameter and ranged in length from 1.0 mm to 3.0 mm depending on the depth of tissue in the donor block. Multiple replicate core samples of normal, HGPIN, and PCA were acquired from each tissue block of each case. Cores were inserted into a 45 x 20 x 12 mm recipient bock and spaced at a distance of 0.8 mm apart. Prostate tumor grading was performed using the system described by Gleason (Gleason, Cancer Chemother Rep., 50:125 [1966]). Pathologic stages for the radical prostatectomies were determined using the TNM staging system (Schroder et al., Prostate Suppl., 4:129 [1992]). Surgical margins were assessed separately and are not included in tumor staging.

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B. Immunohistochemistry

TMA sections were cut at five- micron thick intervals for immunohistochemistry. Initial sections were stained for hematoxylin and eosin to verify histology. TMA slides prepared from formalin- fixed paraffin embedded tissue were heated for 0.5 - 1 hours at 60° centigrade. All slides were placed in 10 millimolar citrate buffer (pH 6.0) and microwaved for 5 minutes. Standard biotin-avidin complex immunohistochemistry was performed. The affinity purified polyclonal Rabbit antibody against hHepsin was used at a 1:40 dilution (original concentration 0.2 mg/ml) for this study. Immunostaining intensity was scored by a dedicated genitourinary pathologist as absent, weak, moderate, or strong. Scoring was performed using a telepathology system in a blinded fashion without knowledge of overall Gleason score (e.g., tumor grade), tumor size, or clinical

outcome (Perrone et al., supra). A total of 738 tissue samples from benign (n=205), high-grade PIN (n=38), localized prostate cancer (n=335) and hormone refractory prostate cancer (n=160) were examined.

Similarly, pim-1 was analyzed using two TMA blocks from a total of 810 PCA samples from 135 patients. Six PCA samples were evaluated from each case and a median score was calculated. In addition, a small number of samples with benign prostatic tissues (e.g., benign epithelium and atrophy) and HG-PIN were examined. Immunohistochemistry was performed as above, using a rabbit polyclonal antibody against the N-terminus of pim-1 (Santa Cruz Biotechnology) at a 1:100 dilution. Pim-1 demonstrated cytoplasmic staining and was graded as either negative, weak, moderate, or strong. All samples were reviewed blinded with respect to all related pathology and clinical data.

C. Statistical methods

A nonparametric ANOVA test (Mann-Whitney [two categories]) was employed to evaluate whether the prostate samples expressed hepsin and pim-1 at different levels based on various parameters (tissue type, Gleason score, and tumor size). Kaplan-Meier analysis was used to estimate the cumulative percentage of PSA free progression ("survival"). The log-rank test was employed to assess the differences in disease free progression hepsin immunostaining. Cox proportional- hazard regression was used for multivariate analysis. Commercial software from SPSS (Chicago, IL) was used for this study.

D. Results

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1. Hepsin

Microarrays used in this study are shown in Figure 3b. Over 700 benign and malignant prostate tissues were immunohistochemically profiled on tissue microarrays (Figure 3c-e) using an affinity-purified hepsin-peptide antibody (Tsuji *et al.*, J. Biol. Chem., 266:16948 [1991]). Figure 3 shows the overexpression of Hepsin, a transmembrane serine protease, in prostate cancer. Figure 3a shows a Northern blot

analysis of human hepsin (top) and normalization with GAPDH (bottom). NAT indicates normal adjacent prostate tissue and PCA indicates prostate cancer. Figure 3b shows tissue microarrays used for hepsin analysis. Staining was done with hemotoxylin and eosin to verify histology.

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Immunohistochemical stains demonstrated absent or weak staining of benign prostate (c1), strong staining in localized prostate cancer (c2-6), and strong staining in a high-grade prostate tumor (magnification 100X was used for all images, samples measure 0.6 mm in diameter). Benign prostate glands demonstrate weak expression in the secretory, luminal cells and strong basal cell staining. In areas where prostate cancer and benign prostate glands are seen, significant hepsin staining differences are observed. Infiltrating prostate cancers (d3-4) demonstrate strong hepsin protein expression. Magnification for all images was 400X. Figure 3c shows a histogram of hepsin protein expression by tissue type. Benign prostate hyperplasia (BPH). High-grade intraepithelial neoplasia (HG-PIN). Localized prostate cancer (PCA). Hormone-refractory prostate cancer (MET). Relative strength of hepsin staining was qualitatively assessed and categorized. Percentage of hepsin staining per category is shown on the y-axis. Figure 3d shows Kaplan Meier Analysis. PSA-free survival was stratified by level of hepsin protein expression into two categories absent/low expression (circles) versus moderate/strong expression (squares).

Internal controls showed that liver tissue, as previously described, stained strongly for hepsin. Overall, hepsin exhibited predominantly membrane staining and was preferentially expressed in neoplastic prostate over benign prostate (Mann-Whitney test, p<0.0001). Importantly, the precursor lesion of PCA, HG-PIN, had the strongest expression of hepsin, and almost never had absent staining (Mann-Whitney, p<0.0001). Most cases of low or absent hepsin staining were seen in benign prostate specimens. In addition, hormone refractory metastatic cancers were intermediate in staining intensity between localized prostate tumors and benign prostate.

Men who develop elevated PSA levels following radical prostatectomy are at a high risk to develop distant metastases and die due to prostate cancer (Pound et al., JAMA, 281:1591 [1999]. Therefore, to assess the usefulness of hepsin as a potential PCA biomarker, PSA failure was defined as a PSA elevation of greater than 0.2 ng/ml

following radical prostatectomy. Analysis was performed on 334 localized prostate cancer samples treating each as an independent sample. PSA elevation following radical prostatectomy was significantly associated with absent and low hepsin immunostaining with a 28% (46/119 samples) PSA failure rate, in contrast to 17% (28/141 samples) PSA failure rate for tumors with moderate to strong hepsin expression (Figure 3d, Log Rank test P=0.03). Multivariate analysis was performed to examine if these results were independent of Gleason score, a well-established histologic grading system for PCA (Gleason, Hum. Pathol., 23:273 [1992]). Based on the results from fitting a Cox proportional hazards model, there is an association of weak or absent hepsin protein expression in PCA with increased risk of PSA elevation following prostatectomy, similar to high Gleason score (corresponding risk ratios were 2.9 (p=0.0004) and 1.65 (p=0.037), respectively). Weak or absent hepsin expression was also associated with large prostate cancers; the median tumor dimension for prostate tumors with moderate to strong expression was 1.3 cm but 1.5 cm for tumors with absent or weak staining (Mann-Whitney Rank test, P=0.043). Taken together, hepsin protein expression in PCA correlated inversely with measures of patient prognosis.

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Hepsin is a 51 kDa transmembrane protein with highest expression in the liver, and like PSA, is a serine protease (Kurachi et al., Methods Enzymol., 244:100 [1994]). The protease domain of hepsin has access to the extracellular space and can potentially activate other proteases or degrade components of extracellular matrix. The function of hepsin is poorly understood. It has been proposed to have a role in controlling cell growth (Torres-Rosado et al., PNAS, 90:7181 [1993], cell morphology, and activating the extrinsic coagulation pathway on the cell surface, leading to thrombin formation (Kazama et al., J. Biol. Chem., 270:66 [1995]). Additionally, hepsin mRNA levels have been shown to be elevated in ovarian carcinomas (Tanimoto et al., Cancer Res., 57:2884 [1997]). The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that the high expression of hepsin in HG-PIN, and not benign prostate, suggests that hepsin plays a role in the establishment of PIN or in the transition from HG-PIN to carcinoma. Subsequent decreases in hepsin expression seen in large localized cancers and hormone-refractory cancers suggest a decreased requirement

of this protease in later stages of PCA. Alternatively, patients with advanced PCA often develop disseminated intravascular coagulation (DIC) (Riddell *et al.*, J. Nucl. Med., 37:401 [1996]) whereby hepsin may play an important role.

2. pim-1

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Tumorigenic growth of the prostate depends on the evasion of normal homeostatic control mechanisms, where cell proliferation exceeds cell death (Bruckheimer and Kyprianou, Cell Tissue Res., 301: 153 [2000]). While it is well known that the oncogene myc is overexpressed in many PCAs (Buttyan et al., prostate 11:327-37 [1987]; Abate-Shen and Shen, supra), the present invention demonstrates that the proto-oncogene pim-1 kinase is similarly up-regulated (cell growth/cell death cluster, Figure 2). Previous studies suggest that the cooperative interaction between pim-1 and myc may induce lymphoid cell transformation by promoting cell cycle progression and blocking apoptosis (Shirogane, et al., Immunity 11:709 [1999]). The present analysis supports a similar co-transcriptional regulation (or gene amplification) of pim-1 and myc possibly mediating a synergistic oncogenic effect in PCA.

Pim-1 kinase protein expression in PCA was also explored using high-denisty TMAs. Figure 4 shows the overexpression of pim-1 in prostate cancer. Immunohistochemical stains demonstrated absent or weak staining of benign prostate, and strong cytoplasmic staining in localized prostate cancer. Benign prostate glands demonstrated absent or weak expression in the secretory, luminal cells. Infiltrating prostate cancers demonstrated strong pim-1 protein expression. Magnification for all images 1000X. Figure 4a shows a histogram of pim-1 protein expression by tissue type as assessed from 810 tissue microarray elements. High-grade intraepithelial neoplasia (HG-PIN). Localized prostate cancer (PCA). Relative strength of pim-1 staining is represented in the included legend. The percentage of pim-1 staining per category shown on y-axis. Figure 4b shows Kaplan-Meier analysis demonstrating that patients with PCA that have negative to weak pim-1 expression (bottom line) are at a greater risk of developing PSA-failure following prostatectomy (log rank p=0.04). PSA-free survival was stratified by level of pim-1 protein expression into two categories absent/weak expression (bottom line) versus moderate/strong expression (top line).

Pim-1 protein was found to be markedly overexpressed in PCA (Figure 4). Negative to weak pim-1 protein expression was observed in the majority of benign prostatic epithelial (97%), prostatic atrophy (73%), and high-grade PIN (82%) samples (Figure 4a). In contrast, moderate to strong pim-1 expression was observed in approximately half of the PCA samples (51%) (Figure 4a). Kaplan-Meier analysis for PSA- free survival demonstrated positive extraprostatic extension, seminal vesicle invasion, Gleason score greater than 7 and decreased pim-1 expression to be associated with a higher cumulative rate of PSA failure (Figure 4b). By univariate Cox models, it was found that Pim-1 expression is a strong predictor of PSA recurrence (hazard ratio (HR)=2.1 (95% CI 1.2-3.8, p=0.01)).

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Among the variables examined, significant predictors of PSA recurrence were Gleason score (HR=1.8 (95% CI 1.1-3.0), p=0.03), Gleason pattern 4/5 PCA (HR=3.9(95%CI 1.8-8.3), p<0.001), extraprostatic extension status (HR=2.6 (95%CI 1.6-4.2), p<0.0001), surgical margin status (HR=2.6 (95%CI 1.2-5.6), p=0.01), seminal vesicle status (HR=3.5 (95%CI 2.0-6.2), p<0.0001), the natural log of pre-operative PSA level (HR=2.5 (95%CI 1.6-3.8), p<0.001), HR=2.4, p<0.001), and maximum tumor dimension (HR=2.7 (95%CI 1.6-4.7), p<0.0001). Presence of Gleason pattern 4/5 PCA (HR=3.8 (95%CI 1.4-10.0), p<0.01), Ln(PSA) (HR=2.1 (95%CI 1.1-3.9), p=0.02), and decreased pim-1 protein expression (HR=4.5 (95%CI 1.6-15.2), p=0.01) were both found to be significant predictors of PSA recurrence by a multivariate Cox model. Thus, even more so than hepsin, decreased expression of pim-1 kinase in PCA correlated significantly with measures of poor patient outcome.

Pim-1 kinase is a proto-oncogene that is regulated by cytokine receptors (Matikainen et al., Blood 93:1980 [1999]). It was first described as a common site of proviral integration in murine retrovirus-induced T cell lymphomas (Cuypers et al., Cell 37:141 [1984]), and was previously thought to be involved exclusively in hematopoietic malignancies (Breuer et al., Nature 340:61 [1989]). Co-transcriptional regulation of pim-1 and myc was observed in the experiments described herein (Figure 2 cell growth/cell death cluster). Chronic overexpression of myc in the ventral prostate of transgenic mice induced epithelial abnormalities similar to low-grade PIN, but progression to adenocarcinoma in this model was never observed (Zhang et al., Prostate 43:278 [2000]).

The present invention is not limited to any one mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that pim-1 overexpression may potentiate myc- induced prostate carcinogenesis.

Figure 8 provides a schematic overview of representative genes differentially expressed in PCA identified by DNA microarray analysis. Genes are grouped functionally and arrows represent up- or down- regulation in metastatic hormone-refractory PCA (MET) and/or localized PCA (PCA) relative to normal prostate epithelium. See Figure 2 for gene expression levels.

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Example 5

AMACR Expression Analysis

The Example describes the analysis of the gene expression data described in Examples 1-4 above to identify AMACR as being consistently over-expressed in prostate cancer.

A. Tissue Samples

In order to examine the widest range of prostate cancer specimens, clinical samples were taken from the radical prostatectomy series at the University of Michigan and from the Rapid Autopsy Program. Both programs are part of the University of Michigan Prostate Cancer Specialized Program of Research Excellence (S.P.O.R.E.) Tissue Core.

Prostatectomy cases for the tissue microarray (TMA) outcomes array were selected from a cohort of 632 patients, who underwent radical retropubic prostatectomy at the University of Michigan as a monotherapy (i.e., no hormonal or radiation therapy) for clinically localized prostate cancer between the years of 1994 and 1998. Clinical and pathology data for all patients was acquired with approval from the Institutional Review Board at the University of Michigan. Detailed clinical, pathology, and TMA data is maintained on a secure relational database (Manley et al., Am. J. Pathol., 159:837 [2001]).

Processing of the prostate specimens began within approximately 15-20 minutes after surgical resection. The prostates were partially sampled and approximately 50% of the tissue was used for research. This protocol has been evaluated in a formal study to assure that partial sampling does not impair accurate staging and evaluation of the surgical margins (Hollenbeck et al., J. Urol., 164:1583 [2000]). Briefly, alternate sections of the prostate gland were submitted for histologic review. The remaining sections were frozen and stored in the SPORE Tissue Core. These samples were collected only from patients who had signed an IRB-approved informed consent. The samples were snap-frozen in OCT embedding media at -80°C and stored in a holding area until the pathology report was finalized. These frozen samples were not available to researchers until adequate diagnosis and staging had been performed. The samples used for cDNA expression array analysis and RT-PCR were all evaluated by the study pathologists. All samples were grossly trimmed such that greater than 95% of the sample represented the desired lesion (e.g., prostate cancer, BPH, or benign prostate). Samples with prostate cancer were also assigned a Gleason score based on the sample used for molecular analysis.

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In order to study hormone refractory prostate cancer, a Rapid Autopsy Protocol was used, which represents a valuable source of metastatic prostate tumors. Modeled after protocols developed at the University of Washington (Seattle, WA.) and Johns Hopkins University (Baltimore, MD), this program allows men with advanced prostate cancer to consent to an autopsy immediately after death. To date, 23 complete autopsies have been performed with a median time of 2 hours from death to autopsy. This procedure has previously been described in detail (Rubin et al., Clin. Cancer Res., 6:1038 [2000]). In brief, patients diagnosed with hormone refractory prostate cancer were asked to take part in a posthumous tissue donor program. The objectives and procedures for tissue donation were explained to the patient. Having agreed to participate in this IRB-approved tumor donor program, permission for autopsy is obtained before the death, with consent provided by the patient, or by next of kin. Hormone refractory primary and metastatic prostate cancer samples were collected using liquid nitrogen. Mirrored samples from the same lesion were placed in 10% buffered formalin. The fixed samples were embedded in paraffin and used for the development of TMAs. As with the

prostatectomy samples, the study pathologist reviewed the glass slides, circled areas of viable prostate cancer, while avoiding areas of necrosis, and used these slides as a template for TMA construction.

B. Pathology and Evaluation

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Prostates were inked before the assessment of surgical margins. Surgical margins from the apex and base were cut perpendicular to the prostatic urethral axis. The seminal vesicles were cut perpendicular to their entry into the prostate gland and submitted as the seminal vesicle margin. The prostates for this study were all partially embedded, taking alternate full sections from the apex, mid, and base. Detailed prostatectomy pathology reports included the presence or absence of surgical margin involvement by tumor (surgical margin status), the presence of extraprostatic extension, and seminal vesicle invasion. Tumors were staged using the TNM system, which includes extraprostatic extension and seminal vesicle invasion but does not take into account surgical margin status (Bostwick *et al.*, Simin. Urol. Oncol., 17:222 [1999]). Tumors were graded using the Gleason grading system (Gleason, Cancer Chemother. Rep., 50:125 [1966]; Gleason, The Veterans Administration Cooperative Urological Research Group. Histologic Grading and Clinical Staging of Prostate Carcinoma. In: Tannenbaum M, editor. Urologic Pathology: The Prostate. Philadelphia: Lea & Febiger; 1977. p. 171-98).

As preparation for the construction of the TMAs, all glass slides were re-reviewed to identify areas of benign prostate, prostatic atrophy, high-grade prostatic intraepithelial neoplasia, and prostate cancer. To optimize the transfer of these designated tissues to the arrays, area of tumor involvement was encircled on the glass slide template as tightly around each lesion as possible. Areas with infiltrating tumor adjacent to benign glands were avoided.

C. RT-PCR

Total RNA integrity was judged by denaturing-formaldehyde agarose gel electrophoresis. cDNA was prepared using 1 µg of total RNA isolated from prostate tissue specimens. Primers used to amplify specific gene products were: AMACR sense, 5' CGTATGCCCCGCTGAATCTCGTG-3' (SEQ ID NO:100); AMACR antisense, 5'-

TGGCCAATCATCCGTGCTCATCTG-3' (SEQ ID NO:101); GAPDH sense, 5'-CGGAGTCAACGGATTTGGTCGTAT-3' (SEQ ID NO:102); and GAPDH antisense, 5'-AGCCTTCTCCATGGTGGTGAAGAC -3' (SEQ ID NO:103). PCR conditions for AMACR and GAPDH comprised 94°C for 5 min, 28 cycles of 95°C for 1 min, 60°C for 1 min (annealing), and 72°C for 1 min, and a final elongation step of 72°C for 7 min. PCR reactions used a volume of 50 μl, with 1 unit of Taq DNA polymerase (Gibco BRL). Amplification products (5 μl) were separated by 2% agarose gel electrophoresis and visualized by ultraviolet light.

10 D. Immunoblot Analysis

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Representative prostate tissue specimens were used for Western blot analysis. Tissues were homogenized in NP-40 lysis buffer containing 50 mmol/L Tris-HCl, pH 7.4, 1% Nonidet P-40 (Sigma, St. Louis. MO) and complete proteinase inhibitor cocktail (Roche, IN, USA). Fifteen µg of protein extracts were mixed with SDS sample buffer and electrophoresed onto a 10% SDS-polyacrylamide gel under reducing conditions. The separated proteins were transferred onto nitrocellulose membranes (Amersham Pharmacia Biotech, Piscataway, NJ). The membrane was incubated for 1 hour in blocking buffer (Tris-buffered saline with 0.1% Tween (TBS-T) and 5% nonfat dry milk). The AMACR antibody (Obtained from Dr. R Wanders, University of Amsterdam) was applied at 1:10,000 diluted in blocking buffer overnight at 4°C. After washing three times with TBS-T buffer, the membrane was incubated with horseradish peroxidase-linked donkey anti-rabbit IgG antibody (Amersham Pharmacia Biotech, Piscataway, NJ) at 1:5000 for 1 hour at room temperature. The signals were visualized with the ECL detection system (Amersham Pharmacia biotech, Piscataway, NJ) and autoradiography.

For \(\beta\)-tubulin western blots, the AMACR antibody probed membrane was stripped with Western Re-Probe buffer (Geno-tech, St. Louis, MO) and blocked in Trisbuffered saline with 0.1% Tween (TBS-T) with 5% nonfat dry milk and incubated with rabbit anti \(\beta\)-tubulin antibodies (Santa Cruz Biotechnologies, Santa Cruz, CA) at 1:500 for two hours. The western blot was then processed as described above.

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E. Immunohistochemistry

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Standard indirect immunohistochemistry (IHC) was performed to evaluate

AMACR protein expression using a polyclonal anti-AMACR antibody. Protein
expression was scored as negative (score=1), weak (score 2), moderate (3) and strong (4).

In order to evaluate whether AMACR protein expression was associated with prostate
cancer proliferation, a subset of samples were evaluated using the monoclonal mouse IgG
Mib-1 antibody for Ki-67 (1:150 dilution, Coulter-Immunotech, Miami, Fl). Microwave
pretreatment (30 minutes at 100 C in Tris EDTA Buffer) for antigen retrieval was
performed using 3,3' diaminobenzidine tetrahydrocloride as a chromogen. Lymph node
tissue with known high Ki-67 positivity was used as a control.

F. Tissue Microarray Construction, Digital Image Capture, and Analysis

Five TMAs were used for this study. Three contained tissue from the prostatectomy series and two contained hormone refractory prostate cancer from the Rapid Autopsy Program. The TMAs were assembled using the manual tissue arrayer (Beecher Instruments, Silver Spring, MD) as previously described (Kononen *et al.*, Nat. Med., 4:844 [1998]; Perrone *et al.*, J. Natl. Cancer Inst., 92:937 [2000]). Tissue cores from the circled areas (as described above) were targeted for transfer to the recipient array blocks. Five replicate tissue cores were sampled from each of the selected tissue types. The 0.6 mm diameter TMA cores were each spaced at 0.8 mm from core-center to core-center. After construction, 4 μm sections were cut and H&E staining was performed on the initial slide to verify the histology.

TMA H&E images were acquired using the BLISS Imaging System (Bacus Labs, Lombard, IL). AMACR protein expression was evaluated in a blinded manner. All images were scored for AMACR protein expression intensity. In addition, all TMA samples were assigned a diagnosis (i.e., benign, atrophy, high-grade prostatic intraepithelial neoplasia, and prostate cancer). This is recommended because the targeted tissues may not be what were actually transferred. Therefore, verification was performed at each step. TMA slides were evaluated for proliferation index using a CAS200 Cell Analysis System (Bacus Labs). Selected areas were evaluated under the 40X objective. Measurements were recorded as the percentage of total nuclear area that was positively

stained. All positive nuclear staining, regardless of the intensity, was measured. Sites for analysis were selected to minimize the presence of stromal and basal cells; only tumor epithelium was evaluated. Specimens were evaluated for Ki-67 expression as previously described (Perrone *et al.*, J. Natl. Cancer Inst. 92:937 [2000]). Each measurement was based on approximately 50–100 epithelial nuclei. Due to the fixed size of TMA samples, 5 repeat non-overlapping measurement was the maximum attainable.

G. Analysis of Prostate Needle Biopsies

In order to evaluate the usefulness of AMACR expression in diagnostic 18 gauge needle biopsies, 100 consecutive biopsies with prostate cancer or atypia that required further work-up were tested for AMACR expression. All cases were immunostained using two basal cell specific markers (34βE12 and p63) and AMACR. Cases were evaluated for cancer sensitivity and specificity. Twenty-six of these cases were seen in consultation with a pathologist and were considered diagnostically difficult, requiring expert review and additional characterization.

H. Results

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Figure 11 shows a schematic of the DNA and tissue microarray paradigm that lead to the discovery and characterization of AMACR in prostate cancer. A) Prostate cancer progression as adapted from Abate-Shen and Shen, (Genes Dev., 14:2410 [2000]). Distinct molecular changes occur at each stage of prostate cancer progression that can be studied using DNA microarray or "chip" technology. B) cDNA generated from tumor (prostate cancer) and reference (benign prostate tissue) samples is labeled with distinguishable fluorescent dyes and interrogated with a DNA microarray that can monitor thousands of genes in one experiment. C) After hybridization, the DNA microarray is analyzed using a scanner and fluorescence ratios determined for each gene (in this case prostate cancer/ benign tissue). D) The ratios are deposited into a computer database and subsequently analyzed using various statistical algorithms. One exemplary method of surveying the data (Eisen et al., PNAS 95:14863 [1998]) assigns color intensity to the ratios of gene expression. In this case, shades of red represent genes that are up-regulated in prostate cancer (e.g., a ratio of 4.0) and shades of green represent

genes that a down-regulated (e.g., ratio of 0.1). Genes that are unchanged between tumor and benign tissues are represented by a black color and missing elements by a gray color.

E) Genes that are identified by DNA microarray can then be validated at the transcript level (e.g., Northern blot, RT-PCR) or at the protein level (e.g., immunoblot). F)

Construction of prostate cancer tissue microarrays facilitates the study of hundreds of patients (rather than hundreds of genes). G) Each tissue microarray slide contains hundreds of clinically stratified prostate cancer specimens linked to clinical and pathology databases (not shown). H) Tissue microarray slides can then be analyzed using various molecular or biochemical methods (in this case immunohistochemistry). I) Both DNA and tissue microarray data have clinical applications. Examples include, but are not limited to: 1. using gene expression profiles to predict patient prognosis, 2. identification of clinical markers and 3. development of novel therapeutic targets.

Figure 12 summarizes AMACR transcript levels as determined by DNA microarray analysis over 57 prostate cancer specimens. Samples (Dhanasekaran *et al.*, Nature 412: 822 [2001]) were grouped according to tissue type and averaged. The experimental sample was labeled in the Cy5 channel while the reference sample (pool of benign prostate tissue) was labeled in the Cy3 channel. The box-plot demonstrates the range of AMACR expression within each group. Tissues were grouped into the following classes benign (normal adjacent prostate tissue), benign prostatic hyperplasia (BPH), clinically localized prostate cancer, and metastatic prostate cancer. In relation to benign prostate tissues, localized prostate cancer and metastatic prostate cancer were 3.1 (Mann-Whitney test, p<0.0001) and 1.67 (Mann-Whitney test, p<0.004) fold upregulated, respectively (represented as Cy5/Cy3 ratios).

DNA microarray results of AMACR mRNA levels were validated using an independent experimental methodology. AMACR-specific primers were generated and RT-PCR performed on the various RNA samples from 28 prostate tissue specimens and 6 prostate cell lines (Figure 13A). GAPDH served as the loading control. Pool, refers to RNA from normal prostate tissues obtained from a commercial source. NAP, normal adjacent prostate tissue from a patient who has prostate cancer. 3+3, 3+4, 4+4, refers to the major and minor Gleason patterns of the clinically localized prostate cancer (PCA) examined. MET, metastatic prostate cancer. Various prostate cell lines are also

examined. RT-PCR without enzyme served as a negative control. An RT-PCR product was clearly observed in the 20 localized prostate cancer samples but not in the benign samples examined. Metastatic prostate cancer and prostate cell lines displayed varying levels of AMACR transcript as compared to localized prostate cancer.

In order to gauge AMACR protein levels, immunoblot analysis was performed on selected prostate tissue extracts (Figure 13B). B-tubulin served as a control for sample loading. Similar to AMACR transcript, over-expression of AMACR protein was observed in malignant prostate tissue relative to benign prostate tissue.

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In order to validate protein expression of AMACR in situ, a separate cohort of 10 prostate samples from those used in the cDNA expression array analysis was used. These prostate samples were taken from the University of Michigan Prostate SPORE Tissue Core and were assembled onto high-density tissue microarrays (schematically illustrated in Figure 11F-H). Moderate to strong AMACR protein expression was seen in clinically localized prostate cancer samples with predominately cytoplasmic localization. A large contrast in levels of AMACR in malignant epithelia relative to adjacent benign epithelia was seen. Prostatic intraepithelial neoplasia (PIN) and some atrophic lesions, which are thought to be potentially pre-cancerous lesions (Putzi et al., Urology 56:828 [2000]; Shah et al., Am. J. Pathol., 158:1767 [2001]), demonstrated cytoplasmic staining of AMACR. High-grade prostate cancer also demonstrated strong cytoplasmic staining. However, no association was identified with AMACR staining intensity and Gleason (tumor) score. Many of the metastatic prostate cancer samples demonstrated only weak AMACR expression. The metastatic samples showed uniform PSA immunostaining, confirming the immunogenicity of these autopsy samples.

In order to assess AMACR protein expression over hundreds of prostate specimens, the tissue microarray data was quantitated. Benign prostate, atrophic prostate, PIN, localized prostate cancer, and metastatic prostate cancer demonstrated mean AMACR protein staining intensity of 1.0 (SE 0), 2.0 (SE 0.1), 2.5 (SE 0.1), 3.0 (SE 0), and 2.5 (SE 0.1), respectively (ANOVA p-value<0.0001). This data is graphically summarized using error bars representing the 95% confidence interval for each tissue category (Figure 14).

The correlation of AMACR levels with tumor proliferation was next investigated using Ki-67 (Perrone et al., supra). There was no significant association between AMACR expression and Ki-67 expression with a correlation coefficient of 0.13 (p-value=0.22). In addition, no significant associations were identified between AMACR protein expression and pathology parameters such as radical prostatectomy, Gleason score, tumor stage, tumor size (cm), or surgical margin status. AMACR protein levels were next evaluated for association with PSA recurrence following surgery in 120 prostatectomy cases with a median follow-up time of 3 years. No statistically significant association was identified. AMACR demonstrated uniform moderate to strong expression in clinically localized prostate cancer with a high sensitivity for tumor and an equally high specificity. In addition, a preliminary survey of normal tissues including ovary, liver, lymph nodes, spleen, testis, stomach, thyroid, colon, pancreas, cerebrum, and striated muscle revealed significant AMACR protein expression in only normal liver.

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The large difference in AMACR protein levels between normal secretory epithelial cells and malignant cells provides a clinical use for testing AMACR expression in prostate needle biopsy specimens. In diagnostically challenging cases, pathologists often employ the basal cell markers 34βE12 (O'Malley et al., Virchows Arch A Patho. Anat. Histopathol., 417:191 [1990]; Wojno et al., Am. J. Surg. Pathol., 19:251 [1995]; Googe et al., Am. J. Clin. Pathol., 107:219 [1997] or p63 (Parson et al., Urology 58:619 [2001]; Signoretti et al., Am. J. Pathol., 157:1769 [2000]), which stain the basal cell layer of benign glands. This second basal cell layer is absent in malignant glands. In many equivocal biopsy specimens, the surgical pathologist must rely on absence of staining to make the final diagnosis of prostate cancer. The clinical utility of AMACR immunostaining on 94 prostate needle biopsies was evaluated. The results are shown in Table 2. The sensitivity and specificity were calculated as 97% and 100%, respectively. These results included 26 cases where the final diagnosis required the use of a basal cell specific immunohistochemical marker (i.e., 34βE12 or p63).

This example demonstrated that AMACR is associated with PCA and that AMACR expression in prostate biopsies is useful for the diagnosis of cancer in inconclusive biopsy samples.

	•	Table 2								
Clinical utility of Assessing AMACR Protein in Prostate Needle Biopsies (n=94)										
Sensitivity	Specificity	Positive Predictive Value	Negative Predictive							
(TP/(TP+FN))	(TN/(TN+FP))	(TP/(TP+FP))	Value (TN/(TN+FN))							
97% ((68/(2+68))	100% ((24/(24+0))	100% ((68/(68+0))	92% ((24/24+2))							

Example 6

Hormone Regulation of AMACR

This example describes studies that indicate that AMACR expression is hormone independent.

A. Sample collection, cDNA array and TMA construction and evaluation

Clinical samples were taken from the radical prostatectomy series and from the Rapid Autopsy Program at the University of Michigan. Both are part of the University of 10 Michigan Prostate Cancer Specialized Program of Research Excellence (S.P.O.R.E.). Primary PCA of metastatic cases as well as lymph node metastases were contributed in collaboration from the University of Ulm, Germany. Detailed clinical and expression analysis as well as TMA data was acquired and maintained on a secure relational database according to the Institutional Review Board protocol of both institutions. 15 Tissue procurement for expression analysis on the RNA level is described in the above examples. For the development of TMA, samples were embedded in paraffin. The study pathologist reviewed slides of all cases and circled areas of interest. These slides were used as a template for construction of the six TMAs used in this study. All TMAs were assembled using a manual tissue arrayer (Beecher Instruments, Silver Spring, MD). At 20 least three tissue cores were sampled from each donor block. Histologic diagnosis of the tissue cores was verified by standard haematoxylin and eosin (H&E) staining of the initial TMA slide. Standard biotin-avidin complex immunohistochemistry (IHC) was performed using a polyclonal anti-AMACR antibody (Ronald Wanders, University of Amsterdam). Digital images were acquired using the BLISS Imaging System (Bacus 25 Lab, Lombard, IL). Staining intensity was scored as negative (score=1), weak (score 2), moderate (3) and strong (4). For exploration of the treatment effect by the means of

hormonal withdrawal before radical prostatectomy, standard slides were used for regular H&E staining and consecutive sections for detection of AMACR expression. In order to test AMACR expression in poorly differentiated colon cancers, cases were used from a cohort of well-described colon tumors. In addition to well-differentiated colon cancers, a recently described subset of poorly differentiated colon carcinomas with a distinctive histopathological appearance, termed large cell minimally differentiated carcinomas, was used. These poorly differentiated colon carcinomas had a high frequency of the microsatellite instability phenotype.

10 B. Cell culture and immunoblot analysis

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Prostate cell lines (RWPE-1, LNCaP, PC3 and DU145) were obtained from the American Tissue Culture Collection. Cells were maintained in RPMI-1640 with 8% decomplemented fetal bovine serum, 0.1% glutamine and 0.1% penicillin and streptomycin (BioWhittaker, Walkersville, MD). Cells were grown to 75% confluence and then treated for 24 and 48 with the antiandrogen bicalutamide (CASODEX, Zeneca Pharmaceuticals, Plankstadt, Germany) at a final concentration of 20 µM or with methyltrienolone (synthetic androgen (R1881); NEN, Life Science Products, Boston, MA) at a final concentration of 1 nM. Cells were harvested and lysed in NP-40 lysis buffer containing 50 mmol/L Tris-HCl, pH 7.4, 1% Nonidet P-40 (Sigma, St. Louis, MO) and complete proteinase inhibitor cocktail (Roche, IN, USA). 15 µg of protein extracts were mixed with SDS sample buffer and electrophoresed onto a 10% SDSpolyacrylamide gel under reducing conditions. After transferring, the membranes (Amersham Pharmacia Biotech, Piscataway, NJ) were incubated for 1 hour in blocking buffer (Tris-buffered saline with 0.1% Tween and 5% nonfat dry milk). The AMACR antibody was applied at 1:10.000 diluted blocking buffer overnight at 4°C. After three washes with TBS-T buffer, the membrane was incubated with horseradish peroxidaselinked donkey anti-rabbit IgG antibody (Amersham Pharmacia Biotech, Piscataway, NJ) at 1:5000 for 1 hour at room temperature. The signals were visualized with the ECL detection system (Amersham Pharmacia biotech, Piscataway, NJ). For B-tubulin blots, membranes were stripped with Western Re-Probe buffer (Geno-tech, St. Louis, MO) and blocked in Tris-buffered saline with 0.1% Tween with 5% nonfat dry milk and incubated

with rabbit anti \(\beta\)-tubulin antibodies (Santa Cruz Biotechnologies, Santa Cruz, CA) at 1:500 for two hours. For PSA expression the membranes were reprobed in the described manner with PSA antibody (rabbit polyclonal; DAKO Corporation, Carpinteria, CA) at 1: 1000 dilution and further processed.

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C. Statistical Analysis

Primary analysis of the cDNA expression data was done with the Genepix software. Cluster analysis with the program Cluster and generation of figures with TreeView was performed as described above. AMACR protein expression was statistically evaluated using the mean score result for each prostate tissue type (i.e., benign prostate, naive localized or advanced prostate cancer, hormone treated and hormone refractory prostate cancer). To test for significant differences in AMACR protein expression between all tissue types, a one-way ANOVA test was performed. To determine differences between all pairs, a post-hoc analysis using the Scheffé method was applied as described above. For comparison of naive primaries to their corresponding lymph node metastases with respect to AMACR protein expression, a non parametric analysis (Mann Whitney test) was performed. To compare AMACR expression intensity to the scored hormonal effect of the pretreated localized prostate cancer cases the Mantel-Haenszel Chi-Square test was applied. AMACR expression scores are presented in a graphical format using error-bars with 95% confidence intervals. P-values <0.05 were considered statistically significant.

D. Results

PCA and metastatic PCA and filtering for only those genes with a 1.5 fold expression difference or greater, clustered the samples into histologically distinct groups as described above. As demonstrated by a TreeView presentation of this data (Figure 15), AMACR was one of several genes that demonstrated over expression at the cDNA level of PCA samples with respect to benign pooled prostate tissue. The highest level of over expression by cDNA analysis was in the clinically localized PCA cases.

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In order to further investigate the role of AMACR protein expression in samples with variable differentiation and exposure to anti-androgen treatment, several TMAs with a wide range of PCA were constructed: a total of 119 benign prostate samples, 365 primary hormone naive PCA samples, 37 naive prostate cancer lymph node metastases, and 41 hormone refractory metastatic PCA samples were evaluated. An additional 49 hormone treated primary prostate cancers (including 22 on standard slides) were. examined for histologic changes associated with anti-androgen treatment and AMACR protein expression. The mean AMACR protein expression levels for each tissue category is presented in Figure 16. Benign prostate, naive primary prostate cancer, hormone treated primary cancer, and hormone refractory metastatic tissue had a mean staining intensity of 1.28 (Standard Error SE 0.038, 95% Confidence Intervals CI 1.20-1.35), 3.11(SE 0.046, CI 3.02-3.20), 2.86 (SE 0.15, CI 2.56-3.15) and 2.52 (SE 0.15, CI 2.22-2.28), respectively). One-way ANOVA analysis revealed a p-value of <0.0001. To specifically examine the difference between different tissue types, a post-hoc pair-wise comparison was performed. Clinically localized PCA demonstrated a significantly stronger AMACR protein expression as compared to benign prostate tissue (post-hoc analysis using Scheffé method, mean difference =1.83, p<0.0001, CI 1.53-2.13). A significant decrease in AMACR protein expression was observed in the metastatic hormone refractory PCA samples with respect to clinically localized PCA (0.59, p=0.002, CI 0.15-1.03). Hormone treated primaries had a mean AMACR expression of 2.86, which was between the expression levels of naive primaries (3.11) and hormone refractory cases (2.52) (post-hoc analysis using Scheffé method, p=0.51, CI -0.66-0.16 and p=0.56, CI -0.23-0.91). There was no significant difference in AMACR expression in the 37 naive primary prostate samples and lymph node metastases derived from the same patient (Mann Whitney test, p=0.8). In other words, matched primaries and lymph. node metastases showed similar AMACR expression pattern.

A subset of 22 PCA cases in which the patients received variable amount and types of anti-androgen treatment prior to surgery was examined. These cases were evaluated blindly with respect to treatment protocol for histological evidence of hormone treatment (H&E slide) and AMACR protein expression. The hormonal effect visible on the H&E slides was classified from 1 to 4 with 1 representing "no effect" and 4 showing

a "very strong effect". 13 cases demonstrated either no or moderate hormonal effect, and 9 cases had a very strong hormonal effect. Statistical analysis revealed a significant difference between these two groups with respect to AMACR expression intensity (Figure 17, Mantel-Haenszel Chi-Square, p=0.009). Figure 17 presents an example of a PCA case treated prior to surgery with anti-androgens that has a strong hormonal effect appreciated on H&E and decreased AMACR protein expression (Figure 17A). In this dataset there was neither a correlation between treatment duration nor treatment type (monotherapy or complete hormonal withdrawal for hormone deprivation) and AMACR expression.

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For further exploration of the hormonal effect on AMACR expression, primary cell culture experiments and Western blot analysis were performed. As demonstrated in Figure 17 Panel B, LNCaP cells, derived from a metastatic lesion but considered hormone responsive, showed a higher baseline AMACR expression as compared to PC3 and DU-145 cells, which are both hormone independent cell lines derived from metastatic lesions. A benign cell line, RWPE-1 (Bello et al., Carcinogenesis 18:1215 [1997]), showed near absent AMACR expression, which is consistent with the in situ protein expression data. To simulate an anti-androgen treatment, the hormone responsive cell line LNCaP was treated with bicalutamide in a final concentration of 20 µM for a time period of 24 and 48 hours. AMACR expression in cell lysates of LNCaP cells did not change at either time point when exposed to anti-androgen therapy. Under the same conditions, PSA, a gene known to be regulated by the androgen receptor, showed decreased protein expression. In addition, when LNCaP cells were exposed to a synthetic androgen R1881, no increase in AMACR expression was observed (Figure 17, Panel B). Therefore, these cell culture experiments provide evidence that AMACR expression is not regulated by the androgen pathway.

The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention.

Nonetheless, it is contemplated that another explanation for these observations was that AMACR over expression occurred in PCA, but as these tumors became poorly differentiated, as in the hormone refractory PCA, AMACR expression was down regulated either directly or indirectly due to the process of de-differentiation. To

elucidate this potential correlation colon cancer samples were examined for AMACR expression (See Example 7). AMACR protein expression is also observed in some other tumor types, with the highest overall expression in colorectal cancers. Colorectal cancers are not known to be regulated by androgens and were therefore used as a control to test this hypothesis. Four well differentiated and seven anaplastic colon cancer samples were chosen. The poorly differentiated tumors have distinct molecular alterations distinguishing them from the common well to moderately differentiated colorectal tumors (Hinoi et al., Am. J. Pathol. 159:2239 [2001]). Strong AMACR protein expression in a moderately differentiated colon cancer was observed. This tumor still forms well defined glandular structures. The surrounding benign colonic tissue does not express AMACR. The anaplastic colon cancers demonstrated weak AMACR protein expression. Primarily data revealed positive AMACR expression in 4/4 well differentiated cases but only 4/7 anaplastic colonic cancers. Three of the anaplastic colon cancers had weak to moderate expression. Metastatic hormone refractory PCA demonstrated weak AMACR protein expression in tissue microarrays.

Example 7

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AMACR Expression in a Variety of Cancers

20 A. Analysis of online EST and SAGE database

The National Cancer Institute Cancer Genome Anatomy Project (CGAP) has several gene expression databases available online for comparing gene expression across multiple samples (See the Internet Web site of the National Cancer Institute). Both EST and SAGE databases offer Virtual Northern blots, which allow users to visualize and compare the expression level of a particular gene among multiple samples. The SAGE database includes over 5 million tags from 112 libraries of multiple benign and malignant tissues.

B. Selection of study cases

A total of 96 cases of cancers from different sites were selected for construction of a multi-tumor tissue microarray. The tissue microarray was constructed to perform a

wide survey of multiple common tumor types. A minimum of three tissue cores (0.6 mm in diameter) was taken for each case. Tumors surveyed included colorectal adenocarcinoma (n=15 cases), renal cell carcinoma (6), prostatic adenocarcinoma (6), urothelial carcinoma (4), cervical squamous cell carcinoma (6), lung non-small cell carcinoma (4), lymphoma (15), melanoma (9) and several other cancer types. Normal adjacent tissue was taken when available. The prostate tissue microarray was constructed from selected patients who underwent radical prostatectomies as monotherapy for clinically localized prostate cancer. This tissue microarray contained a spectrum of prostatic tissue including prostatic atrophy, high-grade prostatic intraepithelial neoplasia (PIN), and clinically localized prostate cancer. In addition, standard slides were used to confirm results for colon cancer. Twenty-four cases of colorectal adenocarcinoma (16 well to moderately differentiated carcinoma and 8 large cell minimally differentiated carcinoma) and 8 endoscopically derived colorectal adenomas were selected for immunostaining for AMACR. For breast carcinoma, a TMA of 52 cases of invasive ductal carcinoma was used. Specimens were collected and analyzed in accordance with the Institutional Review Board guidelines.

C. Immunohistochemistry

Standard avidin-biotin complex immunohistochemistry was used. Pre-treatment was performed by steaming the slides for 10 minutes in sodium citrate buffer in a microwave oven. The slides were then incubated sequentially with primary antibody (1:2000 dilution, polyclonal rabbit anti-AMACR antibody), biotinylated secondary antibody, avidin-biotin complex and chromogenic substrate 3,3'-diaminobenzidine. Slides were evaluated for adequacy using a standard bright field microscope. Digital images were then acquired using the BLISS Imaging System (Bacus Lab, Lombard, IL) and evaluated by two pathologists. Protein expression was scored as negative, weak stain (faint cytoplasmic stain or granular apical staining), moderate (diffuse granular cytoplasmic stain) and strong (diffuse intense cytoplasmic stain). Only moderate and strong staining was considered as positive staining.

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D. Laser Capture Microdissection

Sections of 2 radical prostatectomy samples were frozen in OCT in accordance with an Institutional Review Board protocol. Frozen sections (5 μ m thick) were fixed in 70% alcohol for 10 minutes and then stained in hemotoxylin and eosin. Prostate cancer and benign prostate glands were dissected on a μCUT laser capture microdissector (MMI GmbH, Heidelberg, Germany). Approximately 6000 cells were harvested. Total RNA was isolated using Qiagen micro-isolation kit (Qiagen, San Diego, CA). Reverse transcription was performed using both oligo dT and random hexamer primers. Primers used to amplify specific gene products were: AMACR sense, 5'-CGTATGCCCCGCTGAATCTCGTG-3' (SEQ ID NO:104); AMACR antisense, 5'-

CGTATGCCCGCTGAATCTCGTG-3' (SEQ ID NO:104); AMACR antisense, 5'-TGGCCAATCATCCGTGCTCATCTG-3' (SEQ ID NO:105); GAPDH sense, 5'AGCCTTCTCCATGGTGGTGAAGAC-3' (SEQ ID NO:106); and GAPDH antisense, 5'-AGCCTTCTCCATGGTGGTGAAGAC-3' (SEQ ID NO:107). PCR conditions for AMACR and GAPDH were: heat denaturation at 94°C for 5 min, cycles of 94°C for 1 min, 60°C for 1 min, and 72°C for 1 min (32 cycles for GAPDH, 40 cycles for AMACR), and a final extension step at 72°C for 5 min. PCR products were then separated on 2% agarose gel and visualized by UV illumination.

E. Results

Using the Virtual Northern tool from the online CGAP program, AMACR expression was surveyed in two databases, EST and SAGE libraries. AMACR was found to be expressed in a wide range of tissues, including central and peripheral nervous system, colon, kidney, breast, pancreas, prostate and blood. Compared to their normal counterparts, a number of cancers have elevated AMACR expression, including tumors arising in bone marrow, breast, colon, genitourinary system, lung, lymph node, nervous system, pancreas, prostate, soft tissue and uterus.

To confirm the gene expression data, AMACR immunohistochemistry was performed on a multi-tumor tissue array that included some of the most common cancers from multiple sites. AMACR protein level was increased in many cancers, including colorectal, prostate, ovarian, lung cancers, lymphoma and melanoma (Figure 18). In particular, AMACR over-expression was observed in 92% and 83% of colorectal and

prostate cancer, respectively. Using a breast cancer tissue microarray, it was found that AMACR over-expression was present in 44% of invasive ductal carcinomas. AMACR over expression was not observed in female cervical squamous cell carcinoma (6 cases).

To further characterize AMACR expression in a spectrum of proliferative prostate lesions, a prostate tissue microarray, which included prostate cancer, high grade PIN and atrophic glands, was utilized. Positive AMACR staining (moderate and strong staining) was observed in 83% and 64% of clinically localized prostate cancer and high-grade PIN, respectively. Focal AMACR expression was observed in 36% of the atrophic lesions and in rare morphologically benign glands. To confirm that AMACR protein over-expression was the result of increased gene transcription, laser capture microdissection was used to isolate cancerous and benign prostatic glands. RT-PCR was performed to assess the AMACR mRNA expression. Benign glands had very low baseline expression (Figure 19). In contrast, prostate cancer had much higher mRNA level, confirming that increased AMACR gene transcription leads to elevated protein over expression in prostate cancer.

AMACR expression was studied in 24 colorectal adenocarcinomas, including 16 well to moderately differentiated, and 8 poorly differentiated large cell adenocarcinomas. Overall, 83% (20/24) demonstrated positive AMACR protein expression. All (16/16, 100%) cases of well to moderately differentiated carcinoma had positive staining, compared to 64% (5/8) of poorly differentiated carcinoma. AMACR expression was examined in 8 colorectal adenoma biopsies obtained by colonoscopy. Moderate staining was present in 6 (75%) cases. Compared with well-differentiated adenocarcinomas, adenomas usually showed more focal (10-60% of cells) and less intense staining.

Example 8

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25 Characterization of EZH2 expression in Prostate Cancer

A. SAM Analysis

SAM analysis was performed by comparing gene expression profiles of 7 metastatic prostate cancer samples against 10 clinically localized prostate cancer samples. Data was normalized per array by multiplication by a factor to adjust the aggregate ratio of medians to one, then log base 2 transformed and median centered. This normalized

data was divided into two groups for comparison using a two-class, unpaired t-test. Critical values for the analysis include: Iterations = 500, Random Number Seed 1234567, a fold change cutoff of 1.5 and a delta cutoff of 0.985, resulting in a final largest median False Discovery Rate of 0.898 % for the 535 genes selected as significant (55 relatively up and 480 relatively down regulated between MET and PCA). These 535 genes were analyzed using Cluster (Eisen et al., PNAS 95:14863 [1998]) implementing average linkage hierarchical clustering of genes. The output was visualized by Treeview (Eisen et al., [1998], supra).

10 B. RT-PCR

Reverse transcription and PCR amplification were performed with 1 μg total RNA isolated from the indicated prostate tissues and cell lines. Human EZH2 forward (5'-GCCAGACTGGGAAGAAATCTG-3' (SEQ ID NO:108)), reverse (5'-TGTGCTGGAAAATCCAAGTCA-3' (SEQ ID NO:109)) and GAPDH sense (5'-CGGAGTCAACGGATTTGGTCGTAT-3' (SEQ ID NO:110)), antisense 5'-AGCCTTCTCCATGGTGGTGAAGAC-3' (SEQ ID NO:111)) primers were used. The amplified DNA was resolved on agarose gels and visualized with ethidium bromide.

C. Immunoblot Analysis

Prostate tissue extracts were separated by SDS-PAGE and blotted onto nitrocellulose membranes. Anti-EZH2 (Sewalt et al., Mol. Cell. Biol. 18:3586 [1998]), anti-EED (Sewalt et al., supra), and polyclonal anti-tubulin (Santa Cruz biotechnology) antibodies were used at 1:1000 dilution for immunoblot analysis. The primary antibodies were detected using horseradish peroxidase-conjugated secondary antibodies and visualized by enhanced chemiluminescence as described by the manufacturer (Amersham-Pharmacia).

D. Tissue Microarray Analysis

Clinically stratified prostate cancer tissue microarrays used in this study have

been described previously (See above examples). Tissues utilized were from the radical
prostatectomy series at the University of Michigan and from the Rapid Autopsy Program,

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which are both part of University of Michigan Prostate Cancer Specialized Program of Research Excellence (S.P.O.R.E.) Tissue Core. Institutional Review Board approval was obtained to procure and analyze the tissues used in this study.

EZH-2 protein expression was evaluated on a wide range of prostate tissue to 5 determine the intensity and extent in situ. Immunohistochemistry was performed on three tissue microarrays (TMA) containing samples of benign prostate, prostatic atrophy, high-grade prostatic intraepithelial neoplasia (PIN), clinically localized prostate cancer (PCA), and metastatic hormone refractory prostate cancer (HR-METSs). Standard biotin-avidin complex immunohistochemistry (IHC) was performed to evaluate EZH2 protein expression using a polyclonal anti-EZH2 antibody. Protein expression was scored as negative (score=1), weak (score 2), moderate (3) and strong (4).

Approximately 700 TMA samples (0.6 mm diameter) were evaluated for this study (3-4 tissue cores per case). The TMAs were assembled using a manual tissue arrayer (Beecher Instruments, Silver Spring, MD) as previously described (See above examples). Four replicate tissue cores were sampled from each of the selected tissue types. After construction, 4 µm sections were cut and hematoxylin and eosin staining was performed on the initial slide to verify the histologic diagnosis. TMA hematoxylin and eosin images were acquired using the BLISS Imaging System (Bacus Lab, Lombard, IL). EZH2 protein expression was evaluated in a blinded manner by the study pathologist using a validated web-based tool (Manley et al., Am. J. Pathol. 159:837 [2001]; Bova et al., Hum. Pathol. 32:417 [2001]) and the median value of all measurements from a single patient were used for subsequent analysis.

Clinical Outcomes Analysis E.

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To assess individual variables for risk of recurrence, Kaplan-Meier survival analysis was performed and a univariate Cox proportional hazards model was generated. PSA-recurrence was defined as 0.2 ng/ml following radical prostatectomy. Covariates included Gleason sum, preoperative PSA, maximum tumor dimension, tumor stage, and surgical margin status. To assess the influence of several variables simultaneously including EZH2 protein expression, a final multivariate Cox proportional hazards model of statistically significant covariates was generated. Statistical significance in univariate

and multivariate Cox models were determined by Wald's test. A p-value <0.05 was considered statistically significant.

F. EZH2 Constructs

Myc-tagged EZH2-pCMV was used. The Myc-EZH2 fragment was released with BamHI/XhoI double digest and was sub-cloned into the mammalian expression vector pCDNA3 (Invitrogen). An EZH2-ER in-frame fusion expression construct was generated by replacing the FADD fragment released by Kpn I/Not I double digest of the FADD-ER construct (originally derived from Myc-ER (Littlewood et al., Nuc. Acids.

10 Res. 23:1686 [1995]) with the PCR amplified human EZH2 devoid of its stop codon.

The EZH2 .SET mutant DNA was amplified using the primers

5'GGGGTACCATGGGCGGCCGCGAACAAAAGTTGATT 3' (SEQ ID NO:112) and

5'GGGGAATTCTCATGCCAGCAATAGATGCTTTTT3' (SEQ ID NO:113) and

subsequently sub-cloned into pCDNA3 utilizing the in built KpnI/EcoRI sites.

Expression of these constructs was verified by immunoblot analysis of the expressed proteins using either anti-Myc HRP (Roche, Inc) or anti-EZH2 antibodies.

G. RNA interference

21-nucleotide sense and antisense RNA oligonucleotides were chemically

20 synthesized (Dharmacon Research Inc.) and annealed to form duplexes. The

siRNA employed in the study were targeted to the region corresponding from 85 to 106

of the reported human EZH2 (NM004456). Control siRNA duplexes targeted luciferase,
lamin and AMACR (NM014324). The human transformed prostate cell line RWPE

(Webber et al., Carcinogenesis 18:1225 [1997]) and PC3 were plated at 2x10⁵ cells per

25 well in a 12 well plate (for immunoblot analysis, cell counts, and fluorescence activated

cell sorting (FACS) analysis) and 1.5x10⁴ cell per well in a 96 well plate (for WST-1

proliferation assays). Twelve hours after plating, the cells were transfected with 60

picomoles of siRNA duplex, sense or antisense oligonucleotides (targeting EZH2) using

oligofectamine (Invitrogen). A second identical transfection was performed 24 hours

later. Forty-eight hours after the first transfection, the cells were lysed for immunoblot

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analysis and trypsinized for cell number estimation or FACS analysis. Cell viability was assessed 60 hours after the initial transfection.

H. Cell Proliferation Assays

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Cell proliferation was determined with the colorimetric assay of cell viability, based on the cleavage of tetrazolium salt WST-1 by mitochondrial dehydrogenases as per manufacturers instructions (Roche, Inc.). The absorbance of the formazan dye formed, which directly correlates with the number of metabolically active cells in the culture, was measured at 450nm (Bio-Tek instruments), an hour after the addition of the reagent. Cell counts were estimated by trypsinizing cells and analysis by coulter cell counter.

I. Flow cytometric analysis

Trypsinized cells were washed with phosphate buffered saline (PBS) and cell number was determined by using a coulter cell counter. For FACS analysis, the washed cells were fixed in 70% ethanol overnight. Before staining with propidium iodide, the cells were washed again with PBS and analyzed by flow cytometry (Becton Dickinson).

J. Microarray analysis of EZH2 transfected cells

Initial testing of this transient transfection/transcriptome analysis system demonstrated that transient overexpression of TNFR1 (p55), a receptor for tumor necrosis factor, induced similar expression profiles as was observed with incubation of cells with TNF (Kumar-Smith *et al.*, J. Biol. Chem. 24:24 [2001]). Other molecules have been similarly tested with this approach. Cells were transfected with different EZH2 constructs and transfection efficiency was monitored by beta-galactosidase assay and was approximately 30-50%. EZH2 .SET mutant expressing samples were compared to EZH2 expressing samples using the SAM analysis package (Tusher *et al.*, PNAS 98:5116 [2001]). Data was pre-processed by multiplication by a normalization factor to adjust the aggregate ratio of medians to one, log base 2 transformed and median centered each array, individually. This pre-processed data was divided into 2 groups for comparison using a two-class, unpaired t-test. Critical values for the analysis include: iterations = 5000, (720 at convergence) random Number Seed 1234567, a fold change of 1.5 and a

delta cutoff of 0.45205, resulting in a final largest median False Discovery Rate of 0.45% for the 161 genes selected as significant. These 161 genes were supplemented by the values for EZH2 and then analyzed using Cluster implementing average linkage hierarchical clustering of genes. The output was visualized in Treeview. Selected genes identified as being repressed by EZH2 (e.g., EPC and cdc27) were re-sequenced to confirm identity.

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The molecular identity of a cell is determined by the genes it expresses (and represses). Embryogenesis and cell differentiation intimately depend upon keeping certain genes "on" and other genes "off". When the transcriptional "memory" of a cell is perturbed this can lead to severe developmental defects (Jacobs *et al.*, Semin. Cell Dev. Biol. 10:227 [1999]; Francis *et al.*, Nat. Rev. Mol. Cell. Biol. 2:409 [2001]). Lack of differentiation, or anaplasia, is a hallmark of cancer, which results from normal cells "forgetting" their cellular identity. Thus, it is not surprising that dysregulation of the transcriptional maintenance system can lead to malignancy (Francis *et al.*, *supra*; Jabobs *et al.*, Nature 397:164 [1999]; Beuchle *et al.*, Development 128:993 [2001]).

Studies in Drosophila melanogaster have been instrumental in the understanding of the proteins involved in transcriptional maintenance (Beuchle et al., [[2001], supra; Strutt et al., Mol. Cell. Biol. 17:6773 [1997]; Tie et al., Development 128:275 [2001]). Two groups of proteins have been implicated in the maintenance of homeotic gene expression and include polycomb (PcG) and trithorax (trxG) group proteins (Mahmoudi et al., Oncogene 20:3055 [2001]; Lajeunesse et al., Development 122:2189 [1996]). PcG proteins act in large complexes and are thought to repress gene expression, while trxG proteins are operationally defined as antagonists of PcG proteins and thus activate gene expression (Francis et al., Nat. Rev. Mol. Cell. Biol. 2:409 [2001]; Mahmoudi et al., supra). There are at least twenty PcG and trxG proteins in Drosophila, and many have mammalian counterparts. In human malignancies, PcG and trxG proteins have primarily been found to be dysregulated in cells of hematopoietic origin (Yu et al., Nature 378:505 [1995]; Raaphorst et al., Am. J. Pathol., 157:709 [2000]; van Lohuizzen et al., Cell 65:737 [1991]. EZH2 is the human homolog of the Drosophila protein Enhancer of Zeste (E(z)) ((Laible et al., Embo. J. 16:3219 [1997]), for which genetic data defines as a PcG protein with additional trxG properties (LaJeunesse et al., supra). E(z) and EZH2

share homology in four regions including domain I, domain II, a cysteine-rich amino acid stretch, and a C-terminal SET domain (Laible et al., supra). The SET domain is a highly conserved domain found in chromatin-associated regulators of gene expression often modulating cell growth pathways (Jenuwein et al., Cell. Mol. Life Sci. 54:80 [1998]).

EZH2 is thought to function in a PcG protein complex made up of EED, YY1 and HDAC2 (Satijn *et al.*, Biochim. Biophys. Acta. 1447:1 [1999]). Disruption of the EZH2 gene in mice causes embryonic lethality suggesting a crucial role in development (O'Carroll *et al.*, Mol. Cell. Biol. 21:4330 [2001]).

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In previous studies (See e.g., Example 1), the gene at the top of the "list" of genes significantly up-regulated in metastatic prostate cancer was EZH2, which had a d-score (Tusher et al. PNAS 98:5116 [2001]) of 4.58 and a gene-specific FDR of 0.0012 (also called a "q-value" which is analogous to p-values, but adapted to multiple inference scenarios. Figure 20a displays the 55 up-regulated genes identified by this approach. Figure 20b summarizes the gene expression of EZH2 in 74 prostate tissue specimens analyzed on DNA microarrays made up of 10 K elements. The EZH2 transcript was significantly increased in metastatic prostate cancer with respect to clinically localized prostate cancer (Mann-Whitney test, p=0.001) and benign prostate (p=0.0001).

As independent experimental validation of DNA microarray results, RT-PCR was performed on 18 prostate samples and cell lines. As expected, EZH2 mRNA transcript levels were elevated in malignant prostate samples relative to benign (Fig. 20c). To determine whether EZH2 is up-regulated at the protein level in metastatic prostate cancer, tissue extracts were examined by immunoblotting. In the samples examined by immunoblot analysis, EZH2 protein was markedly elevated in metastatic prostate cancer relative to localized prostate cancer or benign prostate (Fig. 20d).

Importantly, EED, a PcG protein that forms a complex with EZH2 (vanLohuizen et al., supra; Sewalt et al., supra), along with an un-related protein, \(\beta\)-tubulin, did not exhibit similar protein dysregulation. EZH2 protein expression was evaluated on a wide range of prostate tissues (over 700 tissue microarray elements) to determine the intensity and extent of expression in situ (Fig. 21 a,b). When highly expressed, EZH2 expression was primarily observed in the nucleus as suggested previously (Raaphorst et al., supra). The staining intensity was increased from benign, prostatic atrophy, prostatic

intraepithelial neoplasia (PIN), to clinically localized prostate cancer with median staining intensity of 1.7 (standard error [SE], 0.1; 95% confidence interval [CI], 1.5-1.9), 1.7 (SE, 0.2; 95%CI, 1.3-2.0), 2.3 (SE, 0.2.; 95%CI, 1.9-2.7), and 2.6 (SE, 0.1; 95%CI, 2.4-2.8), respectively (Fig. 24b). The strongest EZH2 protein expression was observed in hormone-refractory metastatic prostate cancer with a median staining intensity of 3.3 (SE, 0.3; 95%CI, 2.7-3.9). There was a statistically significant difference in EZH2 staining intensity between benign prostate tissue and localized prostate cancer (ANOVA post-hoc analysis mean difference 0.9, p<0.0001). Although metastatic prostate cancer had a higher mean expression level than localized prostate cancer, the difference did not reach statistical significance (ANOVA post-hoc analysis mean difference 0.7, p=0.3). These findings suggest that as prostate neoplasia progresses there was a trend towards increased EZH2 protein expression, mimicking that seen by DNA expression array analysis. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that this observation suggests that EZH2 levels may indicate how aggressive an individual's prostate cancer is given that the highest level of expression was observed in hormone-refractory, metastatic prostate cancer. Therefore, to test this hypothesis, the utility of EZH2 protein levels to predict clinical outcome in men treated with surgery for clinically localized prostate cancer was examined.

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Two hundred and twenty-five (225) specimens from sixty-four patients (3-4 replicate samples per patient) with clinical follow up were interrogated on a single tissue microarray. These men had a median age of 61 years (range 43-76 years) and a 7.3 ng/ml median pre-operative serum prostate specific antigen (PSA) (range 0.8-21.0 ng/ml). Pathologic examination of their prostatectomy specimens indicated that 77% had organ-confined disease (pT2 stage) and 72% had negative surgical margins. The patient demographics and tumor stages were representative of the over 1500 radical prostatectomy patients. In order to test the utility of EZH2 as a potential tissue biomarker for prostate cancer, the clinical outcome of these 64 cases was examined, taking into account clinical and pathological parameters. Clinical failure was defined as either a 0.2 ng/ml PSA elevation or disease recurrence following prostatectomy (e.g., development of metastatic disease). By Kaplan-Meier analysis (Fig. 21c), EZH2 staining intensity of 3

and greater was significantly associated with clinical failure in 31% (10/32) of patients in contrast to 9% (3/32) of patients with an EZH2 protein levels below 3 (log rank p=0.03). There was no significant correlation between EZH2 levels and Gleason score (<7 versus =7), tumor stage (pT2 versus pT3), or surgical margin status (negative versus positive). There was a significant (p=0.048) albeit weak (Pearson coefficient =0.33) correlation between EZH2 protein levels and proliferation index in situ as assessed by Ki-67 labeling index. Multivariable Cox-Hazards regression analysis revealed that EZH2 protein expression (=3 versus <3) was the best predictor of clinical outcome with a recurrence ratio of 4.6 (95%CI 1.2-17.1, p=0.02), which was significantly better than surgical margin status, maximum tumor dimension, Gleason score, and pre-operative PSA. Thus, monitoring EZH2 protein levels in prostate specimens may provide additional prognostic

information not discernible with current clinical and pathology parameters alone.

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To shed light into the functional role of EZH2 in prostate cancer progression, EZH2 expression in transformed prostate cells in vitro was disrupted using RNA interference. T. Tuschl and colleagues recently reported that duplexes of 21-nucleotide RNA (siRNAs) mediate RNA interference in cultured mammalian cells in a gene-specific fashion (Elbashir et al., Nature 411:494 [2001]). RNA interference has been used effectively in insect cell lines to "knock-down" the expression of specific proteins, owing to sequence-specific, double stranded-RNA mediated RNA degradation (Hammond et al., Nature 404:293 [2000]). siRNAs are potent mediators of gene silencing, several orders of magnitude more potent than conventional antisense or ribozyme approaches (Macejak et al., Hepatology 31:769 [2000]). Thus, a 21- nucleotide stretch of the EZH2 molecule was targeted using criteria provided by Elbashir et al. (supra), and RNA oligonucleotides were synthesized commercially. After the RNA oligos were annealed to form siRNA duplexes, they were tested on the transformed androgen-responsive prostate cell line RWPE (Webber et al., Carcinogenesis 18:1225 [1997]; Bello et al., Carcinogenesis 18:1215 [1997]) as well as the metastatic prostate cancer cell line PC3. Forty-eight hours after transfection with siRNA duplexes, the levels of endogenous EZH2 protein were quntitated. When EZH2 protein was specifically down-regulated in prostate cell lines, the levels of the un-related control protein, β-tubulin, remained unchanged (Fig. 22a). The sense or anti-sense oligonucleotides comprising the EZH2 duplex, as well as un-

related siRNA duplexes, did not affect EZH2 protein levels (Fig. 22a, middle and right panels), verifying the specificity of the siRNA approach in both prostate cell lines.

The phenotype of EZH2 "knock-down" prostate cells was next examined. By phase contrast microscopy, it was observed that siRNA directed against EZH2 markedly inhibited cell number/confluency relative to buffer control. Cell counts taken 48 hrs after transfection with siRNA showed a 62% inhibition of RWPE cell growth mediated by the EZH2 siRNA duplex, which is in contrast to the corresponding sense and anti-sense EZH2 oligonucleotides or control duplexes (targeting luciferase and lamin) which exhibited minimal inhibition (Fig. 22b). The prostate cancer cell line, PC3, demonstrated a similar growth inhibition mediated by EZH2 siRNA, suggesting that the findings are not a peculiarity of the RWPE cell line (Fig. 22b). Using a commercially available cell proliferation reagent WST-1, which measures mitochondrial dehydrogenase activity, a decrease in cell proliferation mediated by the EZH2 siRNA duplex, but not by un-related duplexes, was observed (Fig. 22c). In the time frame considered (48hrs), RNA interference of EZH2 did not induce apoptosis as assessed by propidium idodide staining of nuclei or PARP cleavage. Consistent with this, the broad-spectrum caspase inhibitor, z-VAD-fmk, failed to attenuate EZH2 siRNA induced inhibition of cell proliferation (Fig. 22c). Thus, activation of the apoptosis pathway does not account for the decreases in cell number observed by RNA interference of EZH2.

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Various PcG Group proteins have been suggested to play a role in cell cycle progression (Jacobs et al., Nature 397:164 [1999]; Visser et al., Br. J. Hematol. 112:950 [2001]; Borck et al. Curr. Opin. Genet. Dev. 11:175 [2001]). Flow cytometric analysis of EZH2 siRNA-treated prostate cells demonstrated cell cycle arrest in the G2/M phase (Fig. 22d). Un-related control siRNA duplexes failed to induce a similar cell cycle dysregulation. Few apoptotic cells (sub-G1 cells) were present in any of the experimental samples tested as assessed by flow cytometry (Fig. 22d). The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that these observations suggest that EZH2 plays a role in prostate cell proliferation by mitigating the G2/M transition.

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To further understand the functional role of EZH2 in prostate cells, an epitope-tagged version of wild-type EZH2 and a deletion mutant of EZH2 missing the conserved SET domain in the eukaryotic expression vector pcDNA3 were generated (Fig. 23a). An "inducible"-version of EZH2 was also generated by creating a fusion protein with a modified murine estrogen receptor (ER) (Fig. 26a) (Littlewood *et al.*, Nuc. Acid. Res. 23:1686 [1995]; Juin *et al.*, Genes Dev. 13:1367 [1999]). EZH2-ER fusion was expressed in cells (Fig. 26b) and is inactivated, presumably by sequestration/binding to hsp90 and other proteins (Littlewood *et al.*, *supra*). Upon treatment of cells with 4-hydroxytamoxifen, hsp90 dissociates from the ER fusion and liberates its activity. Expression of the epitope-tagged EZH2 constructs was confirmed by transfection in 293 (Fig. 23b), RWPE and in other mammalian cell lines.

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PcG proteins have been proposed to mediate their functions by repression of target genes (Laible et al., supra; Jacobs et al., Semin Cell Dev. Biol. 10:227 [1999]). To begin to test this hypothesis, RWPE prostate cells were transiently transfected with wild-type EZH2 and global gene expression alterations were monitored using DNA microarrays. While RNA from the experimental (transfected) cell line was labeled with one fluorescent dye, the paired reference sample was labeled with a second distinguishable fluorescent dye. By making direct comparisons between "gene"-transfected cell lines and control vector-transfected cell lines the molecular differences between the samples were observed. When EZH2 was over-expressed in RWPE cells or SUM149 breast carcinoma cells, there was a consistent repression of a cohort of genes (Fig. 23c, d). This exclusive repression of genes was unique compared to other molecules tested in this system including c-myc and TNFR1, among others. When compared to vector-transfected cells the only gene that was significantly up-regulated in EZH2-transfected cells was EZH2 itself (Fig. 23c).

EZH2-mediated transcriptional repression was dependent on an intact SET domain (Fig. 23c), as deletion of this domain did not produce a repressive phenotype and in some cases "de-repressed" genes. EZH2 has been shown to interact with histone deacetylase 2 (HDAC2) via the EED protein (van der Vlag *et al.*, Nat. Genet. 23:474 [1999]). In the experiments described above, EZH2-mediated gene silencing was dependent on HDAC activity, as the commonly used HDAC inhibitor, trichostatin A

(TSA) completely abrogated the effects of EZH2 (Fig. 23c). Thus, EZH2 function requires both an intact SET domain as well as endogenous HDAC activity.

To identify genes that are significantly repressed by EZH2, wild-type EZH2transfected cells were compared with EZH2 .SET-transfected cells. Using this approach, 163 genes were consistently repressed while no genes were activated at an FDR of 0.0045 (Fig. 23d). Examination of the significant gene list identified the PcG group protein EPC, which is the human homolog of the drosophila protein Enhancer of Polycomb (E(Pc)) as being consistently repressed by EZH2 (Fig. 23c). Of the Drosophila PcG proteins, E(Pc) and E(z) are related in that they both act as suppressors of variegation (Su(var)) (Sinclair et al., Genetics 148:211 [1998]) and are the only PcG proteins to have yeast homologs, emphasizing the evolutionary conservation of this PcG pair. In addition to EPC, a host of other transcriptional regulators/activators were transcriptionally silenced by EZH2 including MDNA, RNF5, RNF15, ZNF42, ZNF262, ZNFN1A1, RBM5, SPIB, and FOXF2, among others (Fig. 23c). MDNA, also known as myeloid cell nuclear differentiation antigen, mediates transcriptional repression by interacting with the transcription factor YY1, which is a PcG homolog of Drosophila Pho and shown to be part of the EZH2/EED complex of proteins (Satijin et al., Mol. Cell. Biol. 21:1360 [2001]).

In addition to transcriptional repression in prostate cells, the results also support a role for EZH2 in regulating cell growth (Fig. 23). Transcriptional repression of cdc27 (two independent Unigene clones) was also observed. Cdc27 is part of the anaphase-promoting complex (APC) which mediates ubiquitination of cyclin B1, resulting in cyclinB/cdk complex degradation (Jorgensen *et al.*, Mol. Cell. Biol. 18:468 [1998]). Another family of proteins that was repressed when EZH2 was targeted was the solute carriers. At least 5 distinct members were shown to be repressed (*i.e.*, SSLC34A2, SLC25A16, SLC25A6, SLC16A2, and SLC4A3).

Example 9

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Expression of AMACR in Serum and Urine

This example describes the expression of AMACR in serum and urine. AMACR was detected by standard immunoblotting and by protein microarray using a polyclonal

rabbit anti-AMACR antibody. The results are shown in Figures 24-27. Figure 24 shows the detection of AMACR protein in PCA cell lines by quantitation of microarray data. DUCAP, DU145, and VCAP are prostate cancer cell lines. RWPE is a benign prostate cell line. PHINX is a human embryonic kidney cell line.

Figure 25 shows the detection of AMACR protein in serum by quantitation of microarray data. P1-P7 represent serum from patients with prostate cancer. NS2 and NS3 represent serum from patients that do not have PCA. SNS2 and SNS3 represent serum from patients that do not have PCA that has been spiked with AMACR protein. Figure 26 shows an immunoblot analysis of serum from patients with either negative or positive PSA antigen. Figure 27 shows an immunoblot analysis of the presence of AMACR in urine samples from patients with bladder cancer (females) or bladder cancer and incidental prostate cancer (males). The results demonstrate that AMACR can be detected in the serum and urine of patients with bladder cancer or bladder cancer and prostate cancer.

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Example 10

AMACR as a Tumor Antigen

This example describes the presence of an immune response against AMACR in serum. Figure 28 shows representative data of a humoral response by protein microarray analysis. Tumor antigens including AMACR, PSA, CEA, HSPs were spotted onto nitrocellulose coated slides. The slides were incubated with sera from different patients to detect a humoral response. The microarray was then washed. A Cy5 labeled goat antihuman IgG was used to detect the humoral response. The slide was then scanned using a microarray scanner (Axon). After data normalization, intensity of spots reflects the presence, absence or strength of humoral response to specific tumor antigen. A specific humoral response to AMACR was detected in cancer patients but not in controls. Cancer refers to sera from prostate cancer patients. BPH refers to sera from patients with benign prostate hyperplasia.

Figure 29 shows immunoblot analysis of the humoral response to AMACR.

Figure 29A shows an SDS-PAGE gel containing recombinant MBP (control protein=M) and recombinant AMACR-MBP (A) that was run and transferred to nitrocellulose paper.

Each strip blot was then incubated with human sera. A humoral response to the AMACR was detected using an HRP-conjugated anti-human antibody. Only AMACR and fragments of AMACR were detected in sera from prostate cancer patients and not in controls. Figure 29B shows a control experiment whereby the humoral response is blocked with recombinant AMACR (quenched) and thus shows the specificity of the response.

This example demonstrates that AMACR functions as a tumor antigen in human serum of prostate cancer patients. A specific immune response was generated to AMACR in the serum of PCA patients, but not in controls.

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Example 11

Expression of GP73 in Prostate Cancer

This example describes the association of GP73 with prostate cancer.

15 A. Methods

Microarray analysis, RT-PCR, Western blotting, and immunohistochemistry were performed as described in the above examples.

B. Results

Figure 30 shows GP73 Transcript levels in prostate cancer. Figure 30a shows the level of GP73 in individual samples after microarray analysis. The graph shows the values of Cy5 versus Cy3 ratio wherein the prostate cancer tissue sample RNA were labeled with Cy5 fluorescent dye, while the reference sample (pool of benign tissue RNA) sample was labeled with Cy3 fluorescent dye. A total of 76 individual experiments from different prostate tissue are plotted and they are classified as benign, prostate cancer and metastatic cancer types. Figure 30b shows the result of GP73 transcripts determined by DNA microarray analysis from 76 prostate samples grouped according to sample type and averaged. The experimental samples were labeled with Cy5 fluorescent dye, whereas the reference sample (pool of benign tissue sample) was labeled with Cy3 fluorescent dye. The box plot demonstrates the range of GP73 expression within each group. The middle horizontal bar indicates median values; the

upper and lower limits of the boxes, interquartile ranges; and the error bars, 95% confidence intervals. Figure 30c demonstrates that GP73 transcript levels are elevated in prostate cancer. RT-PCR was used to detect GP73 transcript levels in RNA preparations from prostate tissue extracts. GAPDH served as the loading control.

Figure 31 shows that GP73 protein is upregulated in prostate cancer. Figure 31a shows Western blot analysis of GP73 protein in prostate cancer. Total tissue proteins from benign, cancer and metastatic tissues (10 μg) were analyzed using anti-GP73 antiserum. β-Tubulin serves as control for sample loading. Figure 31b shows an immunoblot analysis of the Golgi resident protein Golgin 97. The Golgin 97 protein levels were analyzed in the prostate tissue sample to indicate the level of Golgi structure in normal and cancerous prostate tissue. β-Tubulin serves as control for sample loading.

Tissue microarray analysis of GP73 protein in normal and cancerous prostate tissue was also performed. GP73 protein expression was analyzed by standard biotinavidin immunohistochemical analysis using a polyclonal mouse antibody to GP73. Protein expression was evaluated on a wide range of prostate tissue using high-density tissue microarrays. High levels of staining were observed in prostate cancer tissue. Some normal epithelial cells did not stain for GP73 in a sub region of prostate cancer tissue.

Figure 32 shows immunoblot analysis of normal and prostate cancer epithelial cells. The epithelial cells were isolated from normal prostate tissue and cancer tissue to specifically isolate the protein from epithelial cell for GP73 immunoblot analysis. For this purpose, laser capture microdissected samples were used. Actin western serves as control.

25 Example 12

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Lethal Markers and Targets

This example describes the identification of lethal markers. The markers serve as potential therapeutic targets. Markers were identified by correlating the number of samples with clinical parameters and gene expression. Specifically, the present study identified markers that have an expression profile similar to EZH2, which serves as a prototypic lethal biomarker of prostate cancer. These genes were identified by a scoring

PCT/US02/24567 WO 03/012067

system that takes into account whether localized prostate cancer has recurred or not recurred. In addition, genes that have highly correlated expression with EZH2 were identified that may serve as markers to supplement EZH2.

-0.024 0.3725 0.7206	. 5			_	·	<u> </u>	<u> </u>	· ·			
-0.024 0.3725 0.7206			Total	16	13	16	6	20			
-0.306 0.1707 0.035 0 0 0 3 3 3 14 17 2506 HN1 -0.348 0.2394 0.1312 0 2 1	mean	dev	High	bph_count	pca_count	pcau_count	pcar_count	met_count	score	UNIQID	NAME
-0.348 0.2394 0.1312 0 2 1 4 14 16 5112 CSF2 0.0623 0.1578 0.3779 0 1 2 3 13 15 6053 ASNS -0.246 0.1689 0.0921 0 2 0 2 15 15 15 1520 NULL ESTS Hs.16304 -0.212 0.1386 0.0648 0 2 0 2 15 15 8273 PRC1 -0.352 0.1458 0.066 0 3 7 3 14 14 34 GPAA1 -0.232 0.2538 0.2153 0 0 1 3 10 13 5239 KIAA1691 -0.141 0.1572 0.1729 0 2 5 3 12 13 8562 NULL Human clone 23614 -0.210 1.083 0.0067 0 4 4 2 15 13 3351 FLJ11715 hypothetical protein -0.22 0.1846 0.1495 0 5 4 5 13 13 2715 NULL ESTS -0.638 0.2696 0.099 1 5 4 3 15 13 9556 FLJ12443 hypothetical protein -0.142 0.1396 0.1371 0 0 2 2 10 12 115 RTGFB -0.142 0.1606 0.1967 0 1 1 3 10 12 5292 NULL ESTS -0.4440 2474 0.0504 0 1 2 2 11 12 3689 NUF2R hypothetical protein -0.205 0.2362 0.2674 0 2 1 2 12 12 1219 ABCC5 -0.090 0.214 0.3526 0 4 2 4 12 12 1360 MEN1 -0.087 0.3367 0.201 0 1 4 2 10 11 3747 H2BFB -0.0146 0.1486 0.1307 0 2 2 1 11 12 3660 MEN1 -0.164 0.1486 0.3367 0.201 0 1 4 2 10 11 3747 H2BFB -0.0164 0.1486 0.3367 0.201 0 1 4 2 10 11 3747 H2BFB -0.0255 0.1542 0.3338 0 3 3 3 11 11 256 EZH2 -0.031 0.1826 0.3346 0 4 4 2 13 11 1979 NULL ESTS Hs.268921 -0.048 0.2967 0.1131 0 2 0 2 10 10 906 MGC5627 hypothetical protein -0.248 0.1486 0.3367 0.201 0 2 2 10 10 906 MGC5627 hypothetical protein -0.0240 0.2530 0.421 0.0411 0 2 8 2 10 10 3728 NULL ESTS -0.0130 0.1826 0.3346 0 4 2 13 11 1979 NULL ESTS Hs.268921 -0.048 0.2967 0.1131 0 2 0 2 10 10 906 MGC5627 hypothetical protein -0.0130 0.1826 0.3346 0 4 2 13 11 1979 NULL ESTS Hs.268921 -0.048 0.2967 0.1131 0 2 0 2 10 10 906 MGC5627 hypothetical protein -0.0130 0.1826 0.3346 0 4 2 13 11 1979 NULL ESTS -0.0130 0.1826 0.3346 0 4 2 2 10 10 978 NULL ESTS -0.0130 0.1826 0.3346 0 4 2 2 10 10 978 NULL ESTS -0.0130 0.1826 0.3346 0 4 9 3928 DGKD -0.0130 0.1826 0.2079 0 2 2 2 10 10 9 3928 DGKD -0.01079 0.1132 0.043 0 4 3 3 10 9 9 7193 KIAA0602	-0.024	0.3725	0.7206	0	4	. 5	6	16	18	5814	NULL ESTs Hs.30237
1	-0.306	0.1707	0.0351	o	0	3	3	14	17	2506	HN1
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0.212 0.1386 0.0648 0 2 0 2 15 15 8273 PRC1 -0.352 0.1458 0.06 0 3 7 3 14 14 34 GPAA1 -0.292 0.2538 0.2153 0 0 1 3 10 13 5239 KIAA1691 -0.141 0.1572 0.1729 0 2 5 3 12 13 8562 NULL Human clone 23614 -0.210 0.1083 0.0067 0 4 4 2 15 13 3351 FLJ11715 hypothetical protein -0.220 0.1846 0.1495 0 5 4 5 13 13 2715 NULL BSTs -0.638 0.2696 0.099 1 5 4 3 15 13 9556 FLJ12443 hypothetical protein -0.142 0.1396 0.1371 0 0 2 2 10 12 1158 TGFBI -0.124 0.1606 0.1967 0 1 1 3 10 12 5292 NULL ESTS -0.444 0.2474 0.0504 0 1 2 2 11 12 3689 NUF2R hypothetical protein -0.205 0.3362 0.2674 0 2 1 2 12 12 1360 MEN1 -0.241 0.1541 0.0673 0 5 3 2 15 12 8476 SARM and HEAT/Armadillo moti -0.874 0.3367 0.201 0 1 4 2 10 11 3747 H2BFB -0.196 0.254 0.3122 0 2 1 3 10 11 4941 VAV2 -0.166 0.1486 0.1307 0 2 4 2 11 11 8636 NULL ESTS Hs.23268 -0.025 0.2536 0.2812 0 4 3 4 11 11 2156 EZH2 -0.031 0.1826 0.3346 0 4 4 2 13 11 11 11 11 11 -0.243 0.1421 0.0411 0 2 8 2 10 10 3728 NULL ESTS Hs.268921 -0.048 0.2967 0.1131 0 2 2 2 2 10 10 3728 NULL ESTS -0.015 0.1806 0.2579 0 2 2 2 2 10 10 3728 NULL ESTS -0.016 0.1806 0.2579 0 2 2 2 2 10 10 3728 NULL ESTS -0.017 0.0617 0 3 2 2 10 9 3928 DGKD -0.0190 0.1132 0.3343 0 3 2 2 10 9 3928 DGKD -0.0107 0.1132 0.3343 0 4 3 3 10 9 7193 KIAA0602	0.0623	0.1578	0.3779	: 0	1	2	3	13	15	6053	ASNS
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0.292 0.2538 0.2153 0	-0.212	0.1386	0.0648	O	2	0	2	15	15	8273	PRC1
0.141 0.1572 0.1729 0 2 5 3 12 13 8562 NULL Human clone 23614 -0.21 0.1083 0.0067 0 4 4 2 15 13 3351 FLJ11715 hypothetical protein -0.22 0.1846 0.1495 0 5 4 5 13 13 2715 NULL ESTS -0.638 0.2696 -0.099 1 5 4 3 15 13 9556 FLJ12443 hypothetical protein -0.142 0.1396 0.1371 0 0 2 2 10 12 1158 TGFBI -0.124 0.1606 0.1967 0 1 1 3 10 12 5292 NULL ESTS -0.444 0.2474 0.0504 0 1 2 2 11 12 3689 NUF2R hypothetical protein -0.205 0.2362 0.2674 0 2 1 2 12 1219 ABCCS -0.09 0.2214 0.3526 0 4 2 4 12 12 1360 MEN1 -0.241 0.1541 0.0673 0 5 3 2 15 12 8476 SARM and HEAT/Armadillo motion -0.874 0.3367 -0.201 0 1 4 2 10 11 3747 H2BFB -0.196 0.254 0.3122 0 2 1 3 10 11 4941 VAV2 -0.166 0.1486 0.1307 0 2 4 2 11 11 8636 NULL ESTS Hs.23268 -0.2055 0.1542 0.3338 0 3 3 3 11 11 280 TOP2A -0.226 0.2536 0.2812 0 4 3 4 11 11 2156 EZH2 -0.031 0.1826 0.3346 0 4 2 13 11 1979 NULL ESTS Hs.268921 -0.48 0.2967 0.1131 0 2 8 2 10 10 3728 NULL ESTS -0.133 0.1806 0.2279 0 2 2 2 10 10 3728 NULL ESTS -0.133 0.1806 0.2279 0 2 2 2 10 10 3728 NULL ESTS -0.0107 0.0179 0.1132 0.3343 0 3 2 2 10 9 3928 DGKD -0.01079 0.1132 0.3343 0 3 2 2 10 9 3928 DGKD -0.01079 0.1132 0.3343 0 4 3 3 10 9 7193 KIAA0602 -0.288 0.1221 0.043 0 4 3 3 10 9 7193 KIAA0602 -0.288 0.1221 0.043 0 4 3 3 10 9 7193 KIAA0602	-0.352	0.1458	-0.06	0	3	. 7	3	14	14	34	GPAA1
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0.124 0.1606 0.1967	-0.638	0.2696	-0.099	1	. 5	4	3	15	13	9556	FLJ12443 hypothetical protein
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-0.241 0.1541 0.0673	-0.205	0.2362	0.2674	0	. 2	1	2	12	12	1219	ABCC5
-0.874 0.3367 -0.201	-0.09	0.2214	0.3526	0	4	2	4	12	12	1360	MEN1
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-0.166 0.1486 0.1307	-0.874	0.3367	-0.201	o	1	· 4	2	10	11	3747	H2BFB
0.0255 0.1542 0.3338	-0.196	0.254	0.3122	0	2	1	3	10	11	4941	VAV2
-0.226 0.2536 0.2812 0 4 3 4 11 11 2156 EZH2 -0.031 0.1826 0.3346 0 4 4 2 13 11 1979 NULL ESTs Hs.268921 -0.48 0.2967 0.1131 0 2 0 2 10 10 906 MGC5627 hypothetical protein -0.243 0.1421 0.0411 0 2 8 2 10 10 3728 NULL ESTs -0.133 0.1806 0.2279 0 2 2 2 10 10 8759 RAB24 -0.192 0.1782 0.1645 0 3 2 2 11 10 2029 FLJ12876 hypothetical protein -0.617 0 -0.617 0 3 2 2 10 9 3928 DGKD -0.1079 0.1132 0.3343 0 3 2 2 10 9 5372 ODF2 -0.288 0.1221 -0.043 0 4 3 3 10 9 7193 KIAA0602	-0.166	0.1486	0.1307	0	2	4	2	11	11	8636	NULL ESTs Hs.23268
-0.031 0.1826 0.3346	0.0255	0.1542	0.3338	0	3	3	3	11	11	280	TOP2A
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-0.243 0.1421 0.0411 0 2 8 2 10 10 3728 NULL ESTS -0.133 0.1806 0.2279 0 2 2 10 10 8759 RAB24 -0.192 0.1782 0.1645 0 3 2 2 11 10 2029 FLJ12876 hypothetical protein -0.617 0 -0.617 0 3 2 2 10 9 3928 DGKD 0.1079 0.1132 0.3343 0 3 2 2 10 9 5372 ODF2 -0.288 0.1221 -0.043 0 4 3 3 10 9 7193 KIAA0602	-0.031	0.1826	0.3346	0	4	4	2	13	11	1979	NULL ESTs Hs.268921
-0.133 0.1806 0.2279 0 2 2 2 10 10 8759 RAB24 -0.192 0.1782 0.1645 0 3 2 2 11 10 2029 FLJ12876 hypothetical protein -0.617 0 -0.617 0 3 2 2 10 9 3928 DGKD 0.1079 0.1132 0.3343 0 3 2 2 10 9 5372 ODF2 -0.288 0.1221 -0.043 0 4 3 3 10 9 7193 KIAA0602	-0.48	0.2967	0.1131	0	2	o	2	10	10	906	MGC5627 hypothetical protein
-0.192 0.1782 0.1645 0 3 2 2 11 10 2029 FLJ12876 hypothetical protein -0.617 0 -0.617 0 3 2 2 10 9 3928 DGKD 0.1079 0.1132 0.3343 0 3 2 2 10 9 5372 ODF2 -0.288 0.1221 -0.043 0 4 3 3 10 9 7193 KIAA0602	-0.243	0.1421	0.0411	0	2	8	2	10	_10	3728	NULL ESTs
-0.617 0 -0.617 0 3 2 2 10 9 3928DGKD 0.1079 0.1132 0.3343 0 3 2 2 10 9 5372 ODF2 -0.288 0.1221 -0.043 0 4 3 3 10 9 7193 KIAA0602	-0.133	0.1806	0.2279	0	2	2	2	10	_10	8759	RAB24
0.1079 0.1132 0.3343	-0.192	0.1782	0.1645	0	3	2	2	11	_10	2029	FLJ12876 hypothetical protein
0.1079 0.1132 0.3343	-0.617	0	-0.617	0	3	2	2	10	. 9	3928	DGKD
-0.288 0.1221 -0.043 0 4 3 3 10 9 7193 KIAA0602	0.1079			0	3	2	2	10	9	5372	ODF2
				0	4	3	3	10	9	7193	KIAA0602
				0	4	2	2	11	9	8535	EHM2

-0.95	0.3504	-0.249	0	4	2	2	- 11	. 9	9824	SLC19A1		
-0.314	0.187	0.06	1	4	2	2	11	9	9447	LIG1		
0.1366	0.1883	0.5132	1	4	3	2	10	8	. 327	NULL ESTs		
-0.586	0.2952	0.0044	0	. 5	2	2	. 11	8	1269	DGKZ		-: :

mean: mean expression in BPH Dev: standard deviation in BPH

High: 2 SD's above the mean (threshold)

5 Bph: # of BPH samples > thresh

PCA: # of PCA samples > thresh (>1yr no recur)
Pcau: # of PCA samples > thresh (<1yr followup)

Pcar: # of PCA samples > thresh (recur) Met: # of metastatic samples > thresh

10 Score: = met + pcar - pca Total: # of samples in category

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Exemplary lethal markers identified using the above methods include ABCC5 (MDR5). This multi-drug resistance gene actively pumps cyclic nucleotides and other small molecules out of cells. An unrelated study found that this enzyme is potently Inhibited by phosphodiesterase inhibitors, including sildenafil (viagra). The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not required to practice the present invention. Nonetheless, it is contemplated that sildenafil may be useful in the treatment of aggressive PCA.

Another lethal marker identified is asparagine synthetase (ASNS). Current therapeutics for the inhibition of ASNS include asparaginase, an enzyme that destroys asparagine in the body. It has been shown that cancers expressing the synthetase are resistant. Analogs are being developed to inhibit the synthetase.

Top2A (topoisomerase 2) and the Vav2 Oncogene were also identified using the methods of the present invention. Vav2 is required for cell spreading, but is dependent on src. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not required to practice the present invention.

Nonetheless, it is contemplated src inhibitors can stop vav2 mediated cell spreading

This example describes the identification of cancer markers overexpressed in prostate cancers. The present invention is not limited to a particular mechanism. Indeed, an understanding of the mechanism is not necessary to practice the present invention.

Nonetheless, it is contemplated that therapeutic compounds that inhibit these lethal markers are useful in the treatment of prostate cancer.

Example 13

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5 Characterization of Annexin Expression in Prostate Cancer

This Example describes the expression of Annexins in prostate cancer.

A. Materials and Methods

Prostate Sample Collection

Prostate tissues were taken from the radical prostatectomy series and the rapid autopsy program available through the University of Michigan Prostate Cancer Specialized Program of Research Excellence (S.P.O.R.E.) Tissue Core. This program is approved by Institutional Review Board at the University of Michigan.

Hormone naïve, clinically localized PCA samples used for this study were taken from a cohort of men who underwent radical retropubic prostatectomy as a monotherapy (i.e., no hormonal or radiation therapy) for clinically localized PCA between the years 1994 and 1998. Processing of the prostatic tissues started within 20 minutes after surgical resection. The prostates were partially sampled and approximately 50% of the tissue was used for research. This protocol has been evaluated in a formal study to assure that partial sampling does not impair accurate staging and evaluation of the surgical margins (Hollenbeck et al., J. Urol. 164:1583 [2000]). The snap frozen samples used for cDNA expression array analysis were all evaluated by one of the study pathologists. All samples were grossly trimmed to ensure greater than 95% of the sample represented the desired lesion.

Hormone refractory PCA samples were collected from the rapid autopsy program (Rubin et al., [2000], supra). Snap frozen samples were used for cDNA expression array analysis. Mirrored samples from the same lesion were placed in 10% buffered formalin. The fixed samples are embedded in paraffin. As with the prostatectomy samples, the study pathologist reviewed the glass slides, circled areas of viable prostate cancer, avoiding areas of necrosis, and used these slides as a template for tissue microarray construction. In this study, twenty (20) hormone refractory metastatic PCAs were

extracted from 15 rapid autopsy cases performed from 1997 to 2000. The patients' ages ranged from 53 to 84 and time from diagnosis to death ranged from 21 to 193 months. All 15 patients died with widely metastatic PCA after extensive treatment, which included antiandrogens and chemotherapy.

Prostatectomy samples were evaluated for the presence or absence of surgical margin involvement by tumor (surgical margin status), the presence of extraprostatic extension, and seminal vesicle invasion. Tumors were staged using the TNM system, which includes extraprostatic extension and seminal vesicle invasion but does not take into account surgical margin status (Bostwick *et al.*, Semin. Urol. Oncol. 17:222 [1999]). Tumors were graded using the Gleason grading system (Gleason, [1966], *supra*).

Immunohistochemistry

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After paraffin removal and hydration, the tissue microarray slides were immersed in 10 mM citrate buffer placed in a pressure cooker chamber and microwaved for 10 minutes for optimal antigen retrieval. Immunostaining was performed using a Dako autostainer (DAKO, Carpinteria, CA). The primary antibody was incubated for 45 minutes at room temperature and a secondary biotin-labeled antibody for 30 minutes. Streptavidin-LSA amplification method (DAKO K0679) was carried out for 30 minutes followed by peroxidase/diaminobenzidine substrate/Chromagen. The slides were counterstained with hematoxylin. Polyclonal antibodies directed against the N-terminus of annexin 1(dilution 1:50), annexin 2 (dilution 1:100), annexin 4 (dilution 1:100), annexin 7 (dilution 1:500), and annexin 11 (dilution 1:100) were obtained from a signal source (Santa Cruz Biotechnology, Santa Cruz, CA). Protein expression as determined by two pathologists immunohistochemistry was scored as negative (score=1), weak (score 2), moderate (3) or strong (4), using the system described above.

Tissue Microarray Construction, Digital Image Capture, and Analysis

Tissue microarrays were constructed as previously described to evaluate protein expression in a wide range of samples ranging from benign prostate tissue taken from the prostatectomy samples to hormone refractory PCA. Three tissue microarrays were used for this study consisting of benign prostate, localized PCAs, and hormone refractory

PCA. The tissue microarrays were assembled using the manual tissue arrayer (Beecher Instruments, Silver Spring, MD) as previously described (Kononen et al., [1998], supra; Perrone et al., [2000], supra). Tissue cores from the circled areas of interest were targeted for transfer to the recipient array blocks. The 0.6mm diameter tissue microarray cores were each spaced at 0.8mm from core-center to core-center. Tissue microarray images were acquired using the BLISS Imaging System (Bacus Lab, Lombard, IL).

Statistical Analyses

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To investigate the statistical significance associated with the differential expression of annexins across 4 independent gene expression studies, standard methods (Hedges et al., Statistical Methods for Meta-analysis meta-analysis. Orlando, Academic Press 1985, pp xxii, 369) were used to combine the results. For each of the studies, a t-statistic was computed (with the two groups being benign tissue compared against localized prostate cancer) and the associated p-values were transformed using a negative logarithmic transformation. These numbers were then doubled and added together to arrive at a summary measure of differential gene expression across the three studies. To assess the statistical significance associated with this summary measure, a permutation-based approach was adopted (Hedges et al., supra). Namely, the tissue types were permutated within studies, and the summary measure was computed for the permutated data. A p-value was computed using the permutation distribution of the summary measure. The issue then arises of whether or not the t-statistics from the three studies are comparable.

Annexin protein expression was statistically evaluated using the mean score results from each tissue microarray sample for each prostate tissue type (*i.e.*, benign, localized PCA, and hormone refractory PCA). To determine differences between all pairs (*e.g.*, localized prostate cancer versus benign), an ANOVA with a post-hoc analysis was performed using the Scheffé method (Scheffae *et al.*, *supra*). The mean expression scores for all examined cases were presented in a graphical format by using error-bars with 95% confidence intervals.

B. Results

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Expression array analysis revealed a significant dysregulation of annexin family members with PCA progression. The cDNA expression of annexins 1, 2, 4, 7 and 11 were significantly decreased in the hormone refractory PCA samples as compared to localized hormone sensitive PCA samples with 2.2, 1.5, 1.3, 1.4 and 1.8 fold decrease, respectively (all p-values < 0.01) (Table 3 and Figure 33). Annexins 1 and 4 showed significant decreases of mRNA expression in localized PCA samples as compared to the benign samples. There were no significant differences between localized hormone naive PCA and the benign samples for annexin 2, 7, and 11. No cDNA dysregulation between the tested prostate samples and annexins 8 and 13 was observed. Annexin 6 demonstrated a slight decrease in cDNA expression between localized PCA and benign samples, which was not statistically significant (Table 3).

In order to cross validate the cDNA expression results for these annexin family members, a meta-analysis of gene expression was performed. Annexin family members cDNA expression results were evaluated using a series of data sets (Welsh *et al.*, Cancer Res. 61:5974 [2001]; Luo *et al.*, Cancer Res. 61:4683 [2001]; Magee *et al.*, Cancer Res. 61:5692 [2001]). The analysis evaluated annexins for each of the individual studies as well as performing a summary statistic, taking into account the significance of the gene expression across the 4 studies. The meta-analysis compared differences between clinically localized PCA and benign prostate tissue as not all of the studies had hormone refractory metastatic PCA. The meta-analysis (Table 4 and Figure 34) demonstrated that annexins 1, 2, 4, and 6 were significantly down regulated across independent studies. Annexin 6 was down regulated to a significant level in 4 of 4 studies. Annexin 1 demonstrated down regulation in 3 of 4 studies. Annexins 2 and 4 were down regulated in 2 studies and overall considered to be significantly under expressed by the meta-analysis. Annexin 7 was not found to be significantly under expressed in any of the 4 studies at the transcript level.

Immunohistochemistry was performed to confirm these results at the protein level (Table 5). By immunohistochemistry, a significant decrease in protein expression for annexins 1, 2, 4, 7 and 11 in hormone refractory PCA samples as compared to localized PCA samples was identified with 2.5 (3.8 vs. 1.5 median expression), 2.4 (4 vs. 1.7

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median expression), 3.6 (4 vs. 1.1 median expression) and 3.3 (4 vs. 1.2 median expression) fold decreases, respectively (Kruskal Wallis test, all p-values p< 0.05). No statistically significant differences were seen between benign and localized PCA samples in any of the annexins tested.

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Table 3: Gene Expression of Select Annexins.

Annexin	Benig	n	BPH ¹		Loc- F	CA ²	Met-P	CA ³	Ratio	p :
	Count	Median	Count	Median	Count	Median	Count	Median	PCA/Met	Value*
1	5	1.56	16	1.35	16	0.69	20	0.31	2.23	<0.001
2	5	0.79	16	0.69	16	0.74	20	0.49	1.51	0.009
4	5	0.91	16	0.97	16	0.9	20	0.69	1.30	0.001
6 ·	5	1.2	16	1.29	16	1.05	20	1.15	0.91	0.377
7	5 .	0.8	16	0.88	16	0.88	20	0.62	1.42	<0.001
8 -	5	1.14	16	1.06	16	0.99	20	1.19	0.83	0.156
11	5	0.99	16	0.76	16	0.94	20	0.52	1.81	<0.001
13	5	1.08	16	1.35	16	1.03	20	0.94	1.10	0.393

^{*} Kruskal Wallis Test. 1, BPH, benign prostatic hyperplasia. 2, Loc-PCA, localized prostate cancer. 3, Met-PCA, metastatic hormone refractory prostatic cancer. Ratio PCA/Met, ratio of expression of localized PCA over hormone refractory PCA.

Table 4: Meta-Analysis of cDNA Prostate Gene Expression Studies for Annexin Family Members

Annexin	Present study	Welsh et al.	Luo et al.	Magee et al.	Summary p-Value
. 6	0.024	0.0001	0.0001	0.026	0.0001
1	0.0001	0.031	0.0007	0.23	0.0001
2	NA	0.0001	NA	0.002	0.0001
11	ŅA	0.010	NA	0.6	0.17
7	0.25	0.48	0.38	0.088	0.20
4	0.33	0.023	0.0093	0.58	0.011

-	13	0.177	NA	1.00	NA	0.48
Ì	8	0.79	NA	0.104	NA	0.29

Table 5: Tissue Microarray Protein Expression for Annexins by Tissue Type

Annexin	Benign		Loc-PO	CA^2	A ² Met-PCA ³		PCA/M	p-value*
	Count	Media	Count	Media	Count	Media	ET	
		n		n		n		
1	37	2.59	360	2.45	162	1.46	1.68	<0.001
2	57	3.95	82	3.62	214	1.47	2.46	<0.001
4	23	3.65	357	3.96	141	1.57	2.52	<0.001
7	26	3.77	350	3.97	126	1.32	3.01	<0.001
11	23	4.00	360	3.99	163	1.30	3.01	<0.001

^{*} Kruskal Wallis Test. 1, BPH, benign prostatic hyperplasia. 2, Loc-PCA, localized prostate cancer. 3, Met-PCA, metastatic hormone refractory prostatic cancer.

Example 14

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Association of CtBP with Prostate Cancer

This example describes the expression of C-terminal binding proteins 1 and 2 (CtBP1 and CtBP2) in prostate cancer. Microarray analysis, Western Blots, immunohistochemistry, and statistical analysis were performed as described in the above examples.

The CtBP transcript was found to be up-regulated in metastatic prostate cancer (Figure 38). Tissue extracts were used to validate this finding at the protein level using an antibody that recognizes CtBP1 and CtBP2 (Sewalt *et al.*, Mol. Cell. Biol. 19:777 [1999]. The results are shown in Figure 35. Figure 35 shows the Expression of CtBP proteins in PCA specimens. Extracts from selected prostate specimens were assessed for expression of CtBP and PcG proteins by immunoblot analysis. Protein level was equalized in each extract before loading and blots were stained with Ponceau S to confirm equal loading. β-tubulin was used as a control protein.

Both CtBPs were over-expressed in metastatic prostate cancer relative to localized prostate cancer and benign tissue. EZH2 protein was also elevated in metastatic prostate cancer relative to localized prostate cancer or benign prostate (Figure 35). EED, a PcG protein that forms a complex with EZH2, along with an un-related protein, β-tubulin, did not exhibit similar protein dysregulation. Thus, both transcriptional repressors (CtBP and EZH2) are mis-expressed in metastatic prostate cancer.

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To determine in situ expression of CtBP, immunohistochemistry of prostate tissue sections were performed using prostate tissue microarrays. Benign prostatic epithelia exhibited exclusively nuclear staining consistent with CtBP's role as a transcriptional repressor. Both clinically localized and metastatic prostate cancer exhibited nuclear staining as well. Most of the metastatic prostate cancer cases and a fraction of the localized prostate cancer cases exhibited distinct cytoplasmic staining of CtBP.

Figure 36 shows tissue microarray analysis of CtBP in prostate cancer that suggests mis-localization during prostate cancer progression. The mean CtBP protein expression for the indicated prostate tissues and sub-cellular compartment is summarized using error bars with 95% confidence intervals. Figure 37 shows the sub-cellular fractionation of LNCaP cells. The results show an increased level of CtBP1 in the cytoplasm relative to the nucleus. CtBP2 is weakly expressed in the cell lines and is not easily apparent. β-tubulin, which is not expressed in the nucleus, is provided as a control. Figure 38 shows a Kaplan-Meier Analysis of prostate cancer tissue microarray data. The results demonstrate that the presence of cytoplasmic CtBP may be associated with a poorer clinical outcome. The median follow up time for all patients was 1 year (range 2 month to 6.5 years). Over this follow up time, 38% of the patients developed a recurrence or PSA elevation greater than 0.2 ng/ml. Prostate tumors from 97 patients demonstrated near uniform nuclear protein expression for CTBP. Cytoplasmic expression was variable with 85 of 97 cases (88%) demonstrating weak cytoplasmic staining and 12 (12%) with moderate to strong CTBP expression. There was a significant association with increased CTBP cytoplasmic staining intensity and PSA recurrence or presence of recurrent disease following prostatectomy with a relative risk of 1.7 (Cox regression analysis p=0.034). The data presented demonstrates a Kaplan-Meier Analysis of outcome stratified by negative/weak cytoplasmic CTBP staining and moderate/strong

staining. CTBP cytoplasmic expression predicted recurrence even when Gleason score was taken into account in a multivariable model, suggesting that CTBP is a prognostic predictor of poor outcome [Gleason relative risk 1.4 (p=0.005) and cCTBP rr 1.6 (p=0.042)].

CtBP has been shown to bind nitric oxide synthase (NOS), which is thought to shift the localization of CtBP from the nuclear compartment to the cytoplasmic compartment (Riefler et al., J. Biol. Chem. 276:48262 [2001]). Weigert and colleagues have proposed a cytoplasmic role for CtBP in the induction of Golgi membrane fission (Weigart et al., Nature 402:429 [1999]). To further support the preliminary immunohistochemical findings, LNCaP (metastatic) prostate cancer cells were fractionated and it was found that CtBP levels were higher in the cytosol relative to the nucleus (Figure 38).

Example 15

15 Methods of Characterizing Cancer Markers

This example describes exemplary methods for the characterization of new cancer markers of the present invention. These methods, in combination with the methods described in the above examples, are used to characterized new cancer markers and identify new diagnostic and therapeutic targets.

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A. Determination of quantitative mRNA transcript levels of cancer markers in prostate cancer specimens

In some embodiments, markers revealed to be over or under expressed in cancer microarrays (See e.g., Example 1 for a description of microarrays) are quantitated using real-time PCR (Wurmbach et al., J. Biol. Chem. 276:47195 [2001]).

In preferred embodiments, cDNA from over 100 prostate samples for archived cDNA samples and associated clinical data are available (See Example 1). The level of expression in the microarray is compared to those obtained by real-time PCR. To identify genes with dysregulation of expression, real-time PCR analysis of cDNA generated from laser-capture microdissected prostate cancer epithelia and benign epithelia is performed.

B. Detection of Mis-localized Transcripts

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In some embodiments, in order to determine if a cancer marker normally present in the nucleus of a cell (e.g., a transcriptional repressor) is mis-localized to the cytoplasm (or other mis-locations) in cancer, the expression of the marker is examined in tissue extracts from preferably at least 20 benign prostate samples, 20 prostate cancer specimens, and 20 metastatic prostate specimens. Expression of the marker in benign prostate cell lines (RWPE), primary prostatic epithelial cells (Clonetics, Inc.) and a panel of prostate cancer cells including LNCaP, DU145, PC3, DUCaP, and VCaP cells is also examined. Once overall expression of prostate cell lines and tissues is established, the cellular localization of the marker is determined by 2 methods. In the first method, the cell and tissue extracts are fractionated into a nuclear fraction and a cytosolic fraction (NE-PER, Pierce-Endogen; Orth et al., J. Biol. Chem. 271:16443 [1996]). Quantitated protein is then analyzed by immunoblotting. Relative levels of cytosolic and nuclear cancer marker are determined by densitometry. To verify clean fractionation, antibodies to β-tubulin and PCNA (or lamin A) are used to assess cytosolic and nuclear fractions, respectively.

In the second method, cells are immunostained with antibodies to the cancer marker followed by detection using anti-rabbit FITC secondary antibody. Confocal microscopy (U of M Anatomy and Cell Biology Core Facility) is used to examine *in situ* localization of the cancer markers.

In some embodiments, mis-localization is further investigated by sequencing the gene in cells containing the mis-located transcript (e.g., metastatic cases) for mutations.

C. Correlation of cancer markers with clinical outcome

In some preferred embodiments, the association of expression or mis-localization of a cancer marker with clinical outcome is investigated. The ratio of total cancer marker to β-tubulin by immunoblot analysis of prostate cancer tissue extracts is first determined and associated with clinical outcome parameters. For markers suspected of being mislocalized in cancer (e.g., CtBP), the ratio of cytoplasmic marker to nuclear marker is next determined by immunoblot analysis of prostate cancer tissue extracts and associated with clinical outcome parameters. For example, it is contemplated that a high

cytoplasmic/nuclear cancer marker ratio may portend a poor clinical outcome. In some embodiments (e.g., where a cancer marker is suspected of being mis-localized), immunohistochemistry of prostate cancer tissue microarrays is used to determine whether the presence of cytoplasmic marker correlates with poor clinical outcome. Tissue microarrays are prepared and performed as described in the above examples.

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Briefly, high-density tissue microarrays (TMA) are constructed as previously described (Perrone et al, supra; Kononen et al., supra). Immunostaining intensity is scored by a genitourinary pathologist as absent, weak, moderate, or strong (or alternatively analyzed separately as for cytoplasmic and nuclear staining). Scoring is performed using a telepathology system in a blinded fashion without knowledge of overall Gleason score (e.g., tumor grade), tumor size, or clinical outcome (Perrone et al., supra). Tumor samples are derived from patients with clinically localized, advanced hormone refractory prostate cancer and naïve metastatic PCA. Cases of clinically localized prostate cancer are identified from the University of Michigan Prostate S.P.O.R.E. Tumor Bank. All patients were operated on between 1993 and 1998 for clinically localized prostate cancer as determined by preoperative PSA, digital-rectal examination, and prostate needle biopsy. All tissues used are collected with institutional review board approval. The advanced prostate tumors are collected from a series of 23 rapid autopsies performed at the University of Michigan on men who died of hormone refractory prostate cancer. The clinical and pathologic findings of these cases have been reported (Rubin et al., [2000], supra).

Statistical analysis of the array data is used to correlate the cancer marker protein measurements on the TMA with clinical outcomes, such as time to PSA recurrence and survival time. This analysis involves survival analysis methods for correlating the measurements with these censored response times. Kaplan-Meier curves are plotted for descriptive purposes. Univariate analyses is performed using the Cox model associating the biomarker with the survival time. In addition, multivariate Cox regression analysis is performed to test whether the biomarker adds any prognostic information over and above that available from known prognostic markers (*i.e.*, Gleason score, tumor stage, margin status, PSA level before surgery).

D. RNA Interference

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In some embodiments, RNA interference of cancer markers is used to investigate the role of the cancer marker in cell culture and well as for application as a therapeutic cancer treatment (See e.g., Example 8 for an example of RNA interference). 21nucleotide RNAs (siACE-RNAi) are synthesized through a commercial vendor (Dharmacon Research, Inc.). RNA interference has been used in mammalian cells (Elbashir et al., Nature 411:494 [2001]). Several siRNA duplexes and controls are designed for each marker. The design of the siRNA duplexes uses criteria provided by Elbashir et al. (Elbashir et al., supra) and Dharmacon Research which include: starting approximately 75 bases downstream of the start codon, locating an adenine-adenine dimer, maintaining G/C content around 50%, and performing a BLAST-search against EST databases to ensure that only one gene is targeted. Multiple (e.g., two) siRNA duplexes are designed for each molecule of interest since whether the siRNA duplex is functional is a relatively empirical process. In addition, it is contemplated that using two siRNA duplexes may provide a combined "knock-down" effect. As a control, a "scrambled" siRNA, in which the order of nucleotides is randomized, is designed for each molecule of interest. Oligonucleotides are purchased deprotected and desalted. Upon arrival, the oligonucleotides are annealed to form a duplex using the manufacturer's provided protocol.

To test the efficacy of each siRNA duplex, prostate cell lines (RWPE, DU145, LnCAP, and PC3) are transfected with the OLIGOFECTAMINE reagent as described (Elbashir et al., supra). The cells are assayed for gene silencing 48 hrs post-transfection by immunoblotting with respective antibodies. A number of controls are included: buffer controls, sense siRNA oligo alone, anti-sense siRNA oligo alone, scrambled siRNA duplex, and siRNA duplexes directed against unrelated proteins. If significant silencing is not appreciated after single transfection, sequential transfection is performed and inhibition is monitored at later time points (i.e., 8 days later) as suggested by others (Breiling et al., Nature. 412: 51 [2001]). This may be necessary with proteins that have a long half-life.

In addition to the transient expression of siRNAs, a method for stable expression of siRNAs in mammalian cells is used (Brummelkamp et al., Science 296:550 [2002]).

Prostate cancer cell lines are generated that express siRNA targeting cancer markers using the pSUPER system. Scrambled siRNA is used as a control. The cell lines facilitate downstream characterization of cancer markers that may be cumbersome using duplexes transiently. If inhibition of a specific cancer marker is found to be toxic to cells, the pSUPER cassette containing siRNA to the marker is cloned into an inducible vector system (e.g., Tet on/off).

E. Generation of Mutants.

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To study the function of cancer markers of the present invention, mutants of cancer markers are generated in eukaryotic expression vectors. myc-epitope tagged versions of cancer marker mutants are generated in both pcDNA3 and pcDNA3-ER (a modified estrogen receptor ligand binding domain). In the case of the ER constructs, the vectors produce an in-frame fusion protein with modified ER, thus generating a post-transcriptionally inducible vector (Littlewood *et al.*, Nucleic Acids Res. 23: 686 [1995]). The ER-ligand domain is mutated and fails to bind endogenous estrogen, yet can be activated by 4-hydroxytamoxifen (Littlewood *et al.*, *supra*). The ER-fusion proteins are inactivated in the absence of ligand presumably due to binding of proteins such as hsp90. In the presence of exogenously added 4-hydroxytamoxifen, ER-fusions become liberated. By using an inducible vector system, cell lines expressing a "toxic" or growth inhibitory version of a cancer marker can still be isolated.

Various N-terminal and C-terminal deletion mutants are generated that encompass function domains of the cancer marker (e.g., the PXDLS, dehydrogenase, and PDZ binding domains of CtBP; Chinnadurai, Mol Cell. 9: 213 [2002]). It is contemplated that some of the mutant versions of the cancer markers of the present invention act as dominant negative inhibitors of endogenous cancer marker function. Expression of epitope-tagged cancer markers and mutants is assessed by transient transfection of human embryonic kidney cells (using FUGENE) and subsequent Western blotting.

F. Establishing Stable Cell Lines Expressing Cancer Markers And Mutants

In some embodiments, cell lines stably expressing cancer markers of the present invention are generated for use in downstream analysis. FUGENE is used to transiently transfect prostate cell lines (RWPE, DU145, LnCAP, and PC3) with cancer markers and

fusions or mutants using the above mentioned vectors and appropriate G418 selection. Prostate cell lines with varied expression levels of endogenous cancer marker protein are used. Both individual clones and pooled populations are derived and expression of cancer markers and mutants assessed by immunoblotting for the epitope tag. By also using an inducible system, clones expressing toxic versions of cancer markers or mutants can be isolated.

G. Cell proliferation and apoptosis studies

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In some embodiments, the role of cancer marker expression in prostate cell proliferation is investigated using a multi-faceted approach that includes 1. RNA interference, 2. transient transfection of cancer markers and potential dominant negative mutants, and 3. comparing stable transfectants of cancer markers and mutants. The following predictions are tested using these methods: 1. whether inhibition of cancer markers will block cell growth and 2. whether overexpression of cancer markers will enhance cell proliferation.

Cell proliferation is assessed by cell counting (Coulter counter) over a time course in culture by using the WST-1 reagent (Roche, Inc.), which is a non-radioactive alternative to [³H]-thymidine incorporation and analogous to the MTT assay. The rate of incorporation of the DNA labeling dye bromodeoxyuridine (BrdU) will also be measured as described previously (Jacobs *et al.*, Nature. 397:164 [1999]). Potential cell cycle arrest induced by siRNA or dominant negative inhibitors of is determined by conventional flow cytometric methods. By using stable cell lines that "activate" cancer markers and mutants in a 4-hydroxytamoxifen-dependent fashion, cell proliferation and cell cycle alterations are monitored in a highly controlled *in vitro* system. To confirm that overexpression or inhibition of cancer markers does not activate the apoptosis pathway, several assays are used including propidium iodide staining of nuclei, TUNEL assay and caspase activation.

If a cancer marker is found to be a regulator of cell proliferation in prostate cells, studies are designed to address how components of cell cycle machinery are modulated by the cancer marker. Thus, in order to study cancer marker mediated effects on the cell cycle machinery of prostate cells, cancer marker functions are modulated with the above

mentioned tools (i.e., siRNA, dominant negative inhibition, etc.) and the expression levels (transcript and protein) of cyclins (cyclin D1,E,A), cyclins-dependent kinases (cdk2, cdk4, cdk6) and cyclin-dependent kinase inhibitors (p21CIP1, p27KIP1, p45SKP2, p16INK4) are monitored.

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H. Cell Adhesion and Invasion Assays

If a cancer marker is suspected of altering cell adhesion (e.g., the transcriptional repression of an epithelial gene program such as E-cadherin), the methods described above are used to investigate whether over-expression of the cancer marker causes increased or decreased cell adhesion. Adhesion to extracellular matrix components, human bone marrow endothelium (HBME) as well as to human umbilical vein endothelial cells (HUVEC) is tested. Cancer markers are further tested for their ability to modulate invasion of PCA.

Known methods are used in these studies (Cooper et al., Clin. Cancer Res. 6:4839 [2000]). Briefly, snap-apart 96-well tissue culture plates are coated with crude bone and kidney matrices. Plates are incubated overnight at room temperature under sterile conditions and stored at 4°C until needed. Assay plates are also coated with extracellular matrix components (e.g., human collagen I, human fibronectin, mouse laminin I) and human transferrin at various concentrations according to the manufacturer's instruction (Collaborative Biomedical Products, Bedford, MA). Endothelial cells (HBME or HUVEC) are seeded onto bone matrices or plastic substrata at a concentration of 900 cells/µl and grown to confluence. Tumor cells are removed from the flask by a 15-20 minute treatment with 0.5mM EDTA in Hank's balanced salt solution. Once the EDTA solution is removed, the cells are resuspended in adhesion medium (e.g., minimum essential medium (MEM) with 1% bovine serum albumin (BSA) supplemented with 10 uCi 51 Cr sodium salt (NEN, Boston, MA)) for 1 hour at 37°C. Cells are then washed three times in isotope free media and 1 x 10⁵ radio-labeled tumor cells are resuspended in adhesion media and layered upon a confluent layer of endothelial cells for 30 min at 37°C. In addition, radiolabeled tumor cells are applied to crude bone matrices. Again, plates are washed three times in phosphate buffered saline and adhesion is determined by

counting individual wells on a gamma counter. Cell adhesion is reported relative to the adhesion of controls (PC-3 cells on plastic), which are set to 100.

Cell invasion assays are performed using a classic Boyden chamber assay. Both strategies to inhibit and overexpress cancer markers are evaluated. Previous reports have correlated increased cell migration in a Boyden Chamber system with increased invasive properties in vivo (Klemke et al., J Cell Biol. 140:61 [1998]. Commercially available 24-well invasion chambers are used (e.g., BD biosciences, Chemicon International).

I. Transcriptional Suppression in Prostate Cancer Cells

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In some embodiments, the effect of cancer markers on gene silencing in prostate cells is assessed. Gene silencing is assayed in several ways. First, gene expression alterations induced by transient transfection of cancer markers and mutants in prostate cell lines (RWPE, DU145, LnCAP, and PC3) is assayed using FUGENE. Twelve to 48 hours after transfection, cells are harvested and a portion is processed to confirm expression of the transfectants by immunoblotting. Using vector-transfected cells as a reference sample, total RNA from transfected cells is then assessed on 20K cDNA microarrays.

In addition to transient transfections, stable cell lines overexpressing cancer markers and cancer marker mutants are generated. Patterns of gene expression from cancer marker and cancer marker mutant expressing cell lines are compared to vector-matched controls in order to identify a gene or group genes that is repressed by a given cancer marker. The present invention is not limited to a particular mechanism. Indeed, and understanding of the mechanism is not necessary to practice the present invention. Nonetheless, it is contemplated that genes identified as repressed by a given cancer marker will be increased (de-repressed) upon knock-down of the cancer marker (e.g., by siRNA inhibition).

All publications and patents mentioned in the above specification are herein incorporated by reference. Various modifications and variations of the described method and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in

connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in the relevant fields are intended to be within the scope of the following claims.

CLAIMS

We claim:

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- 1. A method for characterizing prostate tissue in a subject, comprising:
 - a) providing a prostate tissue sample from a subject; and
- b) detecting the presence or absence of expression of hepsin in said sample, thereby characterizing said prostate tissue sample.
- 2. The method of Claim 1, wherein said detecting the presence of expression of hepsin comprises detecting the presence of hepsin mRNA.
- 3. The method of Claim 2, wherein said detecting the presence of expression of hepsin mRNA comprises exposing said hepsin mRNA to a nucleic acid probe complementary to said hepsin mRNA.
- The method of Claim 1, wherein said detecting the presence of expression of hepsin comprises detecting the presence of a hepsin polypeptide.
- The method of Claim 4, wherein said detecting the presence of a hepsin polypeptide comprises exposing said hepsin polypeptide to an antibody specific to said
 hepsin polypeptide and detecting the binding of said antibody to said hepsin polypeptide.
 - 6. The method of Claim 1, wherein said subject comprises a human subject.
 - 7. The method of Claim 1, wherein said sample comprises tumor tissue.
 - 8. The method of Claim 7, wherein said tumor tissue sample is a post-surgical tumor tissue sample and said method further comprises the step of c) identifying a risk of prostate specific antigen failure based on said detecting the presence or absence of expression of hepsin.

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9. The method of Claim 1, wherein said characterizing said prostate tissue comprises identifying a stage of prostate cancer in said prostate tissue.

- 10. The method of Claim 9, wherein said stage is selected from the group consisting of high-grade prostatic intraepithelial neoplasia, benign prostatic hyperplasia, prostate carcinoma, and metastatic prostate carcinoma.
 - 11. The method of Claim 1, further comprising the step of c) providing a prognosis to said subject.

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- 12. The method of Claim 11, wherein said prognosis comprises a risk of developing prostate specific antigen failure.
- 13. The method of Claim 12, wherein said prognosis comprises a risk of developing prostate cancer.
 - 14. A method for characterizing prostate tissue in a subject, comprising:
 - a) providing a prostate tissue sample from a subject; and
 - b) detecting the presence or absence of expression of pim-1 in said sample, thereby characterizing said prostate tissue sample.
 - 15. The method of Claim 14, wherein said detecting the presence of expression of pim-1 comprises detecting the presence of pim-1 mRNA.
- 25 16. The method of Claim 15, wherein said detecting the presence of expression of pim-1 mRNA comprises exposing said pim-1 mRNA to a nucleic acid probe complementary to said pim-1 mRNA.
- 17. The method of Claim 14, wherein said detecting the presence of expression of pim-1 comprises detecting the presence of a pim-1 polypeptide.

18. The method of Claim 17, wherein said detecting the presence of a pim-1 polypeptide comprises exposing said pim-1 polypeptide to an antibody specific to said pim-1 polypeptide and detecting the binding of said antibody to said pim-1 polypeptide.

- 5 19. The method of Claim 14, wherein said subject is a human subject.
 - 20. The method of Claim 14, wherein said sample comprises tumor tissue.
- 21. The method of Claim 20, wherein said tumor tissue is post-surgical tumor tissue and said method further comprises the step of c) identifying a risk of prostate specific antigen failure based on said detecting the presence or absence of expression of hepsin.
 - 22. The method of Claim 14, wherein said characterizing said prostate tissue comprises detecting a stage of prostate cancer in said prostate tissue.
 - 23. The method of Claim 22, wherein said stage is selected from the group consisting of high-grade prostatic intraepithelial neoplasia, benign prostatic hyperplasia, prostate carcinoma, and metastatic prostate carcinoma.

- 24. The method of Claim 14, further comprising the step of c) providing a prognosis to said subject.
- 25. The method of Claim 24, wherein said prognosis comprises a risk ofdeveloping prostate specific antigen failure.
 - 26. The method of Claim 24, wherein said prognosis comprises a risk of developing prostate cancer.
- 30 27. A method for characterizing prostate tissue in a subject, comprising:
 - a) providing a prostate tissue sample from a subject; and

b) detecting the presence or absence of expression of EZH2 in said sample, thereby characterizing said prostate tissue sample.

- 28. The method of Claim 27, wherein said detecting the presence of expression of EZH2 comprises detecting the presence of EZH2 mRNA.
 - 29. The method of Claim 28, wherein said detecting the presence of expression of hepsin mRNA comprises exposing said hepsin mRNA to a nucleic acid probe complementary to said hepsin mRNA.

- 30. The method of Claim 27, wherein said detecting the presence of expression of EZH2 comprises detecting the presence of a EZH2 polypeptide.
- 31. The method of Claim 30, wherein said detecting the presence of a EZH2 polypeptide comprises exposing said EZH2 polypeptide to an antibody specific to said EZH2 polypeptide and detecting the binding of said antibody to said EZH2 polypeptide.
 - 32. The method of Claim 27, wherein said subject comprises a human subject.
- 20 33. The method of Claim 27, wherein said sample comprises tumor tissue.
 - 34. The method of Claim 27, wherein said characterizing said prostate tissue comprises identifying a stage of prostate cancer in said prostate tissue.
- 25 35. The method of Claim 34, wherein said stage is selected from the group consisting of high-grade prostatic intraepithelial neoplasia, benign prostatic hyperplasia, prostate carcinoma, and metastatic prostate carcinoma.
- 36. The method of Claim 27, further comprising the step of c) providing a prognosis to said subject.

37. The method of Claim 36, wherein said prognosis comprises a risk of developing metastatic prostate cancer.

- 38. A kit for characterizing prostate cancer in a subject, comprising:
- a) a reagent capable of specifically detecting the presence of absence of expression of hepsin; and
 - d) instructions for using said kit for characterizing cancer in said subject.
- The kit of Claim 38, wherein said reagent comprises a nucleic acid probe complementary to a hepsin mRNA.
 - 40. The kit of Claim 38, wherein said reagent comprises an antibody that specifically binds to a hepsin polypeptide.
 - 41. The kit of Claim 38, wherein said instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.
- 20 42. A kit for characterizing prostate cancer in a subject, comprising:

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- a) a reagent capable of specifically detecting the presence of absence of expression of pim-1; and
- d) instructions for using said kit for characterizing cancer in said subject.
- 43. The kit of Claim 42, wherein said reagent comprises a nucleic acid probe complementary to a pim-1 mRNA.
- 44. The kit of Claim 42, wherein said reagent comprises an antibody that specifically binds to a pim-1 polypeptide.

45. The kit of Claim 42, wherein said instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

- 5 46. A kit for characterizing prostate cancer in a subject, comprising:
 - a) a reagent capable of specifically detecting the presence of absence of expression of EZH2; and
 - d) instructions for using said kit for characterizing cancer in said subject.

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- 47. The kit of Claim 46, wherein said reagent comprises a nucleic acid probe complementary to a EZH2 mRNA.
- 48. The kit of Claim 46, wherein said reagent comprises an antibody that specifically binds to a EZH2 polypeptide.
 - 49. The kit of Claim 46, wherein said instructions comprise instructions required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.

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- 50. A method of screening compounds, comprising:
 - a) providing
 - i) a prostate cell sample; and
 - ii) one or more test compounds; and

- b) contacting said prostate cell sample with said test compound; and
- c) detecting a change in hepsin expression in said prostate cell sample in the presence of said test compound relative to the absence of said test compound.
- 30 51. The method of Claim 50, wherein said detecting comprises detecting hepsin mRNA.

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WO 03	3/012067	PC1/0502/24307
	52.	The method of Claim 50, wherein said detecting comprises detecting
hepsin	polype	
	53.	The method of Claim 50, wherein said cell is in vitro.
	54.	The method of Claim 50, wherein said cell is in vivo.
	55.	The method of Claim 50, wherein said test compound comprises an
antiser	ise com	ound.
	56.	The method of Claim 50, wherein said test compound comprises a drug.
	57.	A method of screening compounds, comprising:
		a) providing
		i) a prostate cell sample; and
		ii) one or more test compounds; and
		contacting said prostate cell sample with said test compound; and
		detecting a change in pim-1 expression in said prostate cell sample
	in the	resence of said test compound relative to the absence of said test
	compo	
	Compe	
٠.	58.	The method of Claim 57, wherein said detecting comprises detecting pim-
1 mR1	VA.	
•		
	59.	The method of Claim 57, wherein said detecting comprises detecting pim-
1 poly	peptide	

The method of Claim 57, wherein said cell is in vitro.

The method of Claim 57, wherein said cell is in vivo.

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20 EZH2 polypeptide.

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	62.	The m	nethod o	of Claim 57, wherein said test compound comprises an	:
antiser	ise com	pound.		. · · · · · · · · · · · · · · · · · · ·	
	63.	The m	nethod o	of Claim 57, wherein said test compound comprises a dru	g.
	64.	A met	thod of s	screening compounds, comprising:	
		a)	provid	ding	
			i)	a prostate cell sample; and	
			ii)	one or more test compounds; and	
		b)	contac	cting said prostate cell sample with said test compound; a	nd
		c)	detect	ing a change in EZH2 expression in said prostate cell san	npl
	in the	presenc	ce of sai	id test compound relative to the absence of said test	
	compo	=	·		
	65.	The m	nethod o	of Claim 64, wherein said detecting comprises detecting	
EZH2	mRNA				
		-			
, .	66	The m	nethod o	of Claim 64, wherein said detecting comprises detecting	

- 67. The method of Claim 64, wherein said cell is in vitro.
 - 68. The method of Claim 64, wherein said cell is in vivo.
- 69. The method of Claim 64, wherein said test compound comprises an antisense compound.
 - 70. The method of Claim 64, wherein said test compound comprises a drug.

71. A method for characterizing inconclusive prostate biopsy tissue in a subject, comprising:

- a) providing an inconclusive prostate biopsy tissue sample from a subject; and
- b) detecting the presence of expression of AMACR in said sample,
 thereby characterizing said inconclusive prostate biopsy tissue sample.
- 72. The method of Claim 71, wherein said detecting the presence of expression of AMACR comprises detecting the presence of AMACR mRNA.

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- 73. The method of Claim 72, wherein said detecting the presence of expression of AMACR mRNA comprises exposing said AMACR mRNA to a nucleic acid probe complementary to at least a portion of said AMACR mRNA.
- The method of Claim 71, wherein said detecting the presence of expression of AMACR comprises detecting the presence of a AMACR polypeptide.
- 75. The method of Claim 74, wherein said detecting the presence of a
 AMACR polypeptide comprises exposing said AMACR polypeptide to an antibody
 specific to said AMACR polypeptide and detecting the binding of said antibody to said
 AMACR polypeptide.
 - 76. The method of Claim 71, wherein said subject comprises a human subject.
- The method of Claim 71, wherein the presence of AMACR expression in said inconclusive biopsy tissue is indicative of prostate cancer in said subject.
 - 78. The method of Claim 71, wherein said method further comprises the step of detecting expression of a basal cell marker selected from the group consisting of 34βE12 and p63 and the absence of a basal cell marker expression and the presence of AMACR expression is indicative of prostate cancer in said subject.

- 79. A method of detecting AMACR expression in a bodily fluid, comprising:
 - a) providing
 - i) a bodily fluid from a subject; and
 - ii) a reagent for detecting AMACR expression in said biological fluid; and
- b) contacting said bodily fluid with said reagent under conditions such that said reagent detects AMACR expression in said bodily fluid.
- 10 80. The method of claim 79, wherein said bodily fluid is selected from the group consisting of serum, urine, whole blood, lymph fluid, and mucus.
 - 81. The method of claim 79, wherein the presence of AMACR in said bodily fluid is indicative of cancer.
 - 82. The method of claim 79, wherein said cancer is prostate cancer.
 - 83. A kit for characterizing inconclusive prostate biopsy tissue in a subject, comprising:
- a) a reagent capable of specifically detecting the presence or absence of expression of AMACR; and
 - b) instructions for using said kit for characterizing inconclusive biopsy tissue in said subject.
- 25 84. The kit of Claim 83, wherein said reagent comprises a nucleic acid probe complementary to at least a portion of an AMACR mRNA.
 - 85. The kit of Claim 83, wherein said reagent comprises an antibody that specifically binds to a AMACR polypeptide.

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86. The kit of Claim 83, wherein said kit further comprises a second reagent, said second reagent capable of specifically detecting the expression of a basal cell marker selected from the group consisting of $34\beta E12$ and p63.

87. The kit of claim 86, wherein said instructions further comprise instructions for using said second reagent and said reagent for characterizing inconclusive biopsy tissue in said subject.

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- 88. The kit of Claim 83, wherein said instructions comprise instructions
 10 required by the United States Food and Drug Administration for use in *in vitro* diagnostic products.
 - 89. A method of characterizing tissue in a subject, comprising:
 - a) providing a tissue sample from a subject, said tissue sample selected from the group consisting of breast tissue, ovarian tissue, lymph tissue, and melanoma tissue; and
 - b) detecting the presence or absence of expression of AMACR in said sample, thereby characterizing said breast tissue sample.
- 20 90. A method of diagnosing cancer in a subject, comprising:
 - a) providing a tissue sample from a subject, said tissue sample selected from the group consisting of breast tissue, ovarian tissue, lymph tissue, and melanoma tissue; and wherein said subject is suspected of having breast cancer; and
- b) detecting the presence of expression of AMACR in said sample,
 25 thereby diagnosing cancer in said subject.
 - 91. A method of diagnosing cancer in a subject, comprising:
 - a) providing a blood sample from a subject suspected of having cancer; and
- 30 b) detecting an immune response to AMACR in said blood sample, thereby diagnosing cancer in said subject.

- 92. The method of claim 91, wherein said cancer is prostate cancer.
- 93. The method of claim 91, wherein said detecting an immune response comprises detecting an antibody against said AMACR in said blood sample.
 - 94. A method of inhibiting the growth of cells, comprising
 - a) providing
 - i) a cell that expresses EZH2; and
- i) a reagent for inhibiting EZH2 expression in said cell; and
 - b) contacting said cell with said reagent under conditions such that the expression of EZH2 in said cell is inhibited.
- 95. The method of claim 94, wherein said reagent is an antisense oligonucleotide.
 - 96. The method of claim 94, wherein said reagent is a RNA duplex.
 - 97. The method of claim 94, wherein said reagent is a drug.
 - 98. The method of claim 94, wherein said cell is a prostate cancer cell.
 - 99. The method of claim 94, wherein said cell is in vitro.
- 25 100. The method of claim 94, wherein said cell is in vivo.
 - 101. The method of claim 94, wherein said contacting further results in a decrease in proliferation of said cell.

30

Figure 1

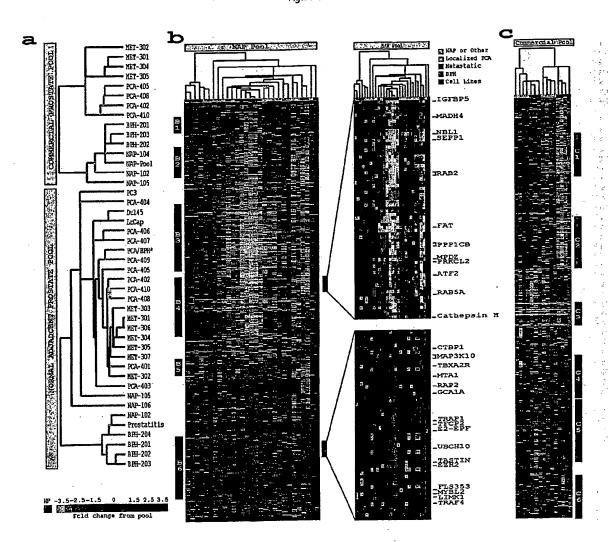


Figure 2

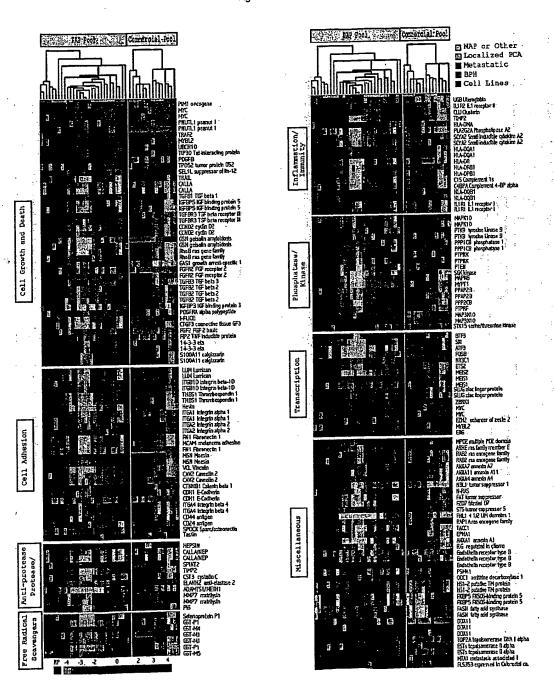
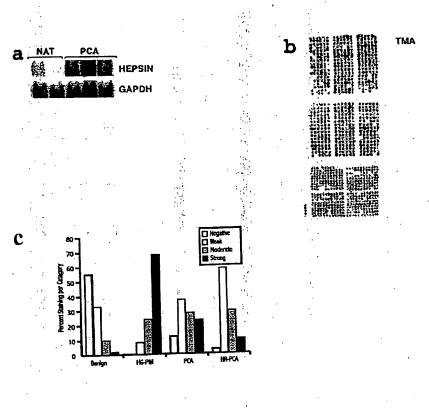
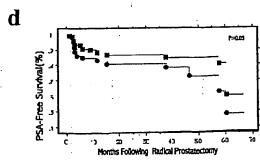


Figure 3

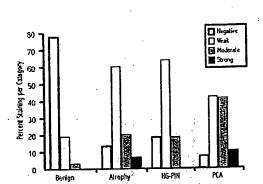




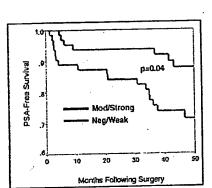
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Figure 4

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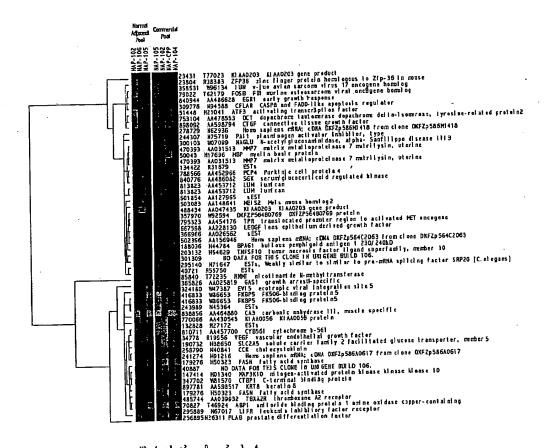


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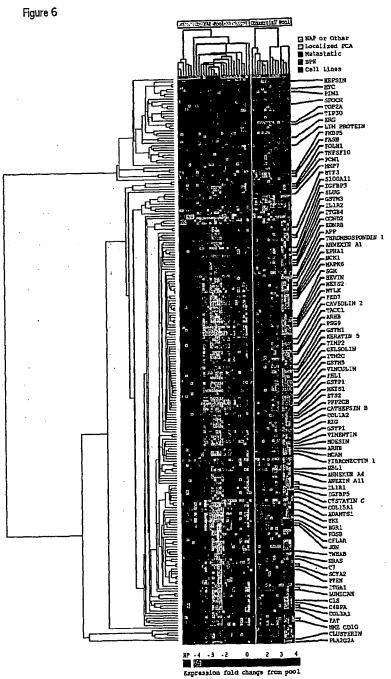


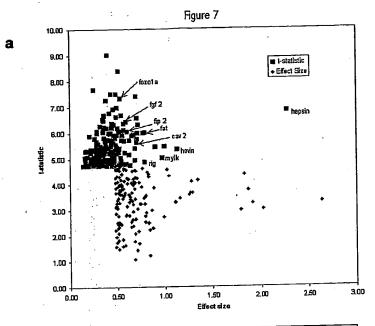
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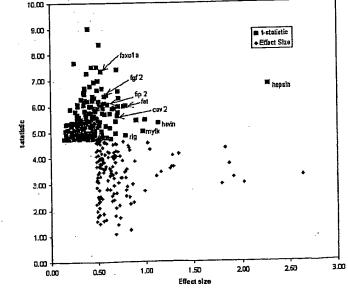
Figure 5



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Figure 8

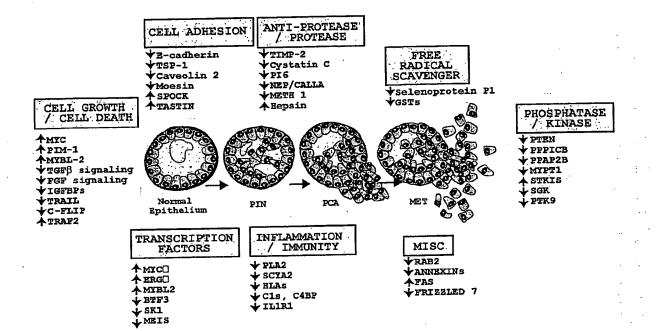


Figure 9

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9	IGFBP3	XM_004689
10	SLUG	XM_011634
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12	IL1R2	X59770
13	ITGB4	X53587
14	CCND2	XM_034568
15	EDNRB	S57283
16	APP	X06989
17	THROMBOSPONDIN 1	X04665
18	ANNEXIN A1	XM_005665
19	EPHA1	M18391
20	NCK1	XM_051968
21	MAPK6	XM_017662
22	SGK	XM_037045
23	HEVIN	XM_011533
24	MEIS2	XM_012430
25	MYLK	XM_042191
26	FZD7	NM_003507
27	CAVEOLIN 2	XM_004966
28	TACC1	XM_049505
29	ARHB	XM_002689
30	PSG9	NM_002784
31 .	GSTM1	NM_000561
32	Keratin 5	XM_006847
33	TIMP2	XM_027036
34	GELSOLIN	XM_016545
35	ITM2C	AA034213
36	GSTM5	XM 002154
37	VINCULIN	XM 011883
38	FHL1	XM 042931

Figure 9 (cont.)

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44	COL1A2	XM_029246
45	RIG	XM_006029
46	VIMENTIN	XM_042952
47	MOESIN	XM_013042
48	MCAM	XM_006077
49	FIBRONECTIN 1	XM_030549
50	NBL1	XM_001434
51	ANNEXIN A4	XM_031594
52	ANNEXIN A11	XM_035906
53	IL1R1	XM_002686
54	IGFBP5	XM 046731
55	CYSTATIN C	XM 009599
56	COL15A1	XM 005592
57		XM 047796
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59	EGR1	XM 033546
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64	NRAS	XM 001317
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66	SCYA2	XM 038982
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73	MMECD10	XM 030168
74	CLUSTERIN	XM 005113
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Figure 9 (cont.)

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90	TRAP1	XM_036666
91	TFCP2	XM_051171
92	E2-EPF	XM_012615
93	UBCH10	XM_009488
94	TASTIN	XM_006826
95	EZH2	XM_004774; NM004456
96	FLS353	AB024704
97	MYBL2	XM_009492
98	LIMK1	XM_051836
99	TRAF4	XM_031428
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115	CTBP2	AF016507
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122	VaV2	XM_005638

Figure 10

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901

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Figure 10 (cont.)

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1381

14/129

Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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     361 cettecagee tttetgteat catetecaca geccaeceat eccetgagea caetaaceae
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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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1861 ctgtacataa caactttgat attgatgaag ttcagcttga tccaagagct ctgtccgatg
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2161 atgaacccaa gcttattata gatctttcca attggaaaga acaaagcaaa gaaaaatctg
2221 ataagaaagg caaatcaaaa tgtgaaagga atggattggt taaagcccag atagcgctag
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2341 attectttat tgcaggaact atteagetta gtteccagea tgageetaet gatgttgttg
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2641 acttggacaa gttctttagc aggaaagaag atactgaaat gctagaaact gagccagtag
2701 aggatgggaa gcttggggag agaggacatg aggaaggatt tctgaacaac agtggggagt
2761 toototttaa caagcagoto gagtocatag goatoccaca gtttcacagt ccagttgggt
2821 caccacttaa gtcaatacag gccacattaa caccttctgc tatgaaatct tcccctcaaa
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2941 ttatctttgt attcttcatg aaatgtgttt tgtctttttt tattactagt gtttaagtca
3001 ttttttactt gaatcagatg gtgtcattta gtaaggattt tatgagttct tgtttttaa
3061 aatccagact ttcttttct acatgtgaga tagttttcat tttaactggc atgtcatttg
3121 cacacaaaaa taaagactag agcaaaataa tgcaacgcag gaggagaaaa gaaatgcact
3181 aagacaagaa cattototoa tagaacattg atotgtttta caggaaacaa acottgoott
3241 gaaatttaca cagtgag
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¹ ggtctttgag cgctaacgtc tttctgtctc cccgcggtgg tgatgacggt gaaaactgag

Figure 10 (cont.)

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241 gagettatga atgecaacce ttetecteca ecaagteett eteageaaat caacettgge
301 ccgtcgtcca atcctcatgc taaaccatct gactttcact tcttgaaagt gatcggaaag
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901 ttgtatgaga tgctgtatgg cctgccgcct ttttatagcc gaaacacagc tgaaatgtac
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1381 cgaatgtttt agttagcctt ttggtggagc cgccagctga caggacatct tacaagagaa
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1681 attatgaaat gtgccttttc tgaagagatt gtgttagctc caaagctttt cctatcgcag
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2161 agaacgtctg tacattgggt tataacacta gtatatttaa acttacaggc ttatttgtaa
2221 tgtaaaccac cattttaatg tactgtaatt aacatggtta taatacgtac aatcettccc
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SEQ ID NO:23

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121 tgtgtgcaga ggggattcaa cttcaatttt tctgcagtgg ctctgggtcc agccccttac
181 ttaaagatct ggaaagcatg aagactgggc tttttttcct atgctctttg
241 ctgcaatccc gacaaatgca agattattat ctgatcattc caaaccaact gctgaaacgg
301 tagcacctga caacactgca atccccagtt taagggctga agctgaagaa aatgaaaaag
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481 tgggattgaa ggatcaagag gacagtgatg gtcacttaag tgtgaatttg gagtatgcac
541 caactgaagg tacattggac ataaaagaag atatgagtga gcctcaggag aaaaaacctc
501 cagagaacac tgatttttg gctcctggtg ttagttcctt cacagattct aaccaacaag
661 aaagtatcac aaagaggag gaaaaccaag aacaacctag aaattattca catcatcagt
721 tgaacaggag cagtaaacat agccaaggcc taagggatca aggaaaccaa gagcaggatc
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Figure 10 (cont.)

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 901 aagacaatac ccaatctgat gatattttgg aagagtctga tcaaccaact caagtaagca
 961 agatgcagga ggatgaattt gatcagggta accaagaaca agaagataac tccaatgcag
1021 aaatggaaga ggaaaatgca tcgaacgtca ataagcacat tcaagaaact gaatggcaga
1081 gtcaagaggg taaaactggc ctagaagcta tcagcaacca caaagagaca gaagaaaaga
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1201 atggagttga tgatgatggc gatgatgatg gcgatgatgg cggcactgat ggccccaggc
1261 acagtgcaag tgatgactac ttcatcccaa gccaggcctt tctggaggcc gagagagctc
1321 aatccattgc ctatcacctc aaaattgagg agcaaagaga aaaagtacat gaaaatgaaa
1381 atataggtac cactgagect ggagageace aagaggeeaa gaaageagag aacteateaa
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1501 gcttccagtg taaaagaggc cacatctgta aggcagacca acagggaaaa cctcactgtg
1561 tetgecagga tecagtgaet tgteetecaa caaaaceeet tgateaagtt tgtggeaetg
1621 acaatcagac ctatgctagt tcctgtcatc tattcgctac taaatgcaga ctggagggga
1681 ccaaaaaggg gcatcaactc cagctggatt attttggagc ctgcaaat
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SEQ ID NO:24

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- 1 agcacactga ggaggcgatc cgccagcagg aggtggagca gctggacttc cgagacctcc
 - 61 tggggaagaa ggtgagtaca aagaccctat cggaagacga cctgaaggag atcccagccg
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Figure 10 (cont.)

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361 agaccetgaa tgccaaggca gtggagagtt ccaagcecet gagcaatgca cagcettcag
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Figure 10 (cont.)

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     3601 aactatctca tetgteagat ttttaaaact ceaacacagg ttttggeate ttttgtgetg
     3661 tatottttaa gigcaigiga aattigiaaa atagagataa giacagiaig tatattiigi
     3721 aaatotooca tittigtaag aaaatatata tigtatitat acattittac titiggatiit
     3781 tgttttgttg getttaaagg tetaccecae tttatcacat gtacagatca caaataaatt
     3841 tttttaaata c
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      601 cattgeteca tügtgtacga gegtaggacg atgettetet tetgteagee tgeaactgag
     661 ccaggattga atacttggac cccaggtctg gagattggga tactgtaata cttctttgtt
     721 attataacat aaaagcacca ctgttctgtt catttcctag ctgttctaat taagaaaact
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     1021 aacttcccat taaaaatgag atttcttatt tgtttgtctg tttttactct gggagtaata
     1081 ctttttaaat tacctttaca tatatagtca ctggcatact gagaatatac aatgatectg
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1 gtttagaaca gcctacagac ccagtggcac gagacgggcc tctctcccaa acatcttcca
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121 tgcagaacte eccacecete tettetgagg getectacea etttgaceca gataactttg
181 acgaatccat ggatcccttt aaaccaacta cgaccttaac aagcagtgac ttttgttctc
241 ccactggtaa tcacgttaat gaaatcttag aatcacccaa gaaggcaaag tcgcgtttaa
301 taacgactac tgaacaagtg aaatttetet gttttetgtt gagtggetgt aaggtgaaga
361 agcatgaaac tcagtetete gecetggatg catgtteteg ggatgaaggg geagtgatet
421 cccagatttc agacatttct aatagggatg gccatgctac tgatgaggag aaactggcat
481 ccacgtcatg tggtcagaaa tcagctggtg ccgaggtgaa aggtgagcca gaggaagacc
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781 gagaagagat aattactaaa gagattgaag caaatgaatg gaagaagaaa tacgaagaga
841 cccggcaaga agttttggag atgaggaaaa ttgtagctga atatgaaaag actattgctc
901 aaatgattga tgaacaaagg acaagtatga cototoagaa gagottocag caactgacca
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1321 aaatetgtga tgagetgatt gcaaagetgg gaaagaetga etgagaeact eeeeetgtta
1381 gctcaacaga totgcatttg gctgcttctc ttgtgaccac aattatcttg ccttatccag
1441 gaataattgc ccctttgcag agaaaaaaaa aaacttaaaa aaagcacatg cctactgctg
1501 cctgtcccgc tttgctgcca atgcaacagc cctggaagaa accctagagg gttgcatagt
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1981 attttgaaat agagteetga eteagaacae caaettaaga atttggggga ttaaagatgt
2041 gaagaccaca gtcttgggtt ttcatatctg gagaagacta tttgccatga cgttttgttg
2101 ccctggtatt tggacactcc tcagctttaa tgggtgtggc ccctttaggg ttagtcctca
2161 gactaatgat agtgtctgct ttctgcatga acggcaatat gggactccct ccaagctagg
2221 gtttggcaag tctgccctag agtcatttac tctcctctgc ctccatttgt taatacagaa
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2401 cacattacca attataaggt gaagaaatgt ttttttccca agtgtgatgc attgttcttc
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2581 agacagettg ttgaatactg agaagaggag tgcaaggaga aggtetgtae taacaaagee
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2701 aatttttcca tataaatctc aatgaattcc ctttcatttg aataggcaaa cccaaatcca
2761 tgcaagtgtt ttaaagcact gtcctgtctt aatcttacat gctgaaagtc ttcatggtga
2821 tatgcactat attcagtata cgtatgtttt cctacttctc ttgtaaaact gttgcatgat
2881 ccaacttcag caatgaattg tgcctagtgg agaacctcta tagatcttaa aaaatgaatt
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3061 ctgtatattc aagcctgttg tcttaacatt ttgtataaaa aagaacaaca gaaattatct
3121 gtcatttgag aagtggcttg acaatcattt gagctttgaa agcagtcact gtggtgtaat
3181 atgaatgotg tootagtggt catagtacca agggcacgtg totoccottg gtataactga
3241 tttccttttt agtcctctac tgctaaataa gttaattttg cattttgcag aaagaaacat
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3481 cctagcacac tggtggaaga gaccccttaa gaacctgacc ccagtgaatg aagctgatgc
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 3661 aatcgtagct tgtacagatc aagagaatat actgggcaga atgaagtatg tttgtttatt
 3721 tttctttaaa aataaaggat tttggaactc tggagagtaa gaatatagta tagagtttgc
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Figure 10 (cont.)

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      421 cgaggtette gagaeggeea egegegeege getgeagaag egetaegget eecagaaegg
      481 ctgcatcaac tgctgcaagg tgctatgagg gccgcgcccg tcgcgcctgc ccctgccggc
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      301 aagagaaaca gtatattcca acgcatccct gctgatccag aatgtcaccc ggaaggatgc
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      781 gaaattcaag acaaagaaga aaaaaactca atgttattgg actaaataat caaaaggata
      841 atgttttcat aattttttat tggaaaatgt gctgattctt tgaatgtttt attctccaga
      901 tttatgaact ttttttcttc agcaattggt aaagtatact tttgtaaaca aaaattgaaa
      961 tatttgcttt tgctgtctat ctgaatgccc cagaattgtg aaactactca tgagtactca
     1021 taggtttatg gtaataaagt tatttgcaca tgttccgtag ttt
SEQ ID NO:31
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Figure 10 (cont.)

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2341 ggctgagctg gctgcctgag gaggggcagg gcccacccat gtcaccggtc agtgcctttt
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2461 tgtgtgtgtg tgtgttgttt ctttttttt tttttacagt atccaaaaat agccctgcaa
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2581 aaaacatttt gaagtgtg
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SEQ ID NO:35

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61 geeeatgact etggggtaet gggacateeg tgggetggee caegeatee gettgeteet
121 ggaatacaca gacteaaget atgtggaaaa gaagtacaeg etgggggaeg eteetgaeta
181 tgacagaage eagtggetga atgaaaaatt eaagetggge etggaettte eeaatetgee
241 etaettgatt gatggggete acaagateae eeagagcaat geeateetge getacattge
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Figure 10 (cont.)

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 841 aggcctcatt ggcttccttt cttctaacat catccctcc cgcatcgagg ctctttaaag
 901 cttcagctcc ccactgtcct ccatcaaagt ccccctccta acgtcttcct ttccctgcac
 961 taacgccaac etgactgett tteetgteag tgettttete ttetttgaga agccagactg
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1081 ggtccctacc ccagctccgt gtgatgccca gtaaagcctg aaccatgcct gccatgtctt
1141 gtcttattcc ctgaggctcc cttgactcag gactgtgctc gaattgtggg tggttttttg
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1261 ggtgatcagg caggttcata aatttecttg gtcatttctg ccctctagcc acatecetet
1321 gttcctcact gtggggatta ctacagaaag gtgctctgtg ccaagttcct cactcattcg
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  601 caaaggttgg ctccgtgacc ctagtgcctc cccaggggat gctggtgagc aggccatcag
  661 acagatetta gatgaagetg gaaaagttgg tgaactetgt geaggeaaag aacgeaggga
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1801 tatcttactt aggaaccctg gaaatcaagc tgcttatgaa cattttgaga ccatgaagaa
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Figure 10 (cont.)

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2461 ggctgccaga cagctccatg atgaagctcg caaatggtcc agcaagggca atgacatcat
2521 tgcagcagcc aagcgcatgg ctctgctgat ggctgagatg tctcggctgg taagaggggg
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2761 agtgaaggec accatgetgg geeggaecaa eateagtgat gaggagtetg ageaggeeae
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2941 aaagactccc tggtaccagt aggcacctgg ctgagcctgg ctggcacaga aacctctact
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3601 tataaaactg tacttcactg tcaggaagaa atcacagaat catatgattc tgcttttacc
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4081 cctaagagtg tttaatggca aggcagccct gtctgaagga cacttcctgc ctaagggaga
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4261 tatggggttc aagagagtaa tgggtttcat atttcttatc accacagtaa gttcctacta
4321 ggcaaaatga gagggcagtg tttccttttt ggtacttatt actgctaagt atttcccagc
4381 acatgaaacc ttattttttc ccaaagccag aaccagatga gtaaaggagt aagaaccttg
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4681 agaggaaata agaaaaatca tgtttgctct cccggttctt ccagtggttt gagacactgg
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SEQ ID NO:38

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Figure 10 (cont.)

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    1381 tetgeatgtt teteatagag cagaaaagtg etaateattt agceaettag tgatgtaage
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    1741 tgtggcatgt tttctgagcg ttcctacttt aaagcatgga acatgcaggt gatttgggaa
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    1861 ctggaagett aacaaaacta accetgetgt cetttttatt gtttttaatt aatatttttg
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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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661

56/129

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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     661 gctcagcagg ccatggacat ccacttccat agcceggcct tccaqcacce qccaacaqaa
     721 ttcatacgag aaggegaega tgaccggact gtgtgccggg agatccgcca caactccacg
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     901 gctgagaggt tgaccaggaa atacaacgag ctgctaaagt cctaccagtg gaagatgctc
     961 aacaceteet eettgetgga geagetgaac gageagttta aetgggtgte eeggetggea
    1021 aacctcacgc aaggegaaga ccagtactat ctgcgggtca ccacggtggc ttcccacact
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    1321 cccagagaga gctctgcacg tcaccaagta accaggcccc agcctccagg cccccaactc
    1381 egeccageet eteccegete tggateetge actetaacae tegactetge tgeteatggg
    1441 aagaacagaa ttgctcctgc atgcaactaa ttcaataaaa ctgtcttgtg agctg
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Figure 10 (cont.)

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Figure 10 (cont.)

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1861 ttgcttagta agtatttcca tagtcaatga tggtttaata ggtaaaccaa accctataaa
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SEQ ID NO:78

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 181 caatgtagaa gaggcattta ttaatacagc aaaagaaatt tatgaaaaaa ttcaagaagg
 241 agtotttgac attaataatg aggcaaatgg cattaaaatt ggccctcagc atgctgctac
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 541 ggagattgta ttcatatcta tttgcatttg atttctaggt caattgatgt gattattttt
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 661 tottgagtat tttaaatogg tttgtgtagt taggtttooc aacatotgtg gttacctaat
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 781 gaaggactet tttaattetg tatttateat ttaetttetg tatatatagt ttaataacet
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SEO ID NO.79

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Figure 10 (cont.)

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      541 acagtaacct tggaagttgg aagaatcaaa gctggtccat tccattcaga gaggaggcca
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     1441 thatgorgtg theageoggg tetheaaaac tgtagggggg aaataacact taagtttett
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SEQ ID NO:83

80/129

Figure 10 (cont.)

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Figure 10 (cont.)

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     1261 gacgcaaaga tcaccagcca tgtgccttag tgtccttctt aacagactca aaccacatgg
     1321 accacgaata ttetttetgt ecagaaggge taetttecae atatagaget ecagggaetg
     1381 tetttetet attegetett caataaacat tgagtgagca cetecceaga tgg
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      121 ggcattgctg gatggccggg actgcacagt ggagatgccc atcctgaagg acgtggccac
      181 tgtggcette tgegaegege agteeaegea ggagateeat gagaaggtee tgaaegagge
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      481 gegggaggge acaegagtee agagegtega geagateege gaggtggegt eeggegetge
      541 caggateege ggggagaeet tgggeateat eggaettgte gegtggggea ggeagtggeg
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      661 gtggageggg egetgggget geagegtgte ageaceetge aggacetget ettecaeage
      721 gactgcgtga ccctgcactg cggcctcaac gagcacaacc accacctcat caacgactte
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      841 gatgagaagg cgctggccca ggccctgaag gagggccgga tccgcggcgc ggccctggat
      901 gtgcacgagt cggaaccett cagetttage cagggeeete tgaaggatge acceaacete
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Figure 10 (cont.)

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    1261 ctgcccctg tggcccaccc gccccacgcc ccttctcctg gccaaaccgt caagcccgag
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     1441 ggccctggca ctgcagagac tggtccgggc tgtcaggagg cgggaggggg cagcgctggg
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    1921 aagacggcat cacgaagcag ctccaaaagg aaaagcttgg gcggtgccca gcgtgcccgc
    1981 tgcccatcga cgtctgtcct ggggacgtgg agggtggcag cgtccccgcc tgcaccagtg
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      481 ageggeegeg tgggegtett eeccageaac taegtggeee eeggegeeee egetgeaeee
      541 gegggeetee agetgeecea ggagateeee tteeaegage tgeagetaga ggagateate
      601 ggtgtggggg gctttggcaa ggtctatcgg gccctgtggc gtggcgagga ggtggcagtc
      661 aaggccgccc ggctggaccc tgagaaggac ccggcagtga cagcggagca ggtgtgccag
      721 gaageeegge tetttggage eetgeageae eecaacataa ttgeeettag gggegeetge
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      901 cggggcatga actacctaca caatgatgcc cctgtgccca tcatccaccg ggacctcaag
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    1081 gggacctacg cctggatggc gccggaggtt atccgtctct ccctcttctc caaaagcagt
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    1261 ccctccacgt gccccgagcc ctttgcccgc ctcctggagg aatgctggga cccagacccc
    1321 cacgggegge cagatttegg tageatettg aageggettg aagteatega acagteagee
    1381 ctgttccaga tgccactgga gtccttccac tcgctgcagg aagactggaa gctggagatt
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    1621 ctgagccagg agaagccccg ggtccgcaag cgcaagggca acttcaagcg cagccgcctg
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     1741 acagtocagg cototocaac totggataag oggaaaggat ocgatggggo cagooccot
    1801 gcaagcccca gcatcatccc ccggctgagg gccattcgcc tgactcccgt ggactgtggt
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Figure 10 (cont.)

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SEQ ID NO:87

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 121 tgcctgctca gctccagacg gcgcccggac ccccgggcgc gggatccagc caggtgggag
 181 ccccgcagat gaggtetetg aaggtgtgcc tgaaccagtg ccagcctgcc ctgtctgcag
 241 categgeetg atggggtggt gaetgateee teagggetee ggageeatgt ggeeeaaegg
 301 cagttccctg gggccctgtt tccggcccac aaacattacc ctggaggaga gacggctgat
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 421 cetgagegtg etggegggeg egeggeaggg gggttegeac acgegeteet cetteeteac
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 661 catggcctca gagegetace tgggtateac eeggeeette tegegeeegg eggtegeete
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Figure 10 (cont.)

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Figure 10 (cont.)

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      361 ttggaaaaac tgcgtcacaa actggtgtct gacggccaag cactgccaga aatggagatt
      421 cacttgcaga ccaatgccga gaaaggcacc atcaccatcc aggatactgg tatcgggatg
      481 acacaggaag agctggtgtc caacctgggg acgattgcca gatcggggtc aaaggccttc
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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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    2041 cagoctcaco otgtoaggta toaaagaaga caacagottg otcaaccagg gottottgca
     2101 ggccaagccc gagaaggcag cagtggccca gaagccccga agccacttca cgacacctgc
    2161 ccctatgtcc agtgcctgga agacggtggc ctgcgggggg accagggacc agcttttcat
    2221 gcaggagaaa gcccggcagc tcctgggccg cctgaagccc agccacacat ctcggaccct
     2281 catcttgtcc tgaggtgttg agggtgtcac gagcccattc tcatgtttac aggggttgtg
     2341 ggggcagagg gggtctgtga atctgagagt cattcaggtg acctcctgca gggagccttc
     2401 tgccaccagc ccctccccag actctcaggt ggaggcaaca gggccatgtg ctgccctgtt
     2461 gccgagccca gctgtgggcg gctcctggtg ctaacaacaa agttccactt ccaggtctgc
     2521 ctggttccct ccccaaggcc acagggagct ccgtcagctt ctcccaagcc cacgtcaggc
     2581 ctggcctcat ctcagaccct gcttaggatg ggggatgtgg ccaggggtgc tcctgtgctc
     2641 accetetett ggtgeatttt tttggaagaa taaaattgee tetetett
SEQ ID NO:98
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       61 gagttgcccg tgtgtgcaag ctgcggccag aggatctatg atggccagta cctccaggcc
      121 ctgaacgegg actggcacge agactgette aggtgttgtg actgeagtge etceetgteg
      181 caccagtact atgagaagga tgggcagctc ttctgcaaga aggactactg ggcccgctat
      241 ggcgagtcct gccatgggtg ctctgagcaa atcaccaagg gactggttat ggtggctggg
      301 gagetgaagt accaececga gtgttteate tgeeteacgt gtgggaeett tateggtgae
      361 ggggacacct acacgetggt ggagcactec aagetgtact gegggeactg ctactaceag
      421 actgtggtga cccccgtcat cgagcagate ctgcctgact cccctggctc ccacctgccc
      481 cacaccetca ccctggtgtc catcccagcc tcatctcatg gcaagegtgg actttcagtc
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541 tecattgace eccegeacyg eccacegyge tytygycaceg agcaetcaca cacegteege 601 ytecaygyay tygateegyg etycatgage ecagatytya agaattecat ecaegtegya

Figure 10 (cont.)

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 781 acactgggcc acgggctggg gcctgagacc agccccctga gctctccggc ttatactccc
 841 agcggggagg cgggcagctc tgcccggcag aaacctgtct tcgcaaggac ctgggtcgct
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961 acggggaggt gctgggcaag ggctgcttcg gccaggctat caaggtgaca caccgtgaga
1021 caggtgaggt gatggtgatg aaggagctga tccggttcga cgaggagacc cagaggacgt
1081 tcctcaagga ggtgaaggtc atgcgatgcc tggaacaccc caacgtgctc aagttcatcg
1141 gggtgctcta caaggacaag aggctcaact tcatcactga gtacatcaag ggcggcacgc
1201 tooggggcat catcaagage atggacagee agtacecatg gagecagaga gtgagetttg
1261 ccaaggacat cgcatcaggg atggcctacc tccactccat gaacatcatc caccgagacc
1321 tcaactccca caactgcctg gtccgcgaga acaagaatgt ggtggtggct gacttcgggc
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1441 cagaccgcaa gaagcgctac accgtggtgg gcaaccccta ctggatggca cctgagatga
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1621 tcaacgtgcg aggattcctg gaccgctact gccccccaaa ctgccccccg agcttcttcc
1681 ccatcaccgt gcgctgttgc gatctggacc ccgagaagag gccatccttt gtgaagctgg
1741 aacactggct ggagaccete egeatgeace tggeeggeea cetgeeactg ggeecacage
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1861 cccaccctga ggtccccgac tga
```

SEQ ID NO:99

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121 acctgcctgc aggagttcct cagtgaagga gtcttcaagt gccctgagga ccagcttcct
181 ctggactatg ccaagatcta cccagacccg gagctggaag tacaagtatt gggcctgcct
241 atccgctgca tccacagtga ggagggctgc cgctggagtg ggccactacg tcatctacag
301 ggccacctga atacctgcag cttcaatgtc attccctgcc ctaatcgctg ccccatgaag
361 ctgagccgcc gtgatctacc tgcacacttg cagcatgact gccccaagcg gcgcctcaag
421 tgcgagtttt gtggctgtga cttcagtggg gaggcctatg aggtggatga gagttctctg
481 ggctttggtt atcccaagtt catctccac caggacattc gaaagcgaaa ctatgtgcgg
541 gatgatgcag tcttcatccg tgctgctgtt gaactgccc ggaagatcct cagctga

SEQ ID NO:104

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Figure 10 (cont.)

```
1141 tcagccgcga agagatttat cagcttaact cagataaaat cattgaaagt aataaggtaa 1201 aagctagtct ctaacttcca ggcccacggc tcaagtgaat ttgaatactg catttacagt 1261 gtagagtaac acataacatt gtatgcatgg aaacatggag gaacagtatt acagtgtcct 1321 accactctaa tcaagaaaag aattacagac tctgattcta cagtgatgat tgaattcta 1381 aaatggttat cattagggct tttgattat aaaactttgg gtacttatac taaattatgg 1441 tagttattct gccttccagt ttgcttgata tatttgttgg tatttaagatc cttgattcta 1501 attttgaatg ggttctagtg aaaaaggaat ggatatattct 1561 tatttacact cttgattcta caatgtagaa aatgaggaaa 1621 taaaagtcac gtgaaacaga ggattggt gcatccagc cttttgtctt ggtgttcatg 1681 atctccctc aagcacattc caaactttag gatactcaga 1741 gaaaagtttc acctgattg aatcagaatg ccttcaactg accaggitat cacactttgt aatttgcaaa 1741 gaaaagttc acctgattg aatcagaatg ccttcaactg acaacattc 1801 atgaggaaat gtgttggct actacggag cctgtttccc cgtgggtctc tggggtgtca gctttccttt 1921 ctccatgtgt ttgatttctc ctcaggctgg tagcaagttc tggatcttat accacacaca 1981 cagcaacatc cagaaataaa gatct
```

```
1 cggaggcgct gggcgcacgg cgcggagccg gccggagctc gaggccggcg gcggcgggag
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 121 aaagagattc tcagcgctga ttttgagatg atgggcttgg gaaacgggcg tcgcagcatg
 181 aagtegeege ceetegtget ggeegeeetg gtggeetgea teategtett gggetteaac
 241 tactggattg cgageteccg gagegtggae etccagacae ggateatgga getggaagge
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 361 ggagagetgg agaageageg ggageagett gacaaaatce agtecageca caacttecag
 421 ctggagageg teaacaaget gtaccaggae gaaaaggegg ttttggtgaa taacateace
 481 acaggtgaga ggctcatccg agtgctgcaa gaccagttaa agaccctgca gaggaattac
 541 ggcaggetge ageaggatgt cetecagttt cagaagaace agaceaacet ggagaggaag
 601 ttctcctacg acctgagcca gtgcatcaat cagatgaagg aggtgaagga acagtgtgag
 661 gagcgaatag aagaggtcac caaaaagggg aatgaagctg tagcttccag agacctgagt
 721 gaaaacaacg accagagaca gcagctccaa gccctcagtg agcctcagcc caggctgcag
 781 gcagcaggcc tgccacacac agaggtgcca caagggaagg gaaacgtgct tggtaacagc
 841 aagtcccaga caccagcccc cagttccgaa gtggttttgg attcaaagag acaagttgag
 901 aaagaggaaa ccaatgagat ccaggtggtg aatgaggagc ctcagaggga caggctgccg
 961 caggagccag gccgggagca ggtggtggaa gacagacctg taggtggaag aggcttcggg
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1141 gagcaggaag ctgccgggga agggagaaac cagcagaaac tgagaggaga agatgactac
1201 aacatggatg aaaatgaagc agaatctgag acagacaagc aagcagccct ggcagggaat
1261 gacagaaaca tagatgtttt taatgttgaa gatcagaaaa gagacaccat aaatttactt
1321 gatcagcgtg aaaagcggaa tcatacactc tgaattgaac tggaatcaca tatttcacaa
1381 cagggccgaa gagatgacta taaaatgttc atgagggact gaatactgaa aactgtgaaa
1441 tgtactaaat aaaatgtaca tctgaagatg attattgtga aattttagta tgcactttgt
1501 gtaggaaaaa atggaatggt cttttaaaca gcttttgggg gggtactttg gaagtgtcta
1561 ataaggtgtc acaatttttg gtagtaggta tttcgtgaga agttcaacac caaaactgga
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1861 aatcaatggc ctagagcact gactgttaac acaaacgtca ctagcaaagt agcaacagct
1921 ttaagtctaa atacaaagct gttctgtgtg agaatttttt aaaaggctac ttgtataata
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2041 gtctttattt actttgcttc gtgtgtgggc aaagcaacat cttccctaaa tatatattac
2101 caagaaaagc aagaagcaga ttaggttttt gacaaaacaa acaggccaaa agggggctga
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```

Figure 10 (cont.)

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 541 caggaaatcc acgagaaggt tctaaacgaa gccgtgggcg ccatgatgta ccaccatc
 601 acceteacea gggaggacet ggagaagtte aaggeeetga gagtgategt geggatagge
 661 agtggctatg acaacgtgga catcaaggct gccggcgagc tcggaattgc cgtgtgcaac
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 841 gagcagatcc gcgaggtggc ctcgggagcg gcccgcatcc gtggggagac gctgggcctc
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1381 acaggtogca toccagaaag ottaagaaat tgtgtgaaca aggaattott tgtcacatca
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1561 atcatecetg gaggeatece agtgacteae aaceteeega cagtggeaca teetteecaa
1621 gegecetete ecaaceagee cacaaaacae ggggacaate gagageacee caacgageaa
1681 tagcagagaa tgccagaagg taatcactca gatacacttg ggaccaagag acagtgaaaa
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1861 cggaagcgct gaaagactag gatgtgattt attaacgacc aacttctgtt attgtgtgtt
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1981 tecettgggc acagcaggte etgaacacce tgetetacaa tgttgcatca agagttcaaa
2041 Caacaaaata aaaaatatta agaggaaatc cccatcctgt gacttgagtc ccttaagtct
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```

Figure 10 (cont.)

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2281 cagtaaaacc ctctgatgat gcaaaaaaaa aaaaaaagta ttaagtttca caagctgttt
2341 gtactcaaat atattttctc agtttcag
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SEQ ID NO:116

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 121 tgggtctgtc aaagcctata ctaactttga tgctgagcgg gatgctttga acattgaaac
 181 agccatcaag accaaaggtg tggatgaggt caccattgtc aacattttga ccaaccgcag
 241 caatgcacag agacaggata ttgccttcgc ctaccagaga aggaccaaaa aggaacttgc
 301 atcagcactg aagtcagcct tatctggcca cctggagacg gtgattttgg gcctattgaa
 361 gacacctgct cagtatgacg cttctgagct aaaagcttcc atgaaggggc tgggaaccga
 421 cgaggactet eteattgaga teatetgete cagaaccaac caggagetge aggaaattaa
 481 cagagtetac aaggaaatgt acaagactga tetggagaag gacattattt eggacacate
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1321 gtactgtgtc ataaacagat gaataaactg aatttgtact tt
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 661 acacategtt catgttecag egagtgetgg tgtetetgte agetggtggg agggatgaag
 721 gaaattatot ggacgatgot otogtgagac aggatgooca ggacotgtat gaggotggag
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 961 tgaggaacaa atctgcatat tttgctgaaa agctctataa atcgatgaag ggcttgggca
1021 ccgatgataa caccctcatc agagtgatgg tttctcgagc agaaattgac atgttggata
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Figure 10 (cont.)

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1321 agaatataga etgtetgtat tattatteae etataattag teattatgat getttaaage
1441 gtattccatg tttttaaaag attacttct actttgtgtt tcacagacat tgaatatatt
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2101 aagatgtact tggatttaat taaaaagttc actttgt
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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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Figure 10 (cont.)

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4561 gagaaaattg tttcgaaagc agtcacaagc aagaaatcca agggggagag tgatgacttc
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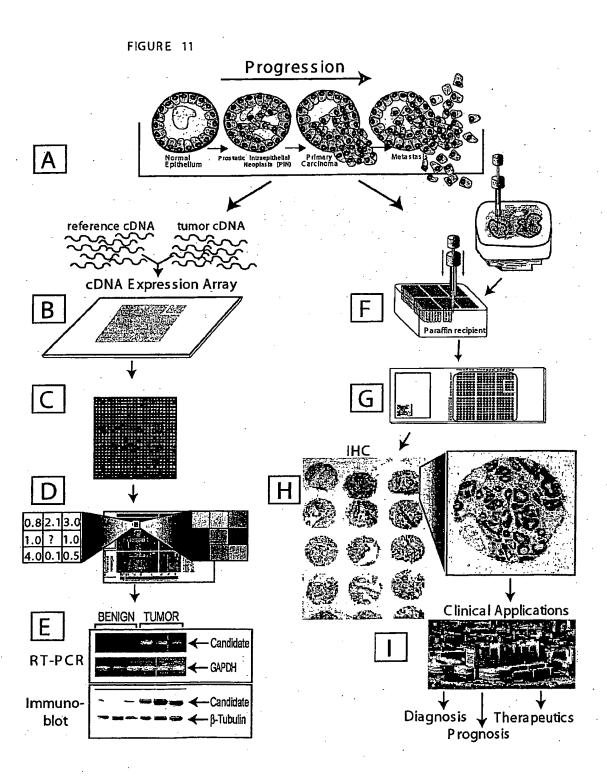
Figure 10 (cont.)

4681 ataaagtacc tggaagagtc agatgaagat gatctgtttt aaaatgtgag gcgattattt 4741 taagtaatta tcttaccaag cccaagactg gttttaaagt tacctgaagc tcttaacttc 4801 ctcccctctg aatttagttt ggggaaggtg tttttagtac aagacatcaa agtgaagtaa

```
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     5101 ctccttttct actttcagta gatatgagat agagcataat tatctgtttt atcttagttt
     5161 tatacataat ttaccatcag atagaacttt atggttctag tacagatact ctactacact
     5221 cagoctotta tgtgccaagt ttttctttaa gcaatgagaa attgctcatg ttcttcatct
     5281 teteaaatea teagaggeea aagaaaaaca etttggetgt gtetataaet tgacacagte
     5341 aatagaatga agaaaattag agtagttatg tgattatttc agctcttgac ctgtcccctc
     5401 tggctgcctc tgagtctgaa tctcccaaag agagaaacca atttctaaga ggactggatt
     5461 gcagaagact cggggacaac atttgatcca agatcttaaa tgttatattg ataaccatgc
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SEQ ID NO:122
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      181 acatcaactt ceggeegeag atgteceagt ttetgtgttt gaagaacata egeacettee
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481 acgactgcgt cccgtgtgag gatggagggg acgacatcta cgaggacatc atcaaggtgg
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      601 ggaactgctg cctgctggag atccaggaga ccgaggccaa gtactaccgc accctggagg
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     1981 gccggccgcc atcccgggag atcgactaca ctgcataccc ctggtttgca ggtaacatgg
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```

Figure 10 (cont.)

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2161 acatcaaggt					
2221 gcctcctgga					
2281 tggacaccac	actcaagtac	ccctacaagt	cccgggaacg	ttcggcctcc	agggcctcca
2341 gccggtcccc	agcttcctgt	gcttcctaca	acttttcttt	tctcagtcct	cagggcctca
2401 gctttgcttc	tcagggcccc	tccgctccct	tctggtcagt	gttcacgccc	cgcgtcatcg
2461 gcacagctgt	ggccaggtat	aactttgccg	cccgagatat	gagggagctt	tegetgeggg
2521 agggtgacgt	ggtgaggatc	tacagccgca	tcggcggaga	ccagggctgg	tggaagggcg
2581 agaccaacgg	acggattggc	tggtttcctt	caacgtacgt	agaagaggag	ggcatccagt
2641 gacggcagga	acgtggacaa	gactcgcaga	ttttcttggg	agagtcactc	cagccctgaa
2701 gtctgtctct	agctcctctg	tgactcagag	gggaaatacc	aacctcccag	tet



. FIGURE 12

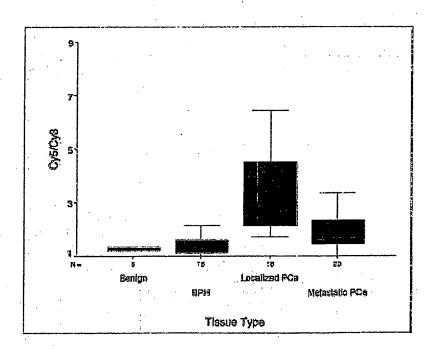


FIGURE 13

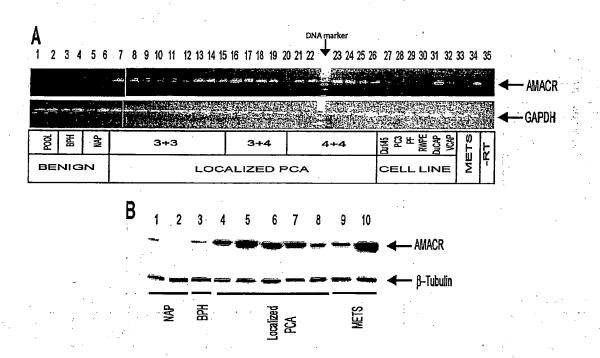
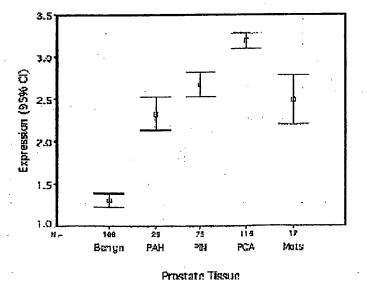


FIGURE 14



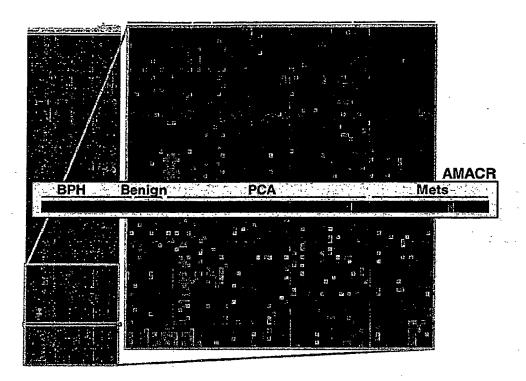
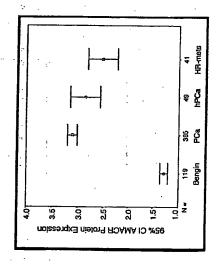


Figure 15



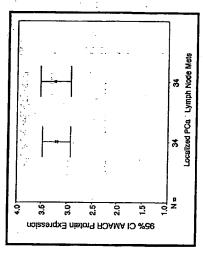
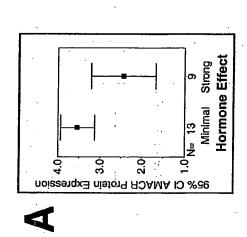
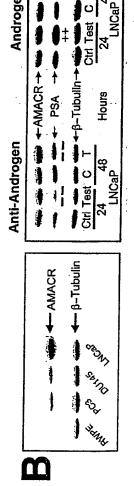


Figure 16

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igure 17

Figure 18

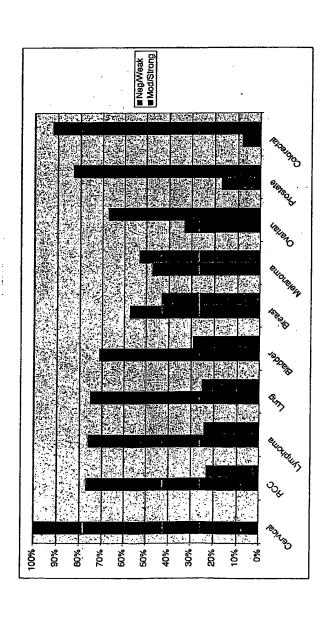


Figure 19

AMACR-

DNA marker
Benign
PCA
Benign
PCA

Figure 20

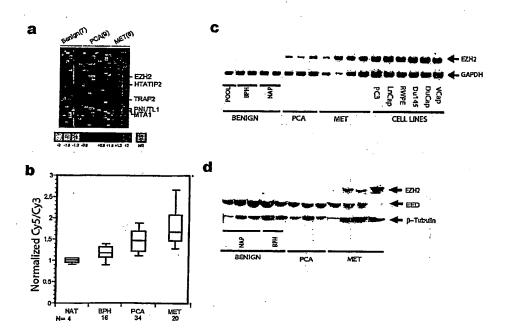


Figure 21

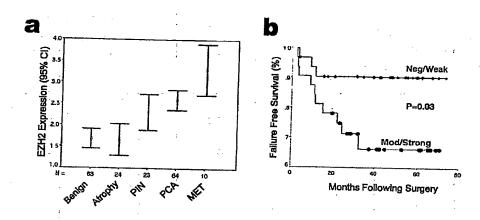
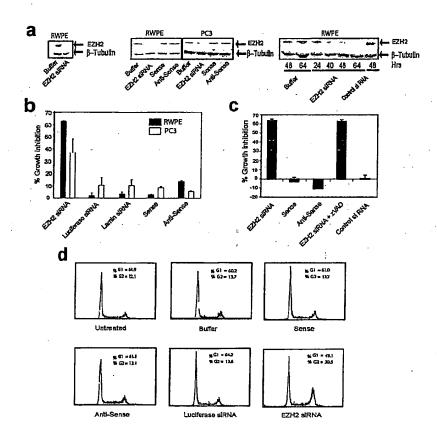
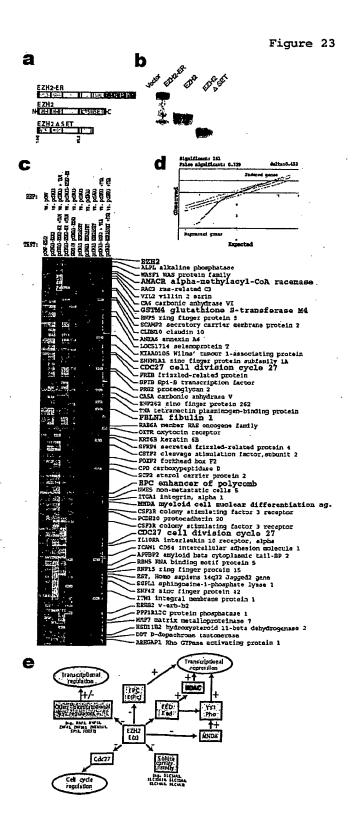
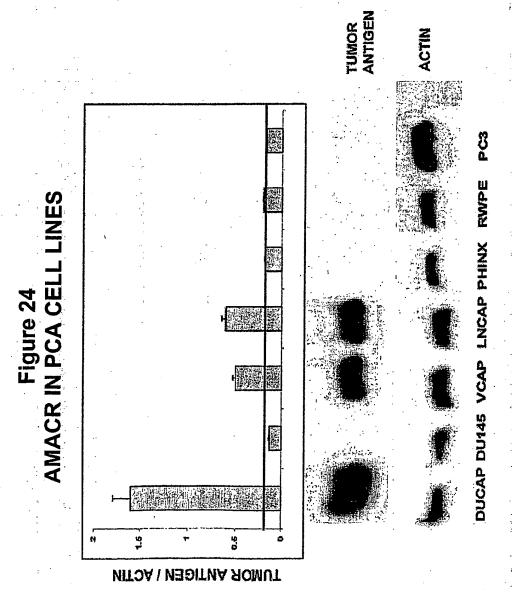


Figure 22







116/129

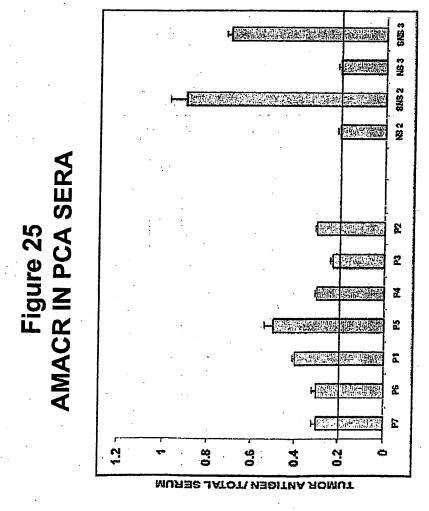


Figure 26 Immunoblot Analysis for AMACR in Prostate Cancer and Normal Sera



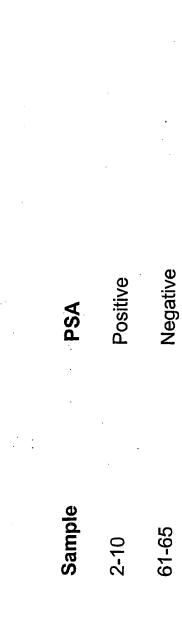
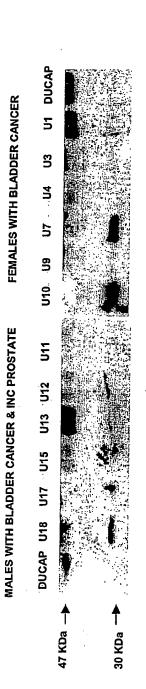
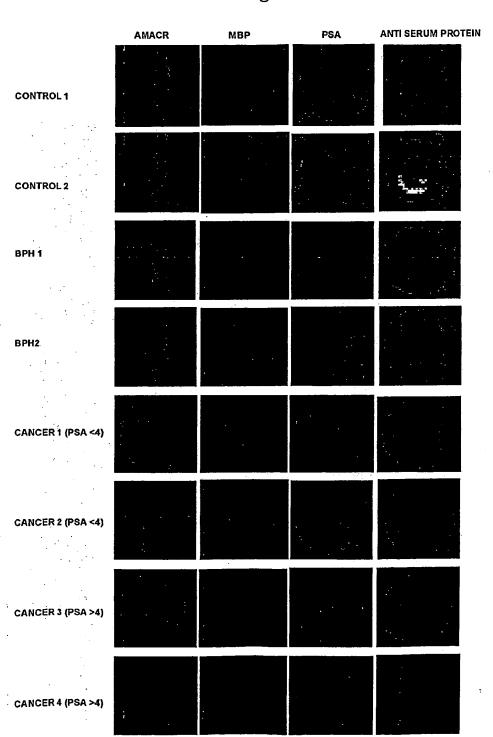


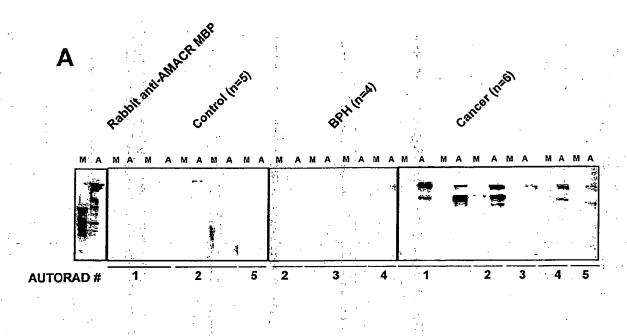
Figure 27
IMMUNOBLOT ANALYSIS
OF URINE SAMPLES FOR AMACR



U1-U10 : FEMALES WITH BLADDER CANCER U11-U20 : MALES WITH BLADDER CANCER AND INC PROSTATE

Figure 28



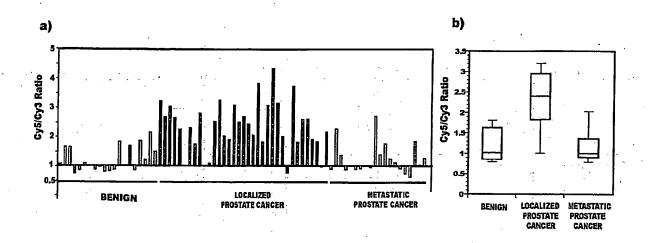


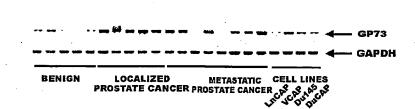
B

Figure 29

C)

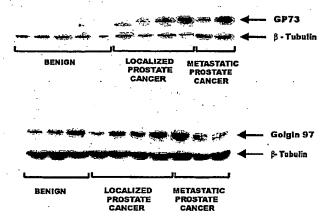
Figure 30





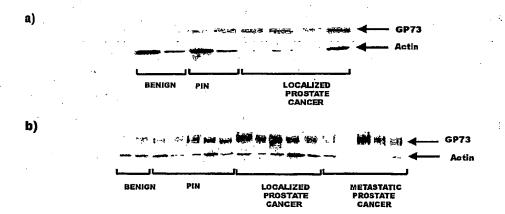
· *: ·

Figure 31



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Figure 32



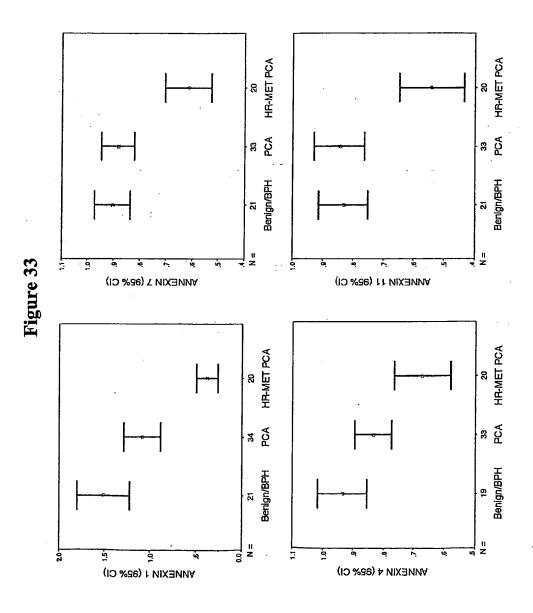




Figure 34

Figure 35

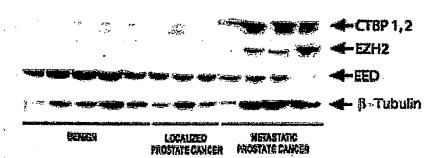


Figure 36

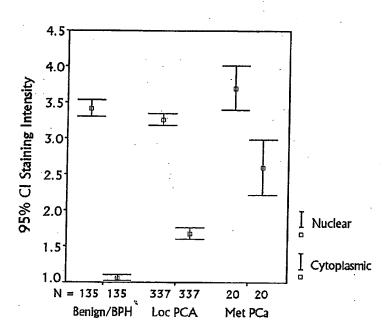


Figure 37

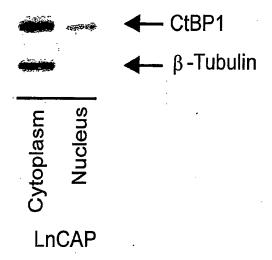
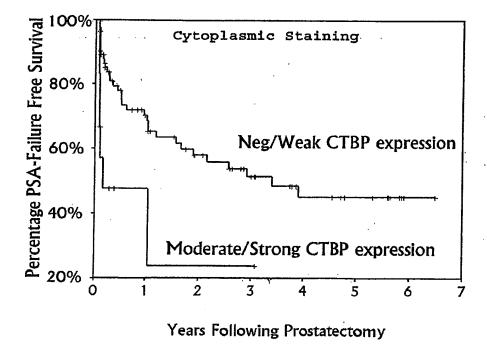


Figure 38



(19) World Intellectual Property Organization

International Bureau





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(10) International Publication Number -WO 2003/012067 A3

(51) International Patent Classification7: G01N 33/53, C07K 16/00, C07H 21/04 C12Q 1/68,

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- (74) Agents: ARENSON, Tanya, A. et al.; Medlen & Carroll, LLP, 101 Howard Street, Suite 350, San Francisco, CA 94105 (US).
- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

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- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments
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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: EXPRESSION PROFILE OF PROSTATE CANCER

(57) Abstract: The present invention relates to compositions and methods for cancer diagnostics, including but not limited to, cancer markers. In particular, the present invention provides gene expression profiles associated with prostate cancers. Genes identified as cancer markers using the methods of the present invention find use in the diagnosis and characterization of prostate cancer. In addition, the genes provide targets for cancer drug screens and therapeutic applications.



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/2456

			•	10170302224007		
A. (IPC(7)	SSIFICATION OF SUBJECT MATTER : C12Q 1/68; G01N 33/53; C07K 16/00; C07I : 435/6, 7.1; 530/387.1; 536/24.3	H 21/04			
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B. F	TEL	DS SEARCHED	<u> </u>			
		cumentation searched (classification system follows 35/6, 7.1; 530/387.1; 536/24.3	d by classi	fication symbols)		
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Docume	ntati	on searched other than minimum documentation to	he extent t	hat such documents are include	d in the fields searched	
			· ·	·		
Electron	ic de	ata base consulted during the international search (n	ame of date	a base and, where practicable, s	earch terms used).	
				•	•	
C. D	XOC	UMENTS CONSIDERED TO BE RELEVANT				
Category	y *	Citation of document, with indication, where	appropriate	, of the relevant passages	Relevant to claim No.	
Х		US 6,518,028 B1 (O'BRIEN) 11 February 2003 (1-3,6-7,9-10	
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<u> </u>		documents are listed in the continuation of Box C.	<u> </u>	See patent family annex.		
	•	ecial categories of cited documents:	-1-	later document published after the inter date and not in conflict with the applica-	tion but cited to understand the	
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"E" carlie		dication or patent published on or after the international filing date	•X•	document of particular relevance; the c considered novel or cannot be considered when the document is taken alone	laimed invention cannot be ed to involve an inventive step	
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"O" docu	ment i	referring to an oral disclosure, use, exhibition or other means		combined with one or more other such being obvious to a person skilled in the		
		published prior to the international filing date but later than the	·&•	document member of the same patent fa		
Date of th	he ac	tual completion of the international search	Date of n	nailing of the international sear	ch report	
20 Septen	nber	2004 (20.09.2004)	1	7 NOV 2004.		
		iling address of the ISA/US	Authoriz	ed officer / 20/1-1/	allen	
Mail Stop PCT, Attn: ISA/US Commissioner for Patents				Lindson Savis Del Harris for		
	P.O.	Box 1450	Tolonbon			
Alexandria, Virginia 22313-1450 Facsimile No. (703) 305-3230				e No. 571-272-1600		

International	enalization	N۵
THICH TRACTORES	application:	NO.

INTERNATIONAL SEARCH REPORT

PCT/US02/24567

This interr	national report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
_	
1.	Claim Nos.:
	because they relate to subject matter not required to be searched by this Authority, namely:
. •	
. —	
2	Claim Nos.:
. —	because they relate to parts of the international application that do not comply with the prescribed requirements to
	such an extent that no meaningful international search can be carried out, specifically:
<u></u>	
3. 🔝	Claim Nos.:
	because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule
6.4(a).
Box II (Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
	more among a man or man
Thic Interr	stional Consulting Authority found multimle inventions in this international application, as follows:
I IIIS IIIIGI II	ational Searching Authority found multiple inventions in this international application, as follows:
ı. 🔲	As all required additional search fees were timely paid by the applicant, this international search report covers all
-	searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite
سببه	payment of any additional fee.
,	
·	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
	report covers only mose claims for which fees were paid, specifically claims ros
∇	ar 1 1 100 and a sector of the
Ь. 🖂	No required additional search fees were timely paid by the applicant. Consequently, this international search report
	is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-13 and 38-41
Remark on	Protest The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees.